

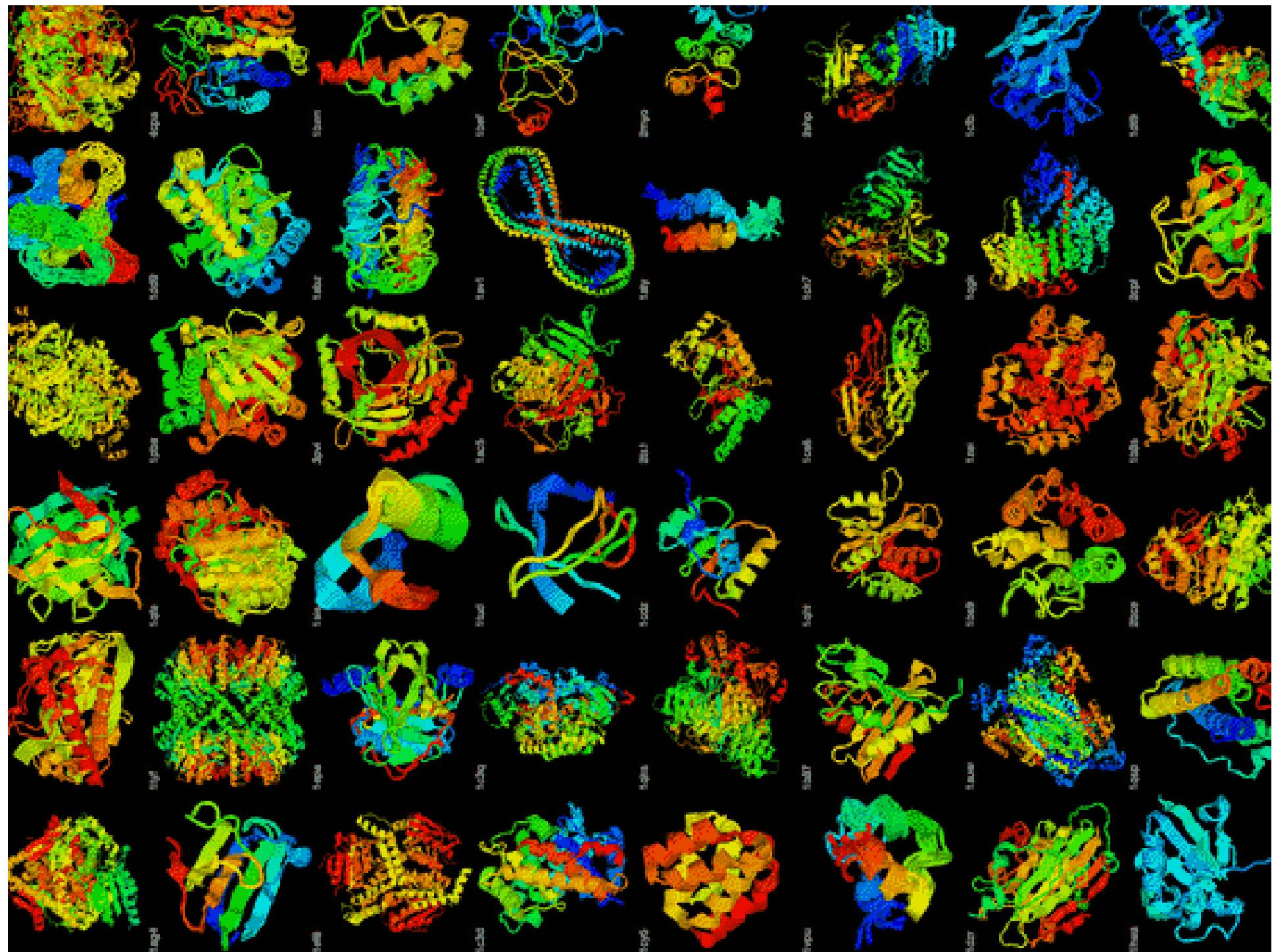
# Time resolved diffraction & scattering studies of membrane protein dynamics using XFEL radiation

Richard Neutze  
University of Gothenburg

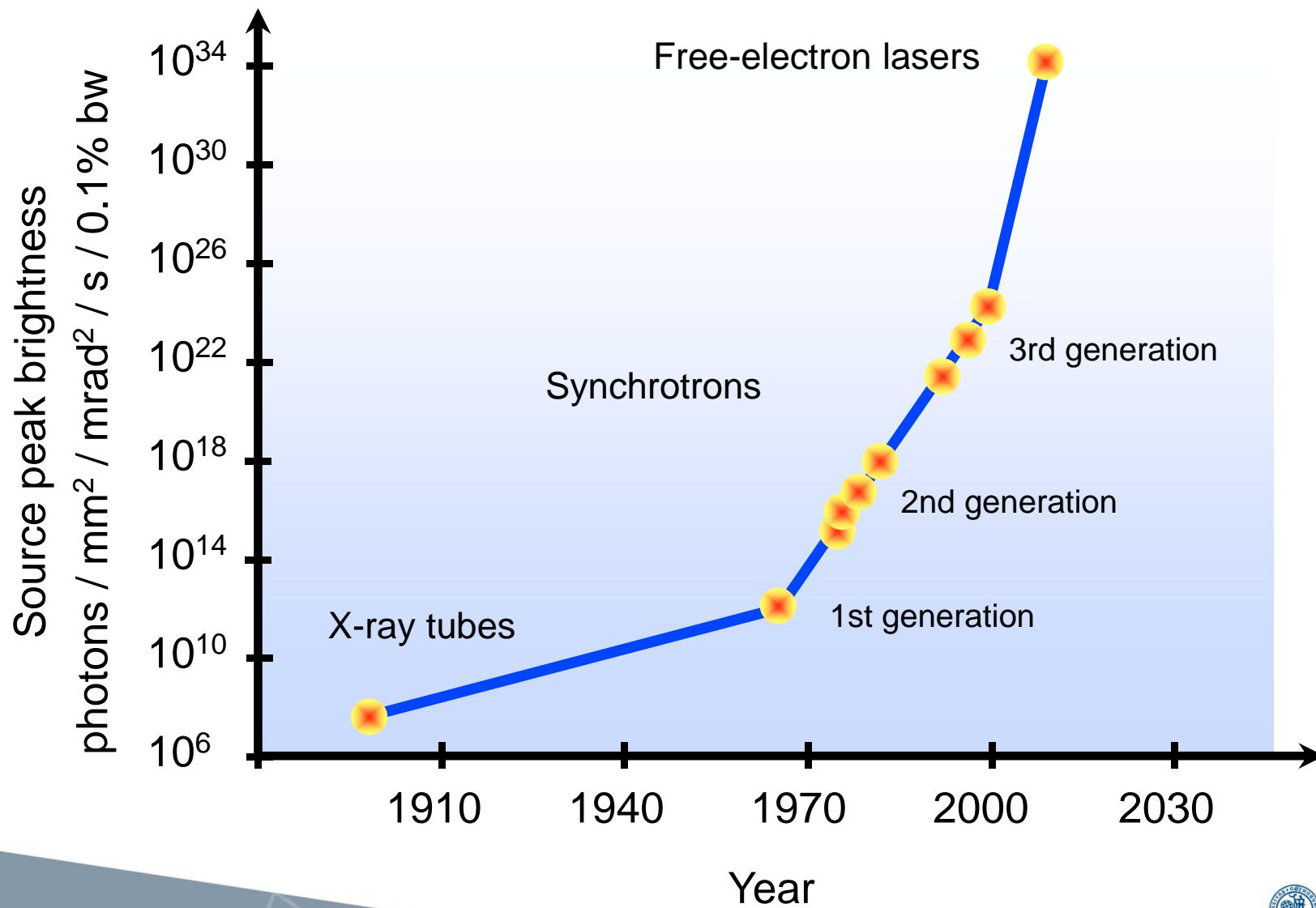


UNIVERSITY OF GOTHENBURG

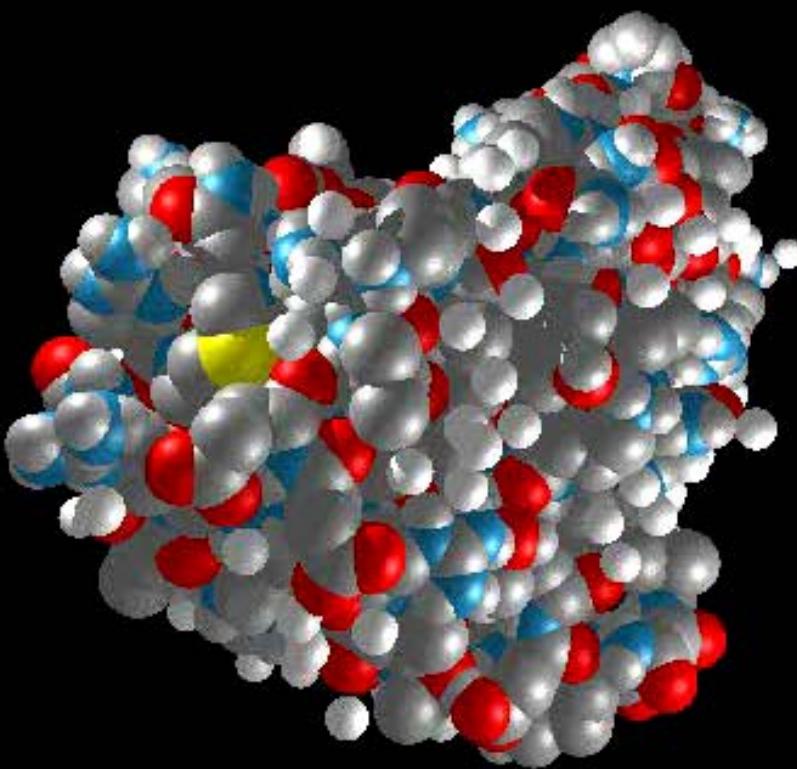




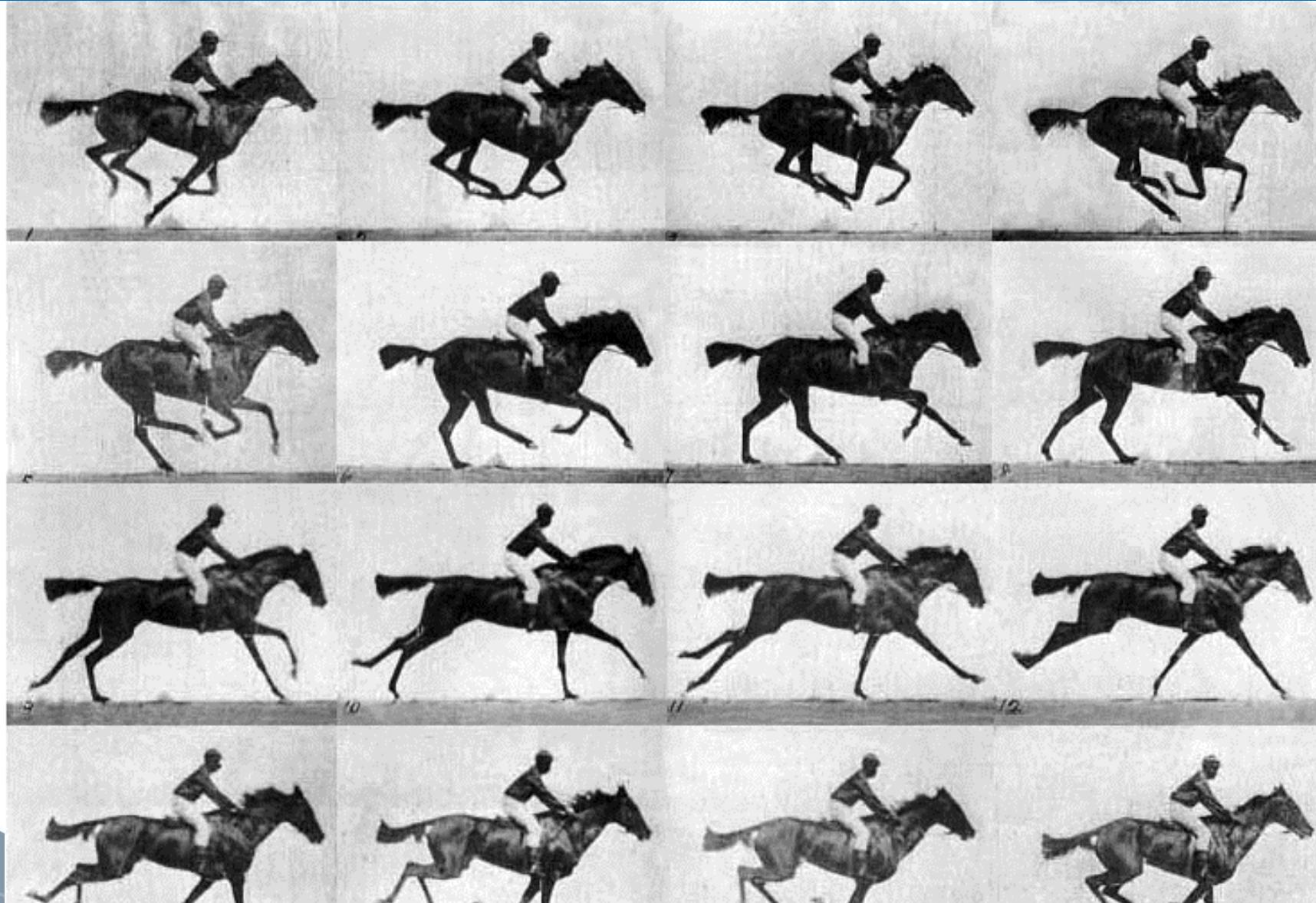
# Peak Brilliance



# Diffraction before Destruction



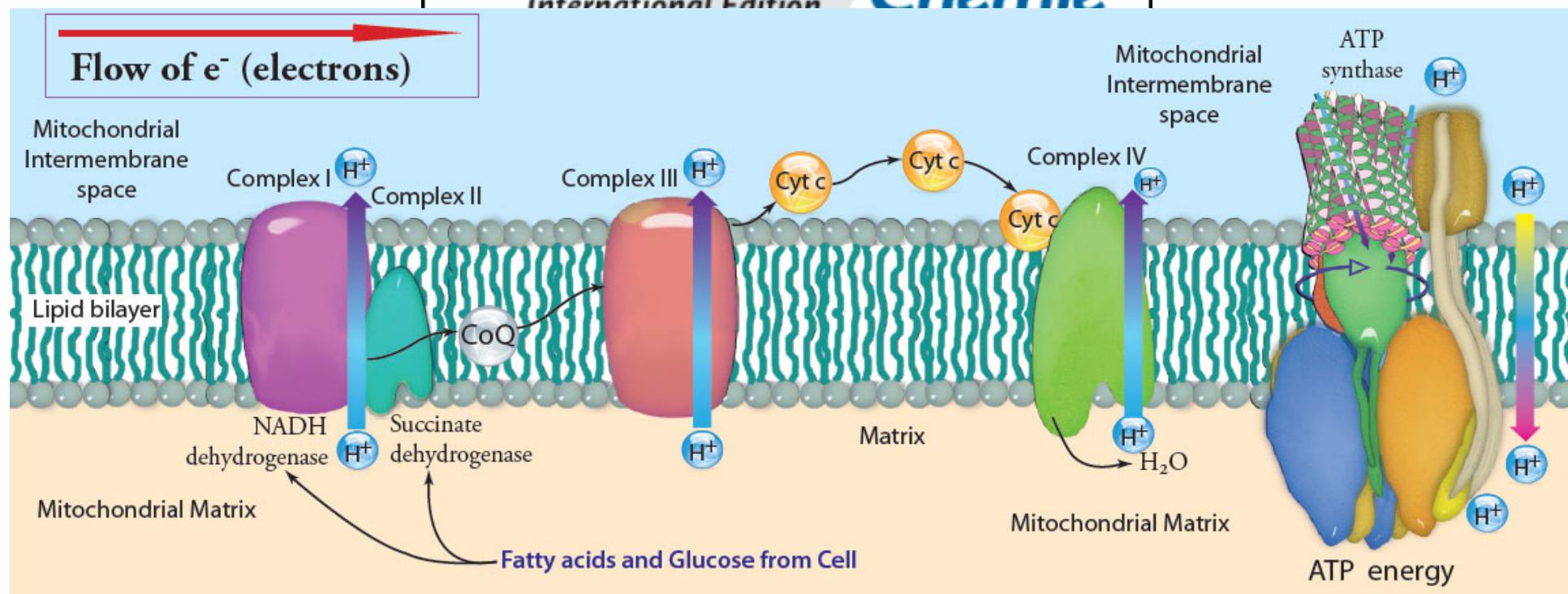
# Reaction Dynamics



Muybridge's freeze-frame sequence revealed a horse's gait.

# Energy Transduction

A journal of the Gesellschaft Deutscher Chemiker  
**Angewandte**  
International Edition



**Fluorination**  
Review by T. Ritter et al.  
**Flexible Mesocrystals of Calcite**  
Highlight by D. Gréber  
**Inhibitors of Cysteine Proteases**  
Highlight by J. Rademann and C. Arkona  
ACIEP 52 (52) 8171–8404 (2013) · ISSN 1433-7851 · Vol. 52 · No. 52

125 **Angewandte**  
Chemie

WILEY-VCH

Faculty of Science



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# Photosynthetic Proteins

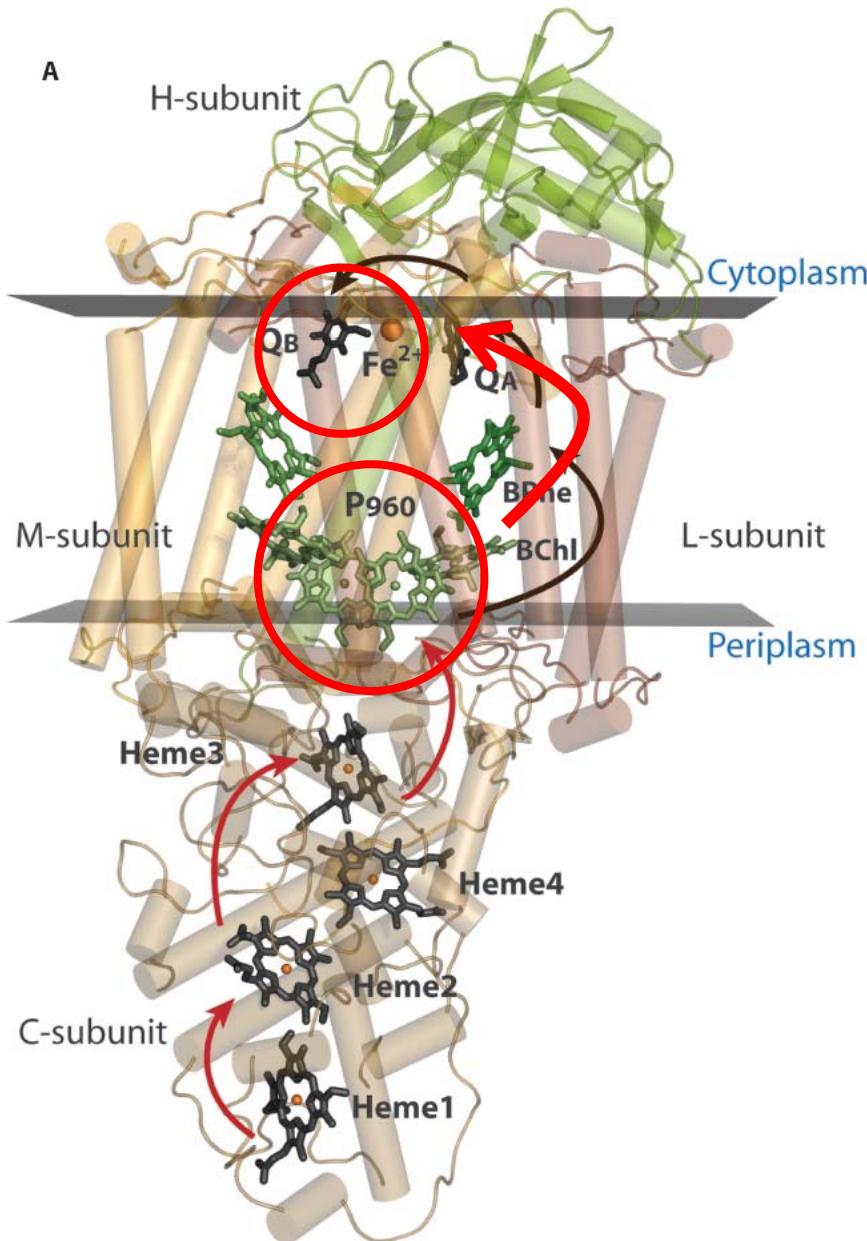


# Photosynthetic Purple Bacteria



Blooming purple sulfur bacteria in a coastal lagoon

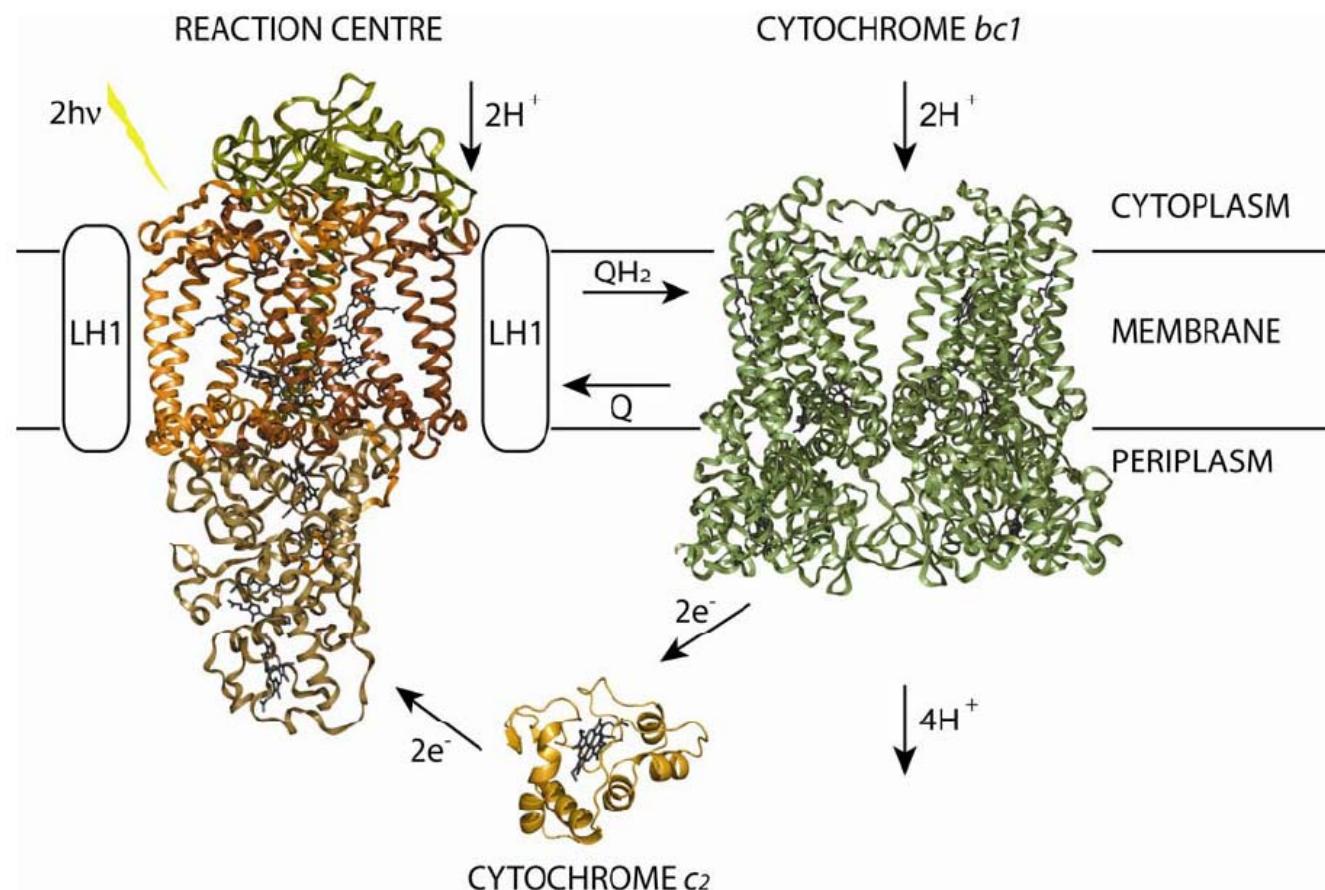
# Photosynthetic reaction centres



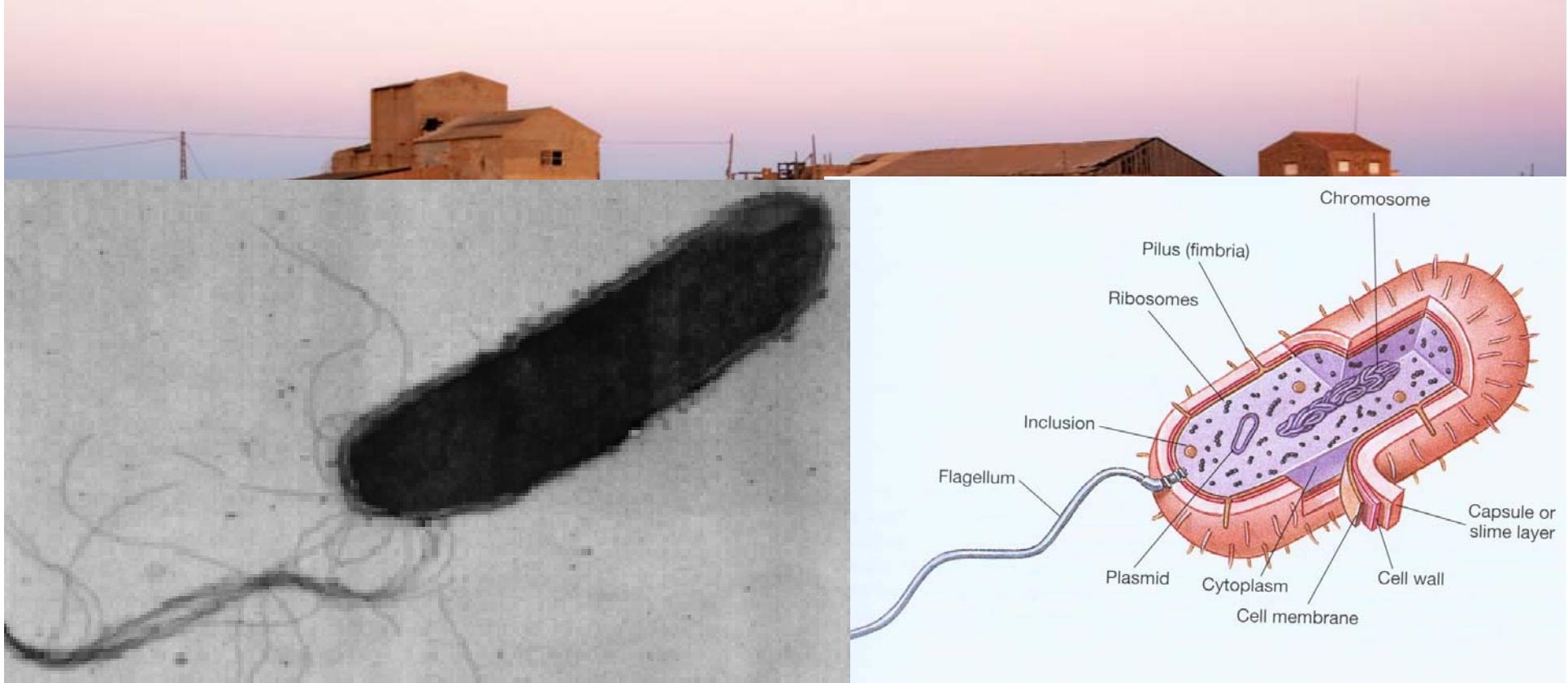
- 135 kDa Membrane protein.
- Complex light driven proton pump.
- Electron movements driven by light.
- Coupled redox reactions pump protons.
- Descendent created O<sub>2</sub> rich atmosphere.

# Proton pumping by reaction centres

- Electron movements driven by light.
- Coupled redox reactions pump protons.

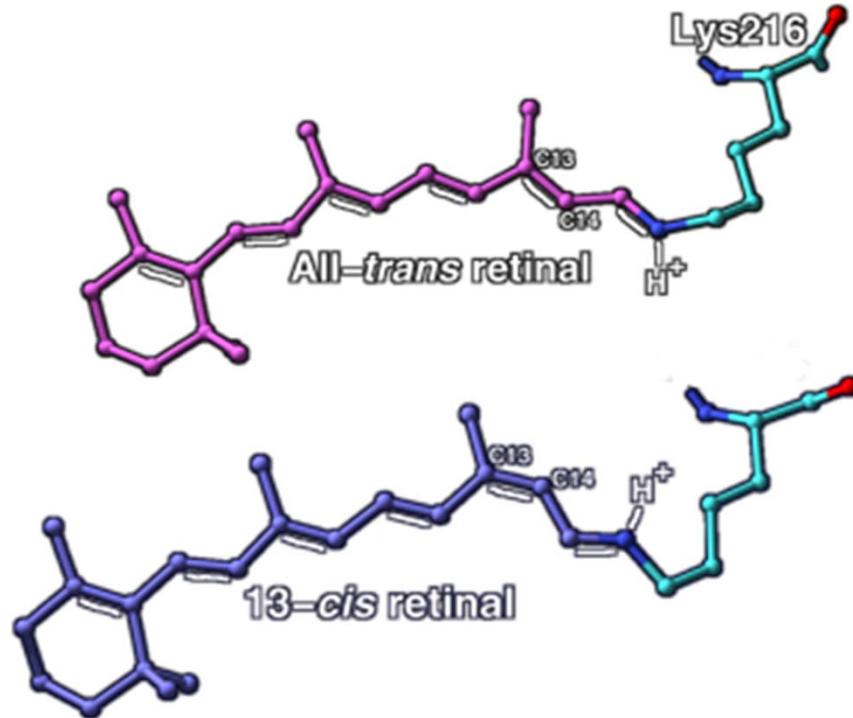


# Halo-archaea

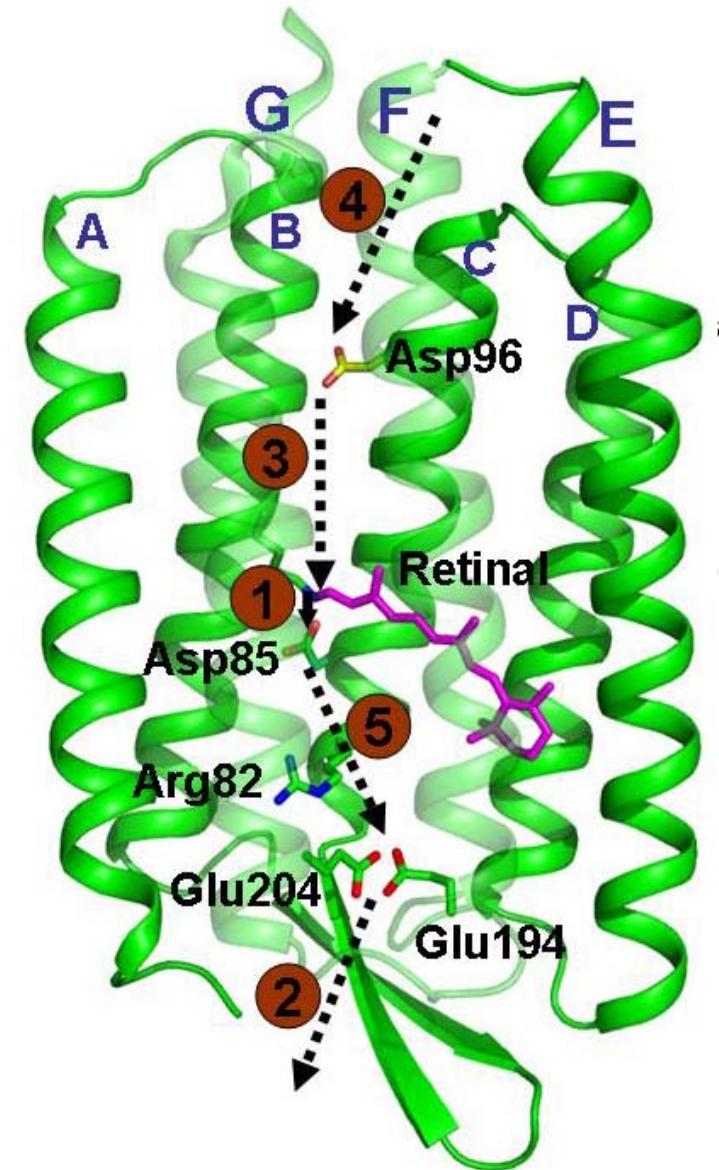


Halobacterium Salinarium in high-salt ponds

# Bacteriorhodopsin



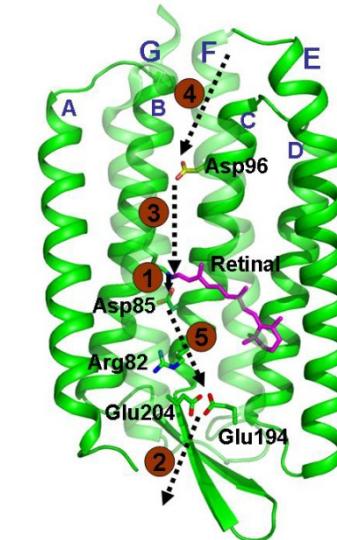
- Retinal isomerisation starts a sequence of structural changes.
- One proton pumped per photon.



# Scientific Question

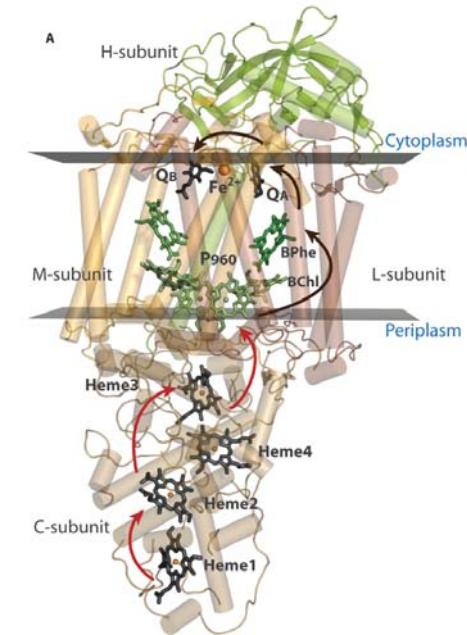
## Bacteriorhodopsin:

- What structural changes are needed to achieve proton pumping up-hill against a proton gradient?

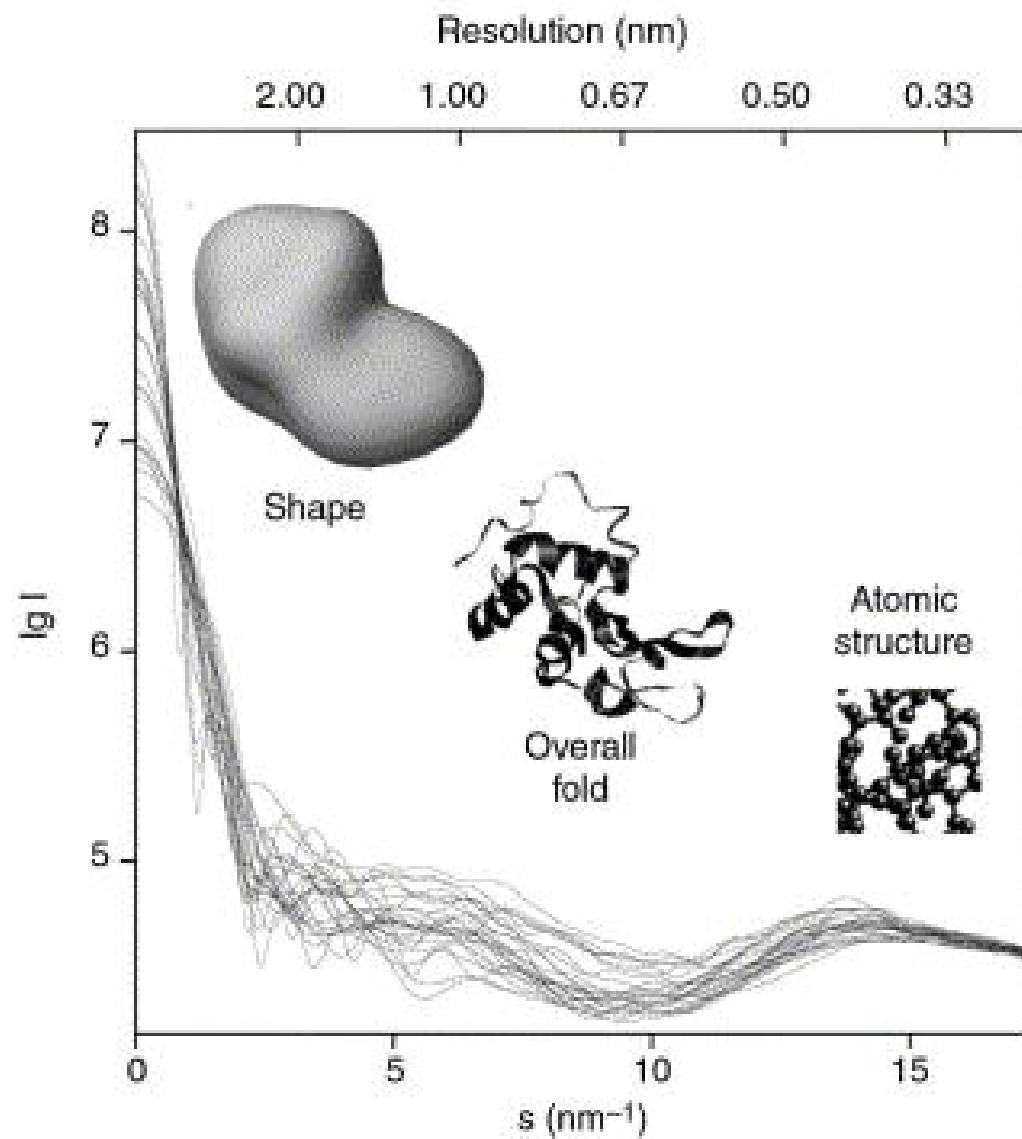


## Photosynthetic reaction centres:

- Do ultrafast conformational changes contribute to the primary charge separation reactions of photosynthesis?

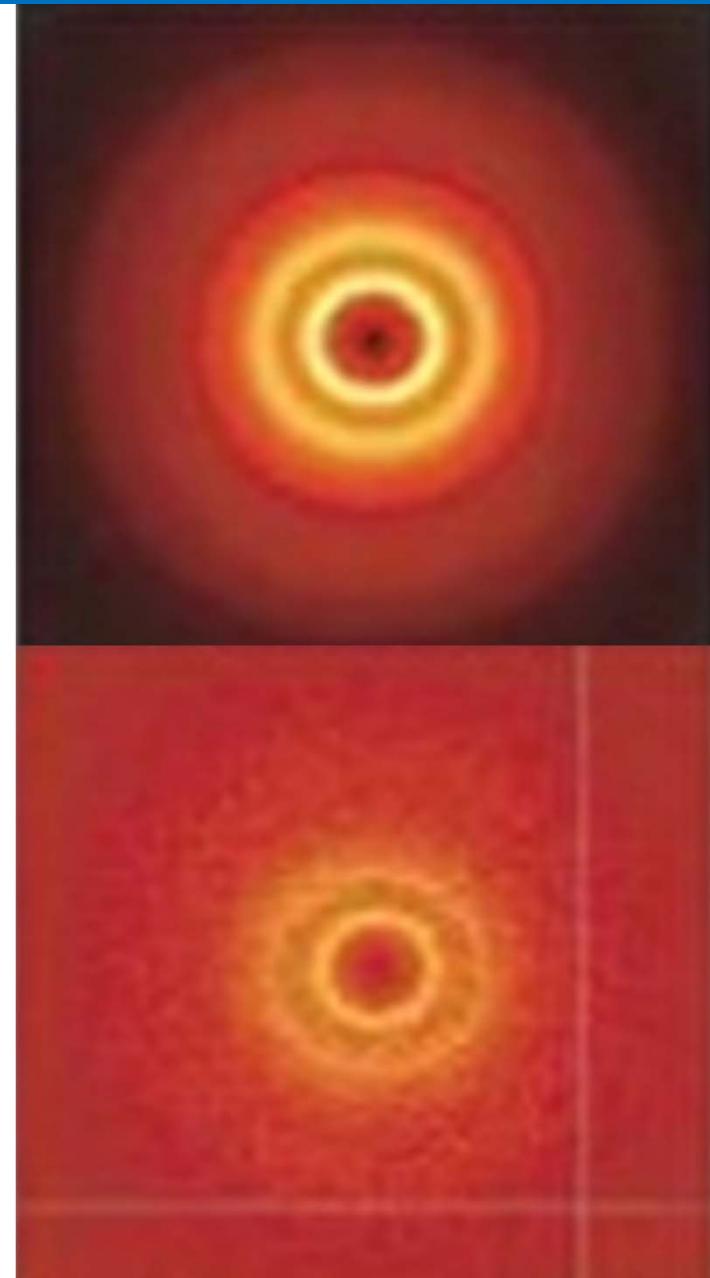
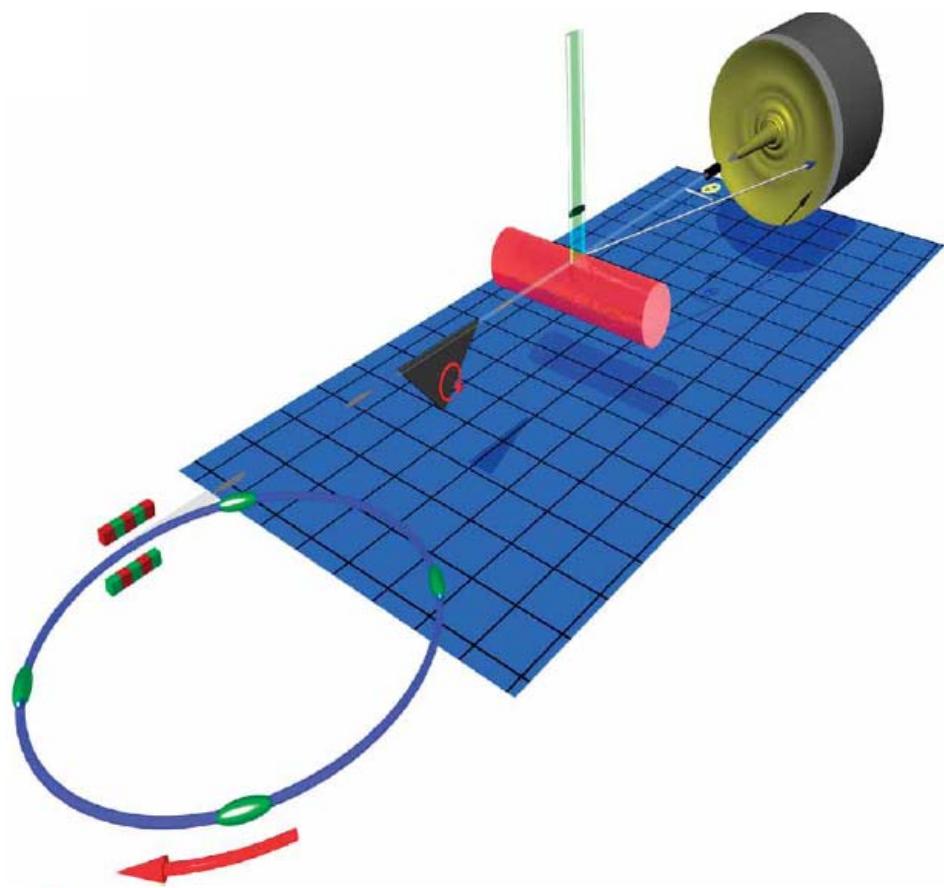


# X-ray scattering



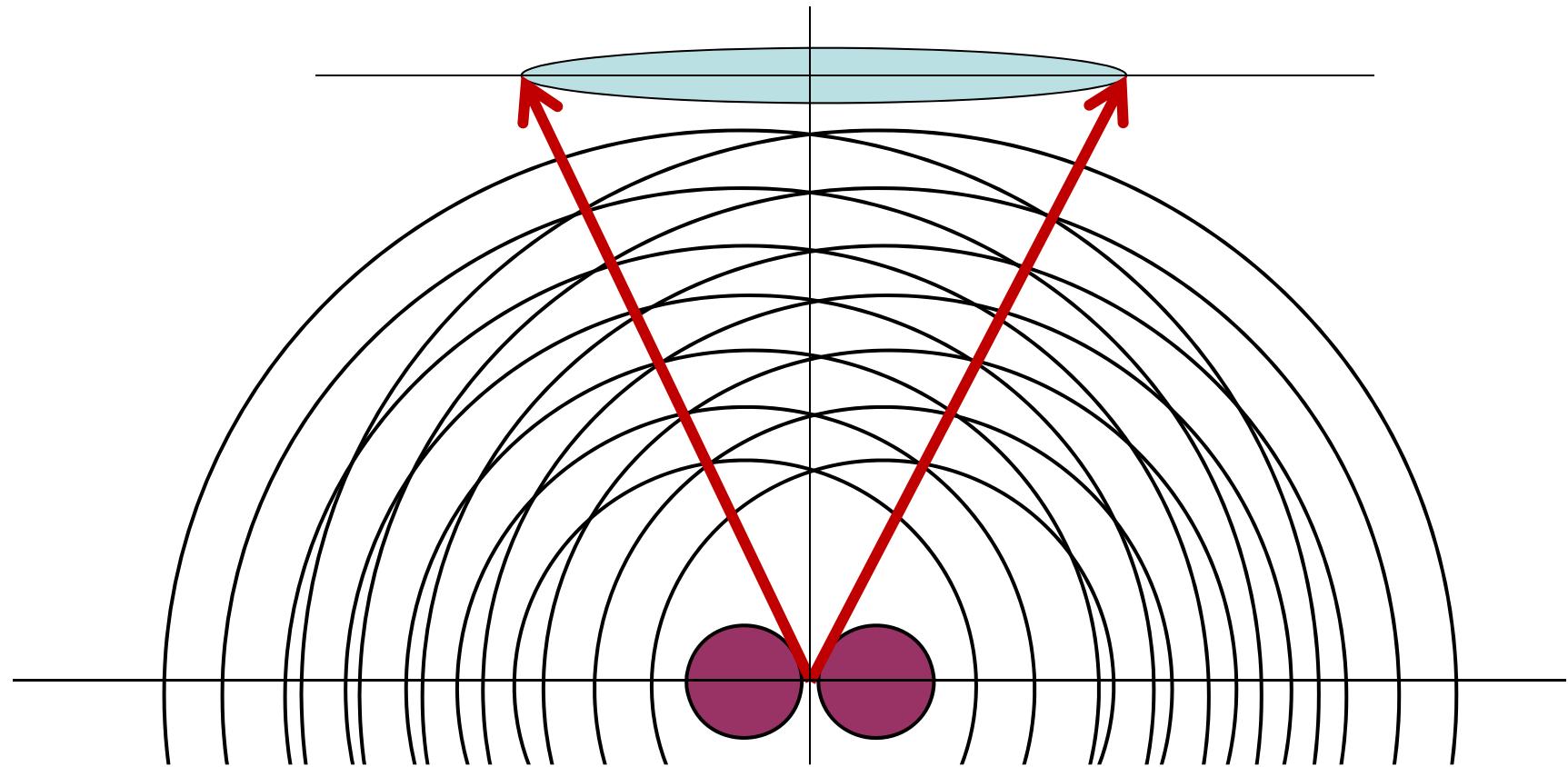
Svergun & Koch, *Biophys. Meth.* (2002)

# Time-resolved Wide Angel X-ray Scattering

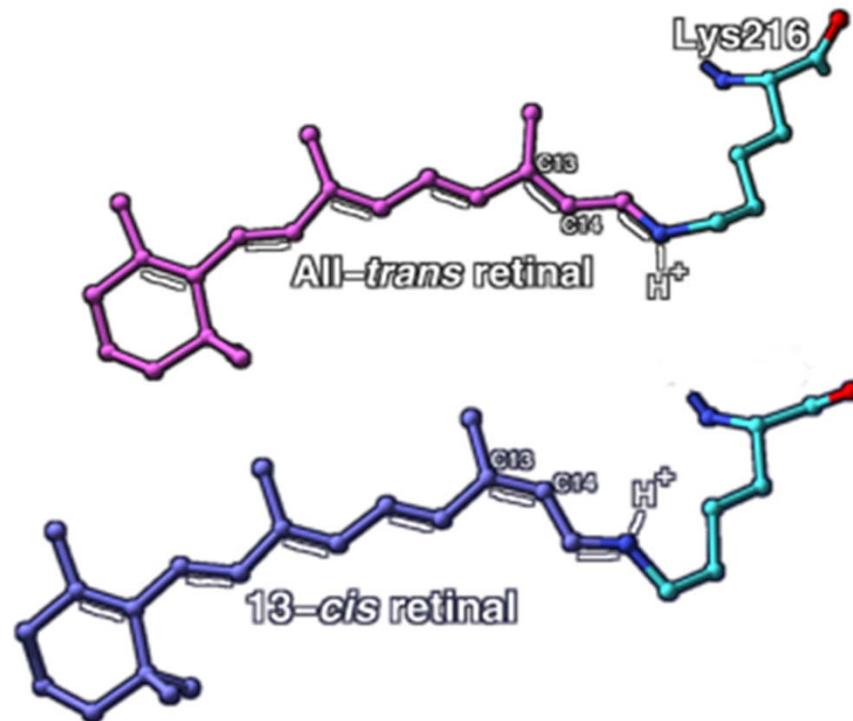


Cammarata et al., Nature Methods (2008)

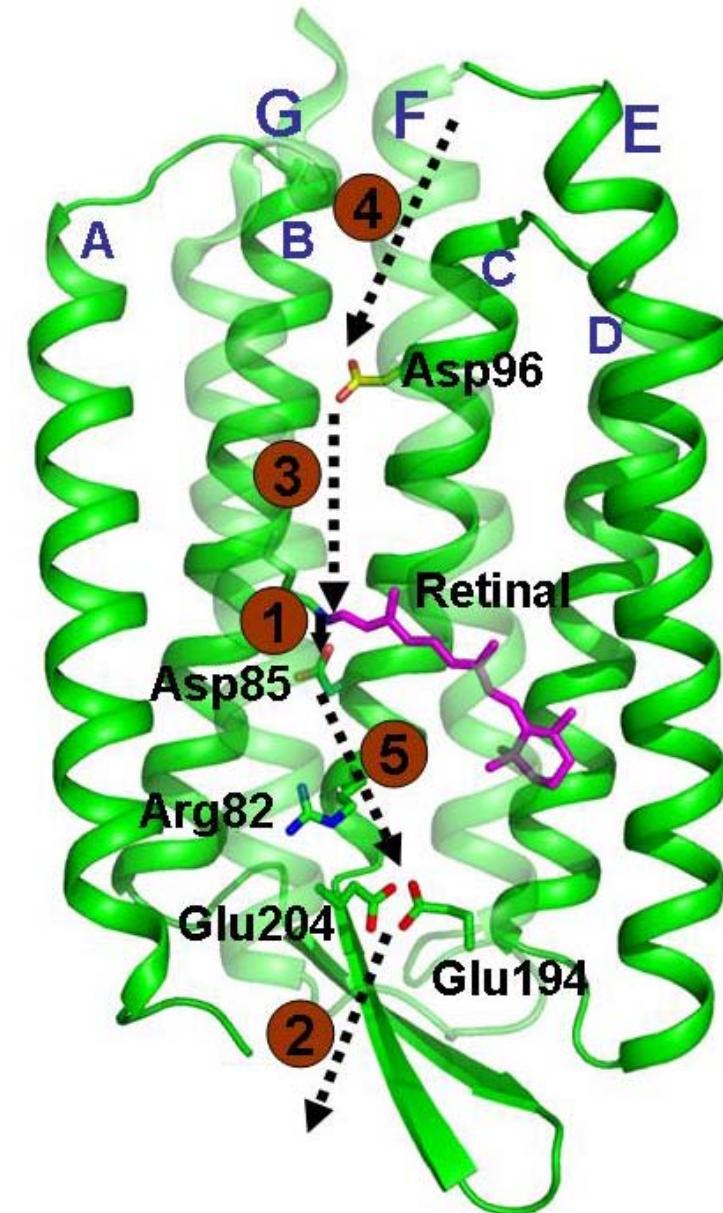
# Time Resolved Wide Angle X-ray Scattering



# Bacteriorhodopsin



- Retinal isomerisation starts a sequence of structural changes.
- One proton pumped per photon.



# European Synchrotron Radiation Facility

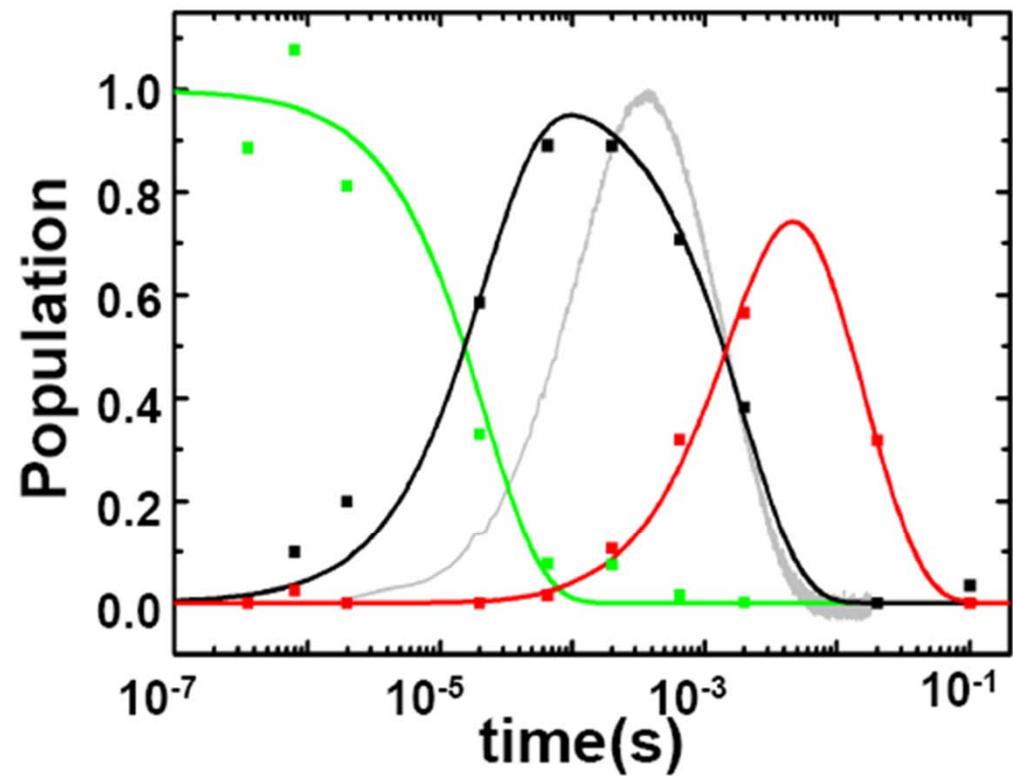
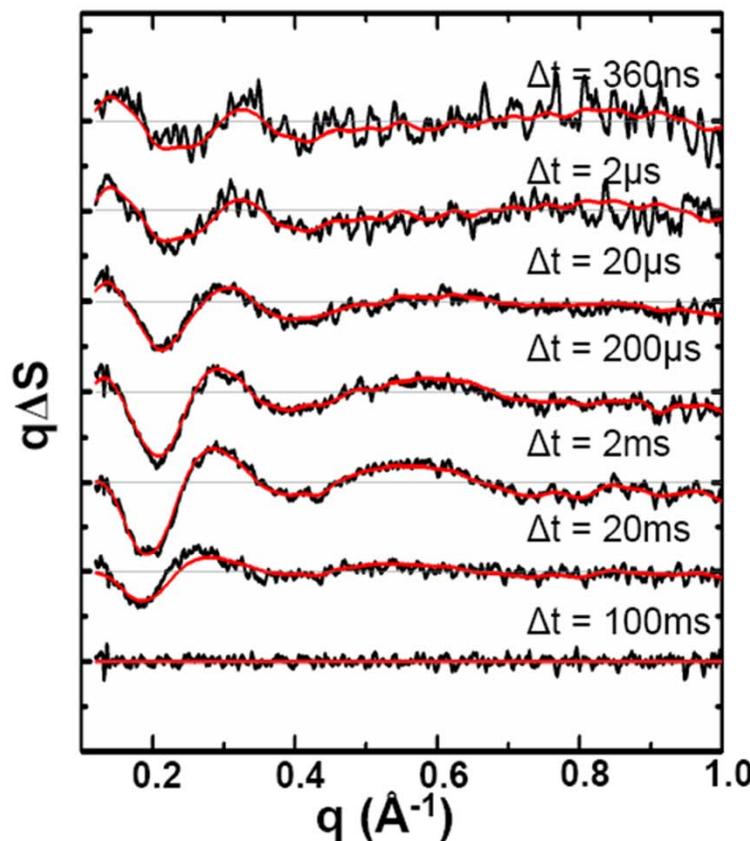


Michael Wulff

Marco Cammarata

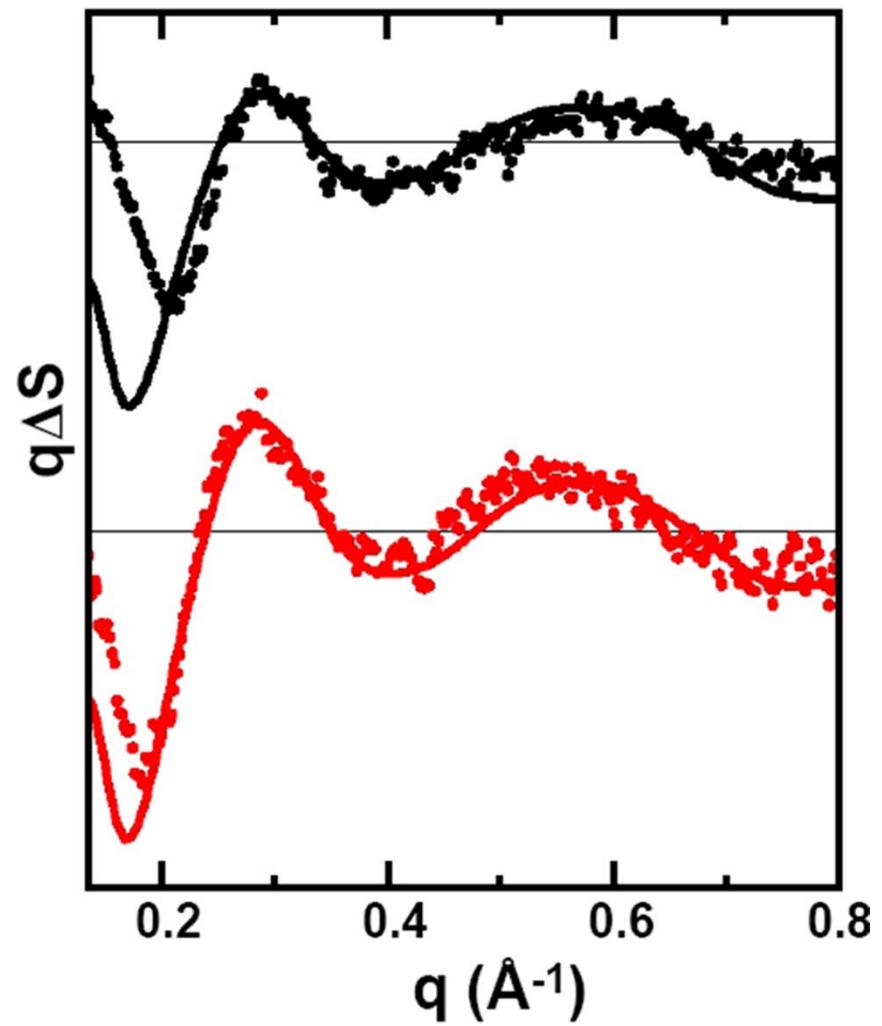
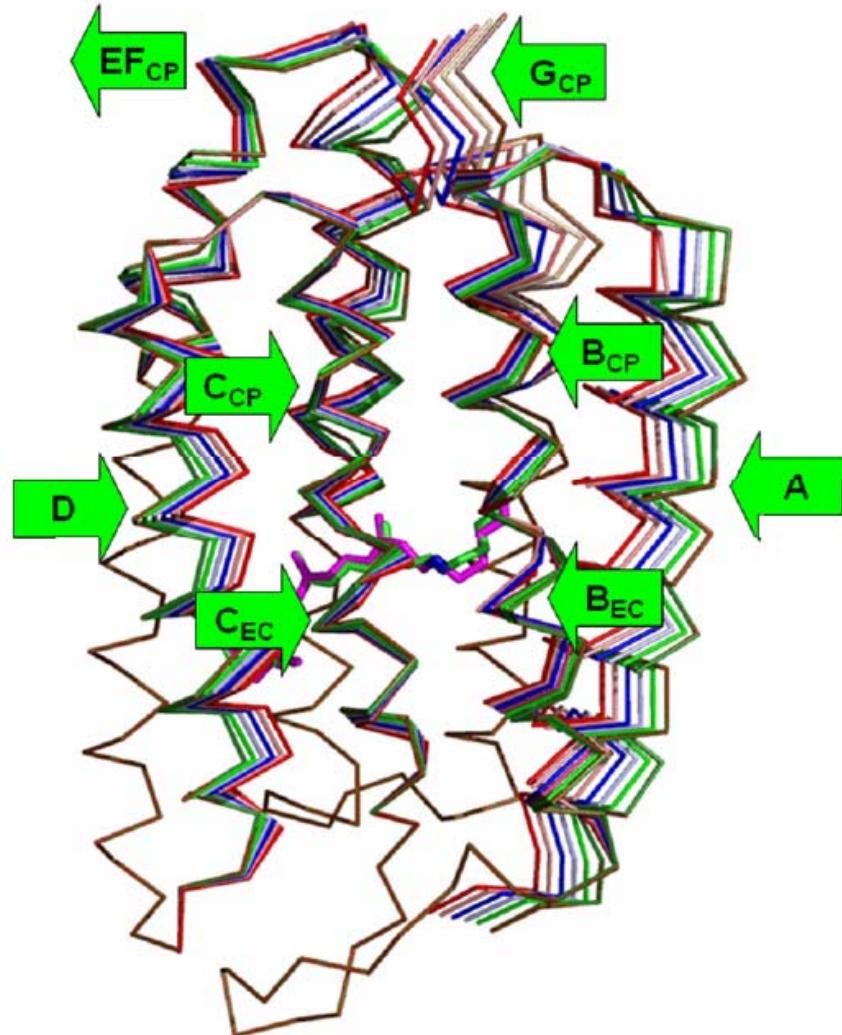


# Experimental data & spectral decomposition



Andersson *et al.*, *Structure* (2008)

# Structural Refinement

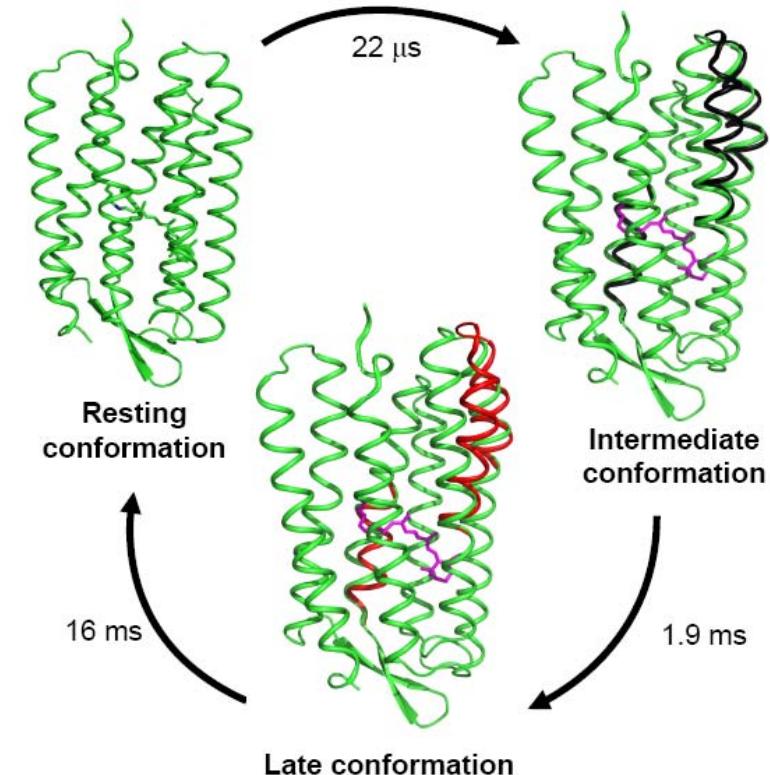


Andersson *et al.*, *Structure* (2008)

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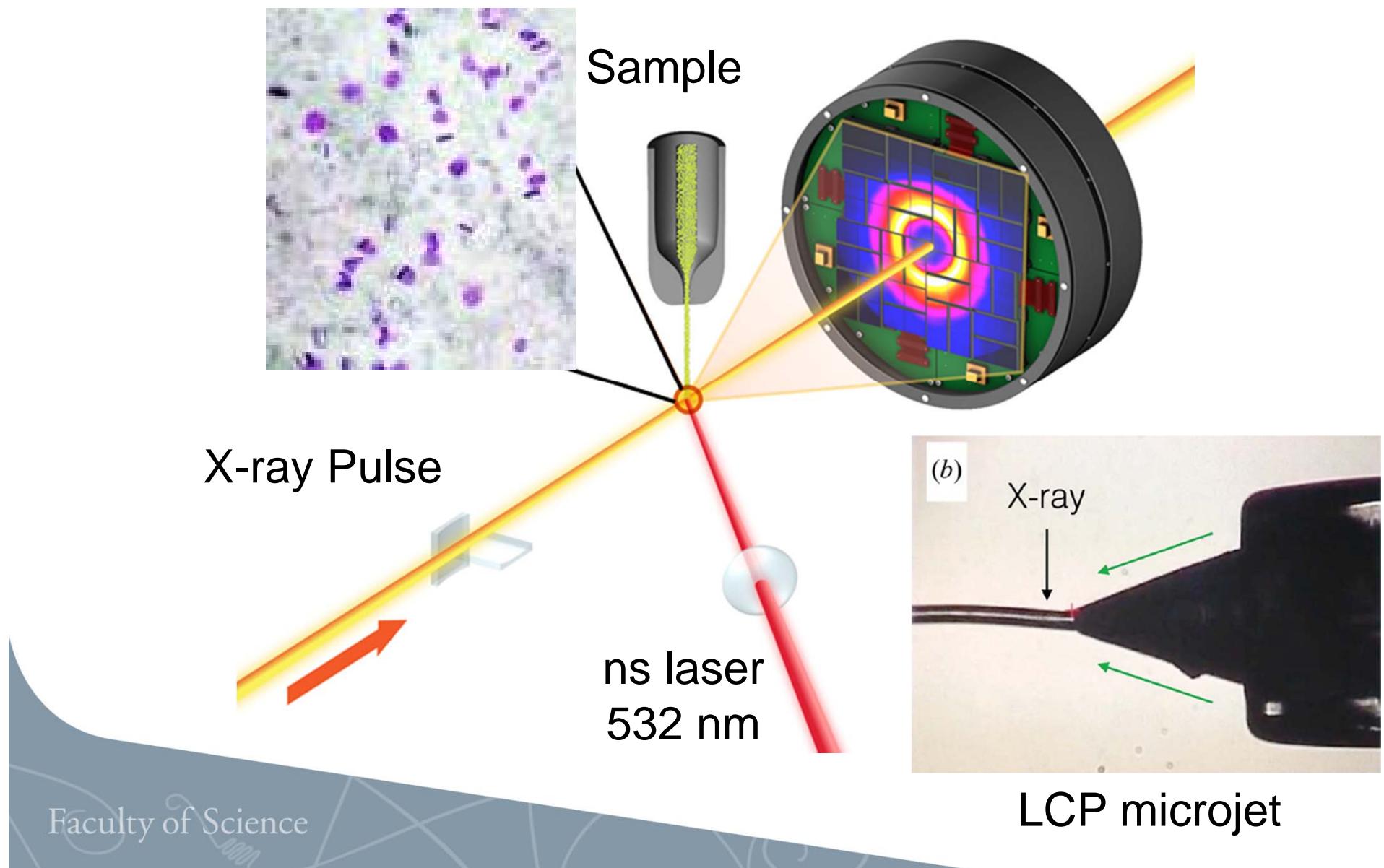
# Structural Conclusions

- Two intermediate conformations.
- Movements of helices E, F & C used to model these changes.
- 2/3 of the movement occurs prior to the first proton transfer step.



Andersson *et al.*, *Structure* (2008)

# Time Resolved Serial Crystallography





Eriko Nango

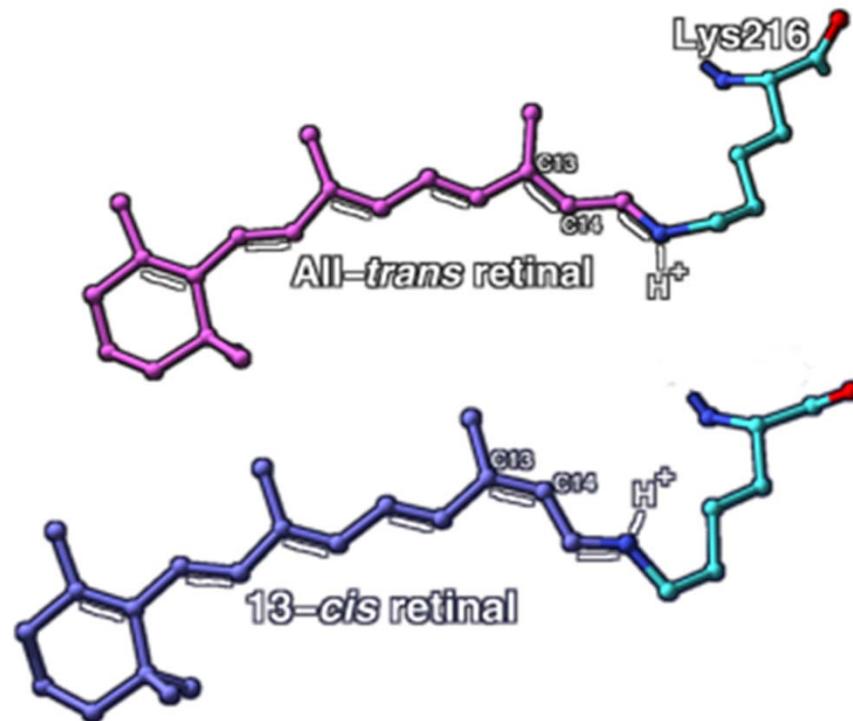
So Iwata

## SACLA at SPring8 Hyogo, Japan

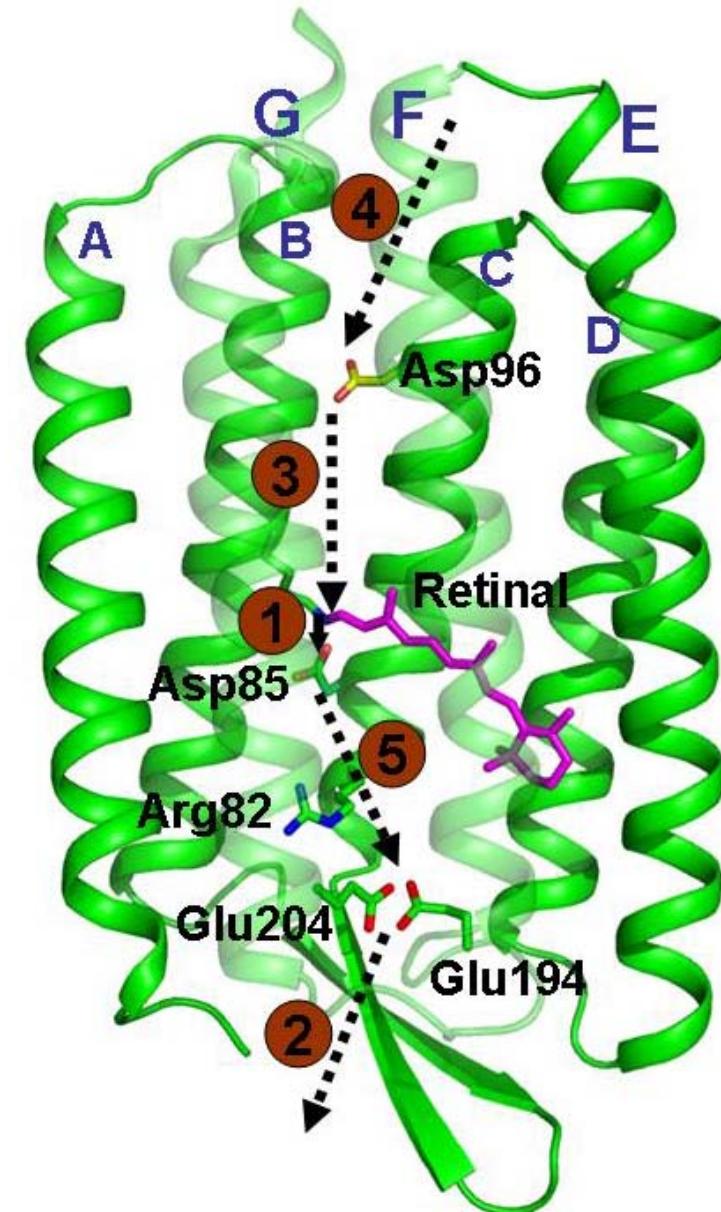


UNIVERSITY OF GOTHENBURG

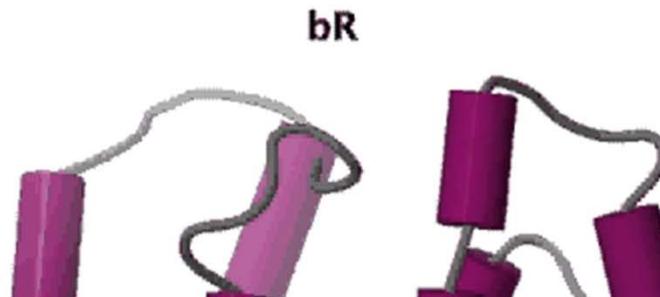
# Bacteriorhodopsin



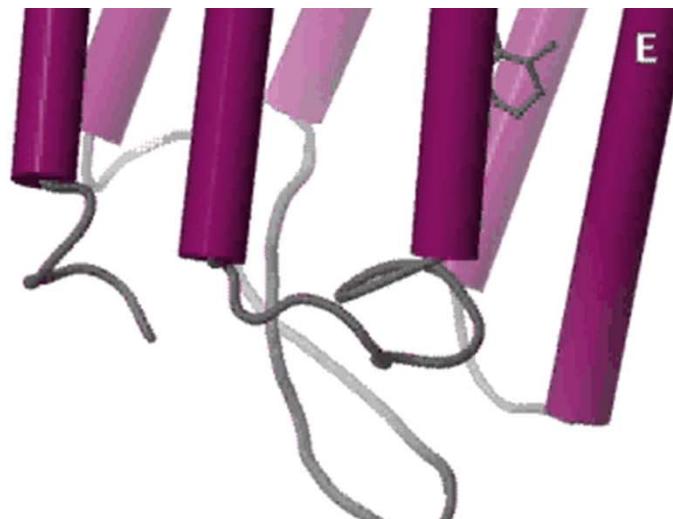
- Retinal isomerisation starts a sequence of structural changes.
- One proton pumped per photon.



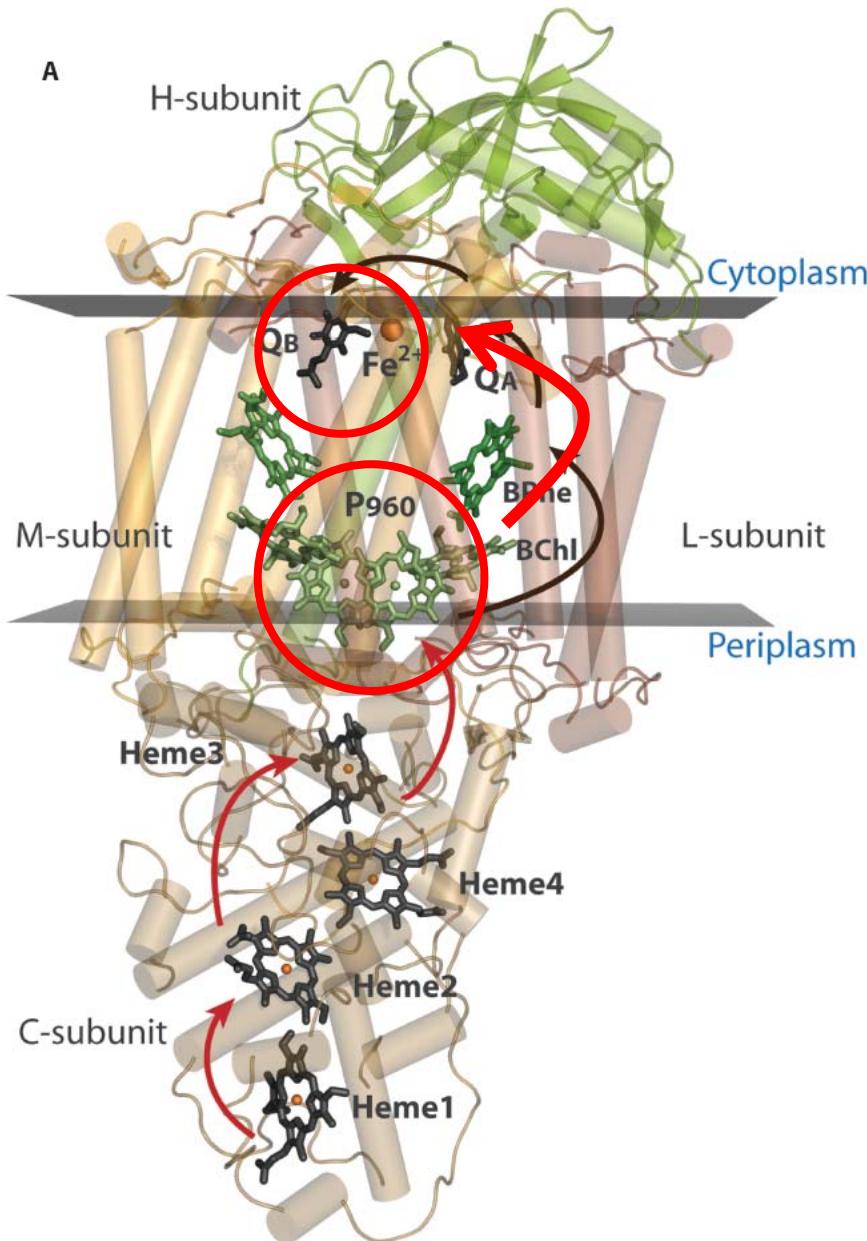
# Photocycle of bacteriorhodopsin



- Highly coordinated structural changes control the pKa of key proton-exchange groups.
- Understanding the nature & timing of these events central to untangling the mechanism of proton pumping.



# Photosynthetic reaction centres



- 135 kDa Membrane protein.
- Complex light driven proton pump.
- Electron movements driven by light.
- Coupled redox reactions pump protons.
- Descendent created O<sub>2</sub> rich atmosphere.

# Time-resolved spectroscopy

*Nature* 363, 320 (1993)

ARTICLES

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## **Visualization of coherent nuclear motion in a membrane protein by femtosecond spectroscopy**

**Marten H. Vos<sup>\*</sup>, Fabrice Rappaport<sup>\*</sup>, Jean-Christophe Lambry<sup>\*</sup>, Jacques Breton<sup>†</sup>  
& Jean-Louis Martin<sup>\*‡</sup>**

“implicates coherent nuclear motion in the primary electron transfer reaction in functional reaction centres”



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# Time-resolved spectroscopy

*Science* 316, 747 (2007)

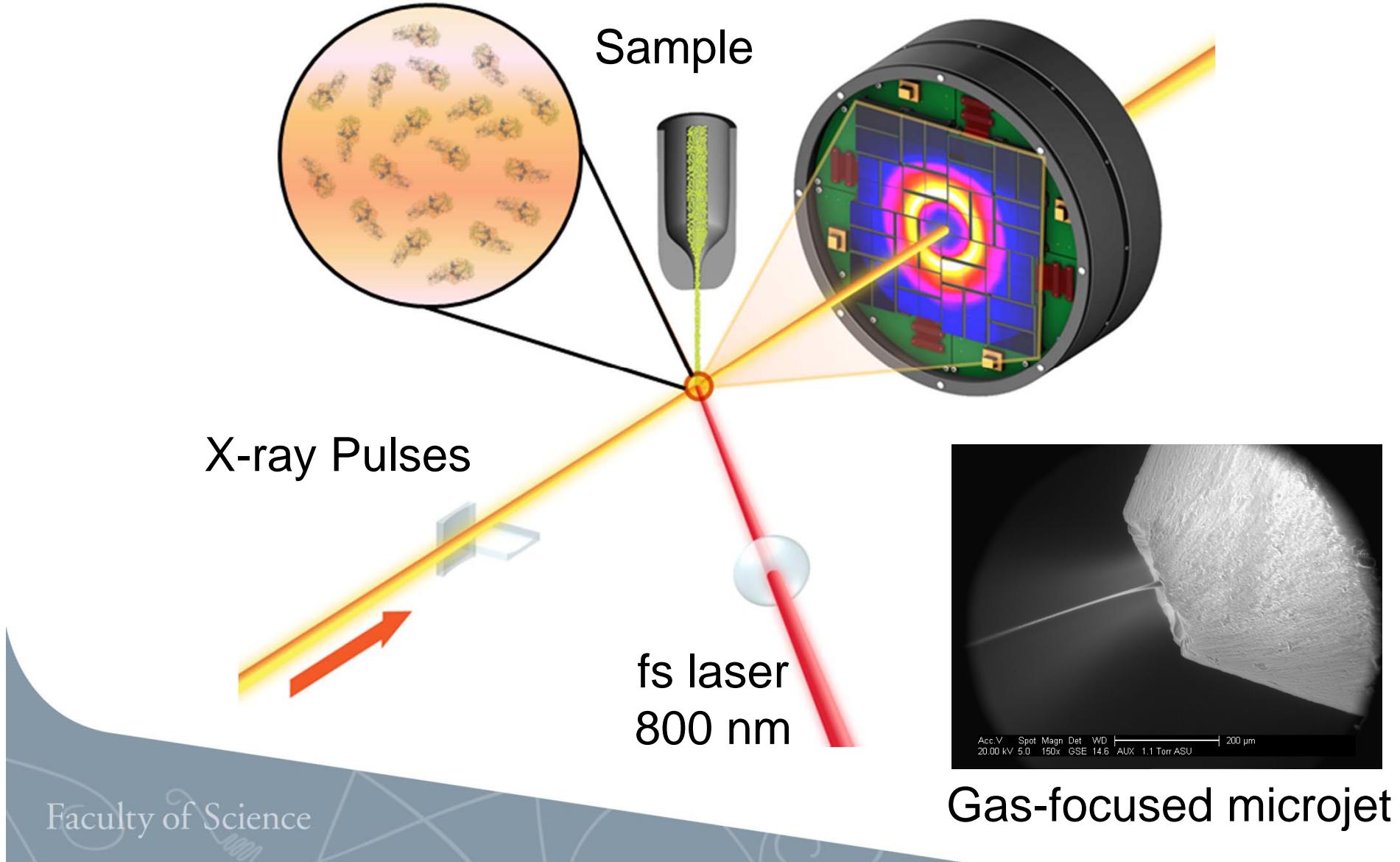
## Protein Dynamics Control the Kinetics of Initial Electron Transfer in Photosynthesis

Haiyu Wang,<sup>1,2</sup> Su Lin,<sup>1,2</sup> James P. Allen,<sup>2</sup> JoAnn C. Williams,<sup>2</sup> Sean Blankert,<sup>1,2</sup>  
Christa Laser,<sup>1,2</sup> Neal W. Woodbury<sup>1,2\*</sup>

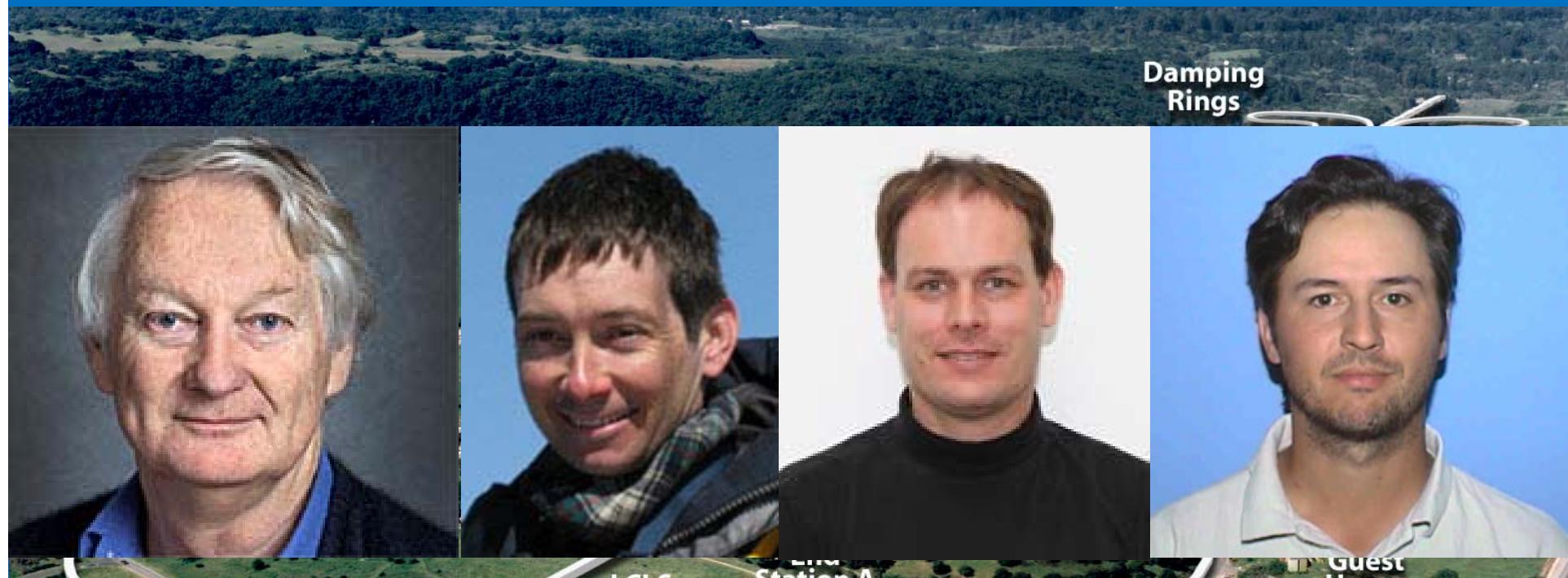
“These results indicate that initial photosynthetic charge separation is limited by protein dynamics rather than by a static electron transfer barrier”



# Time resolved wide angle X-ray scattering



# Linac Coherent Light Source (LCLS) Stanford, USA



John Spence

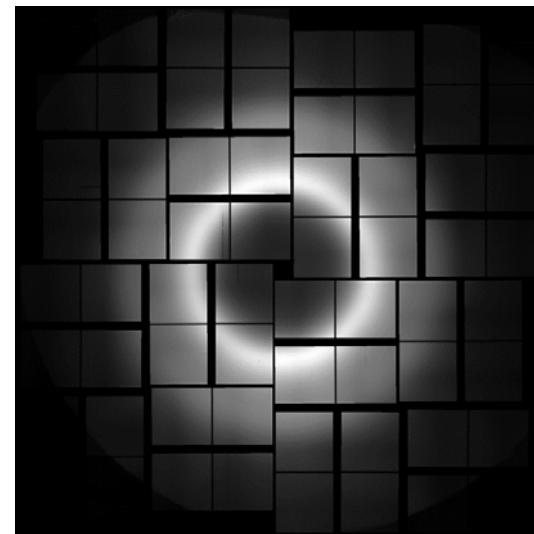
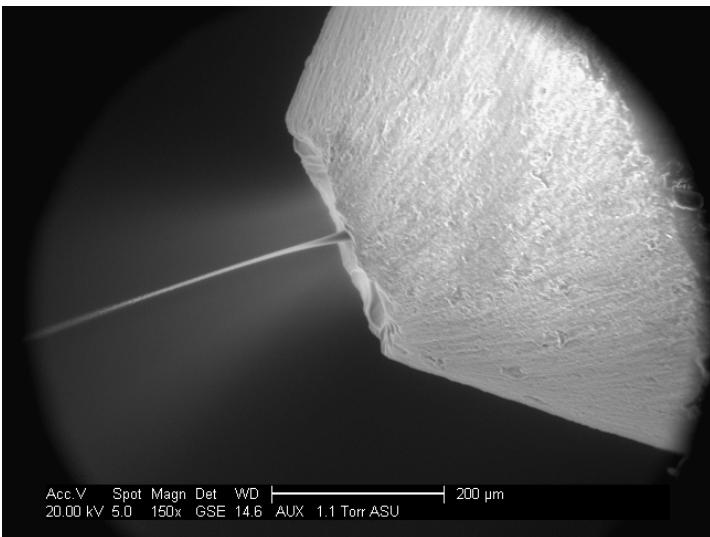
Henry Chapman

Anton Barty

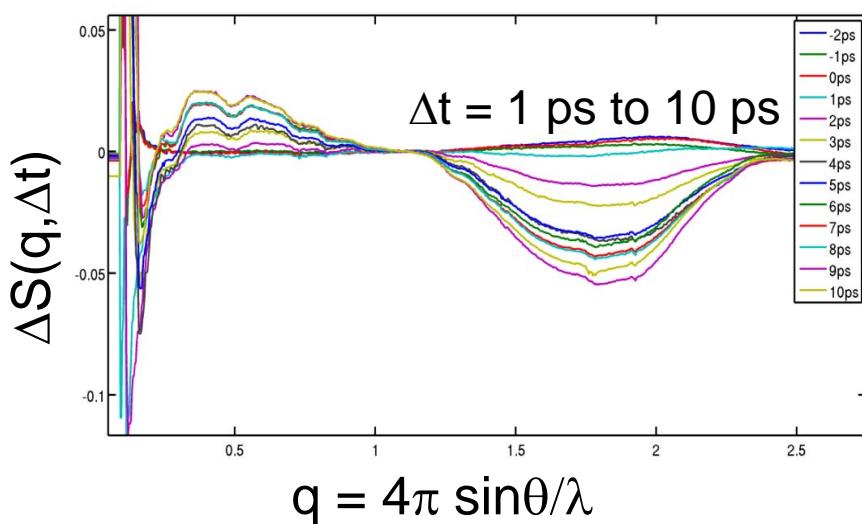
Sébastien Boutet



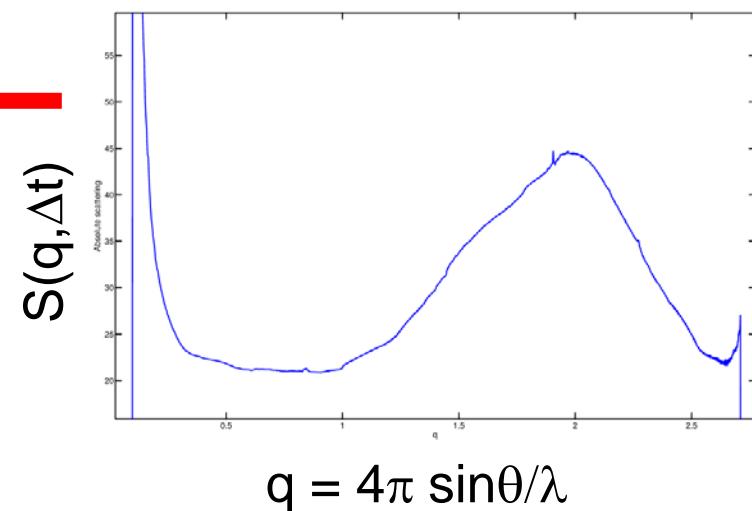
# Time resolved wide angle X-ray scattering



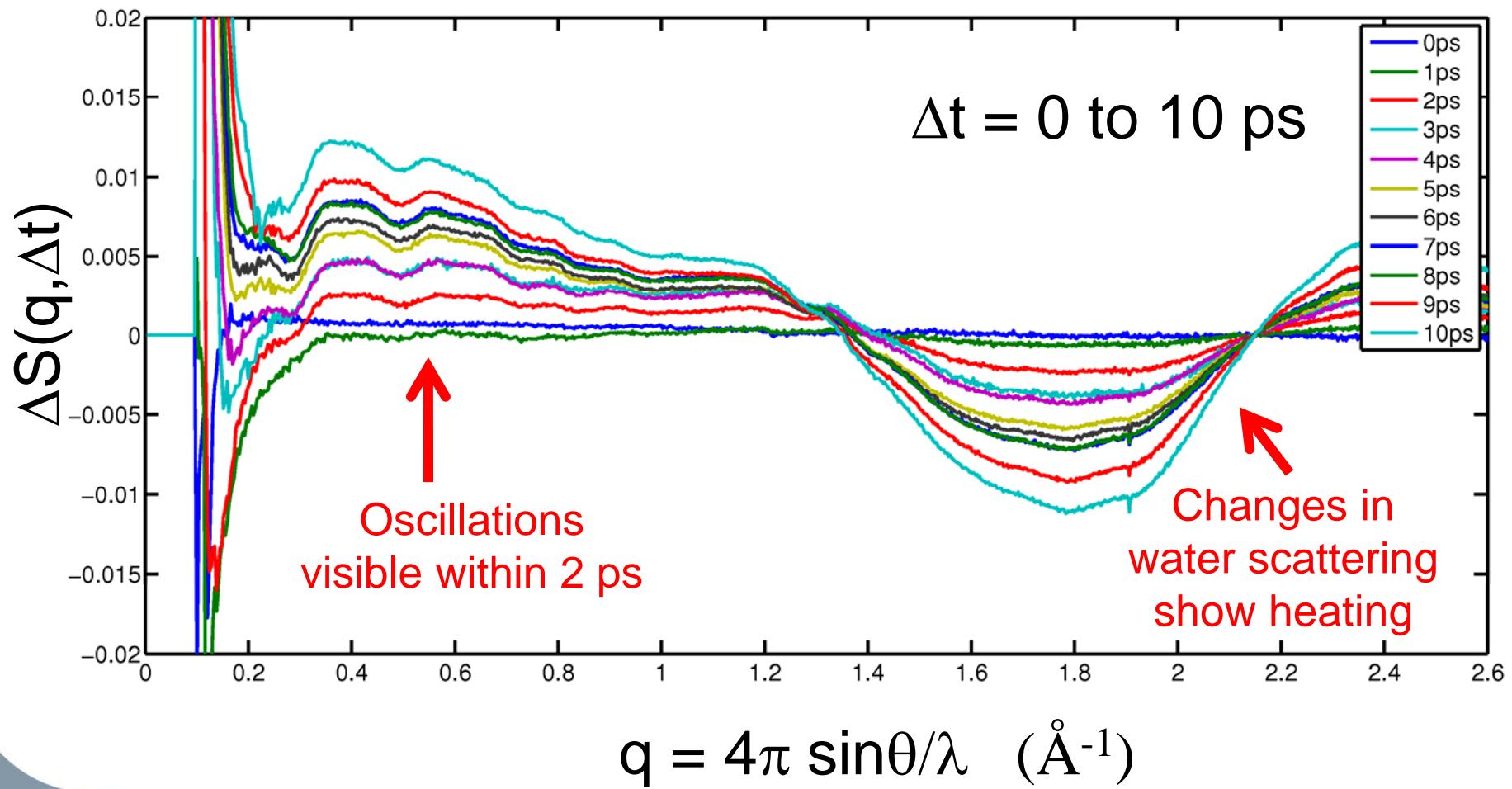
Laser on minus laser off



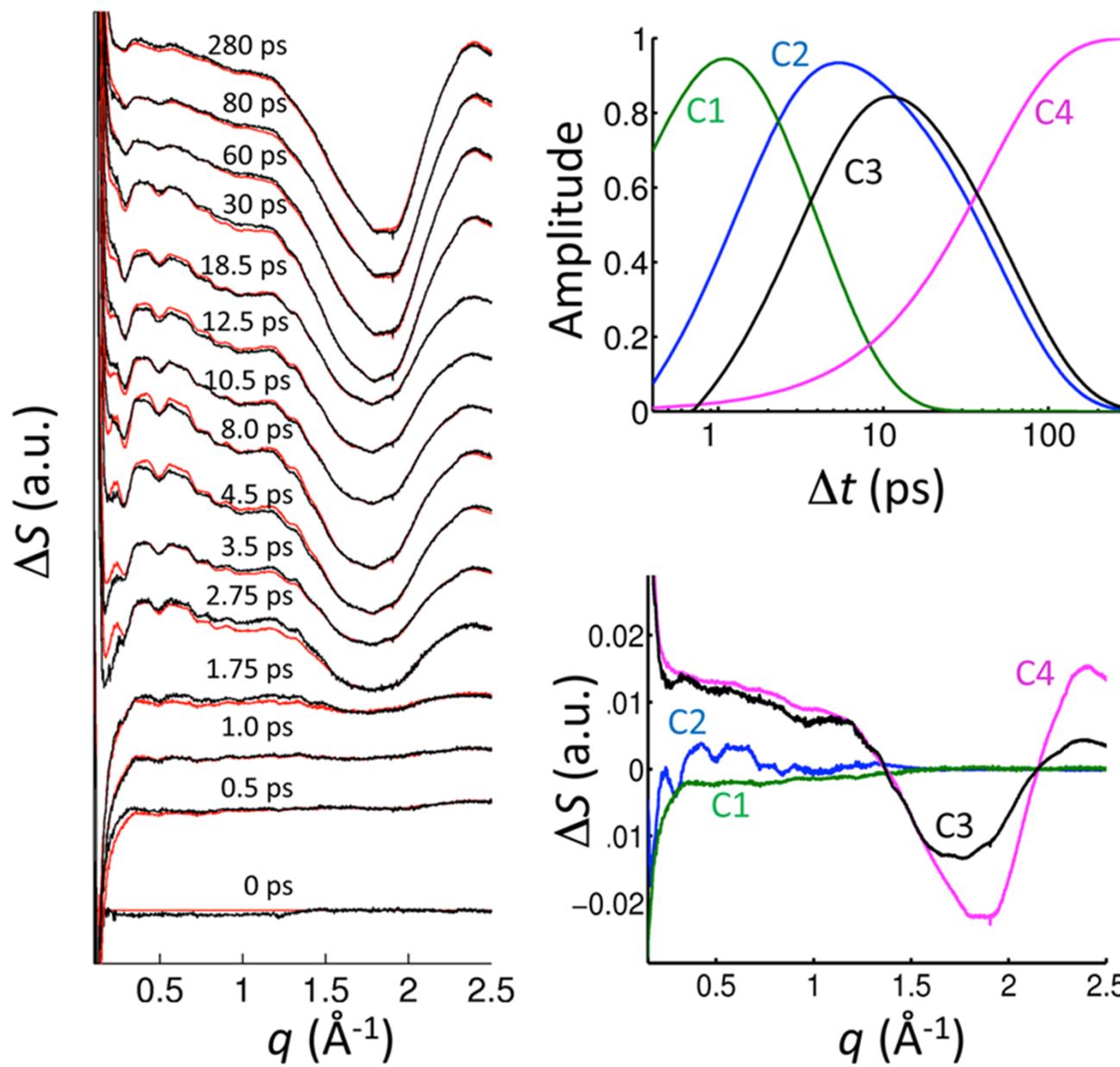
Radial Integration



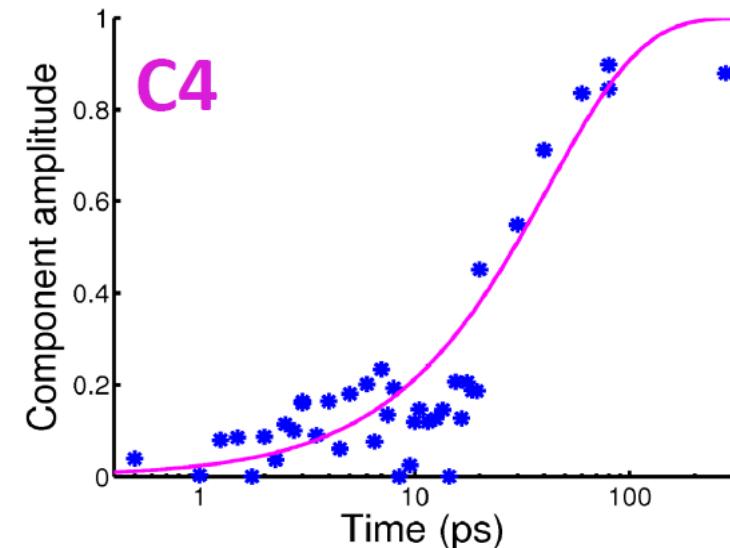
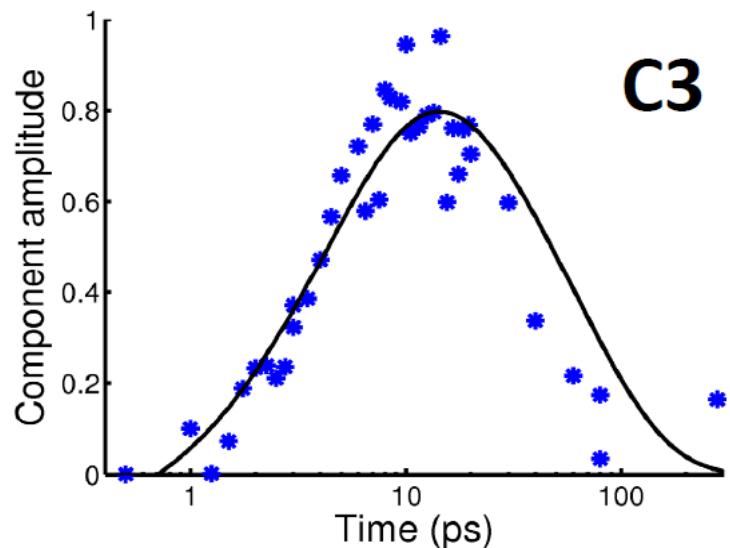
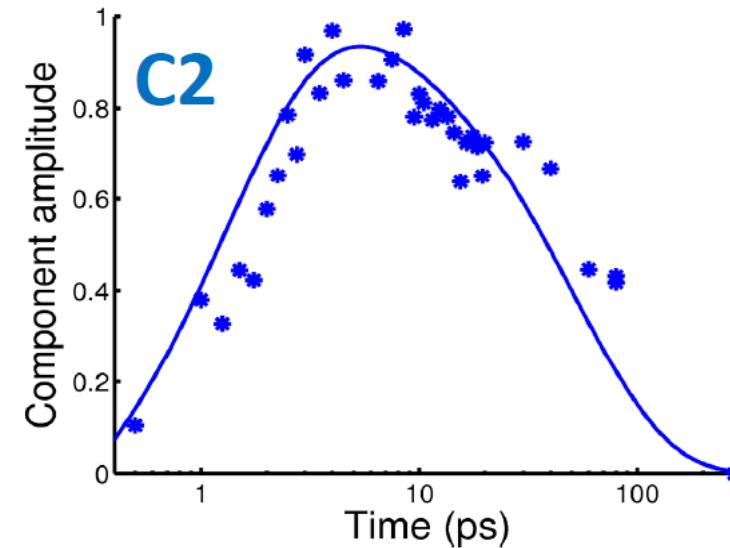
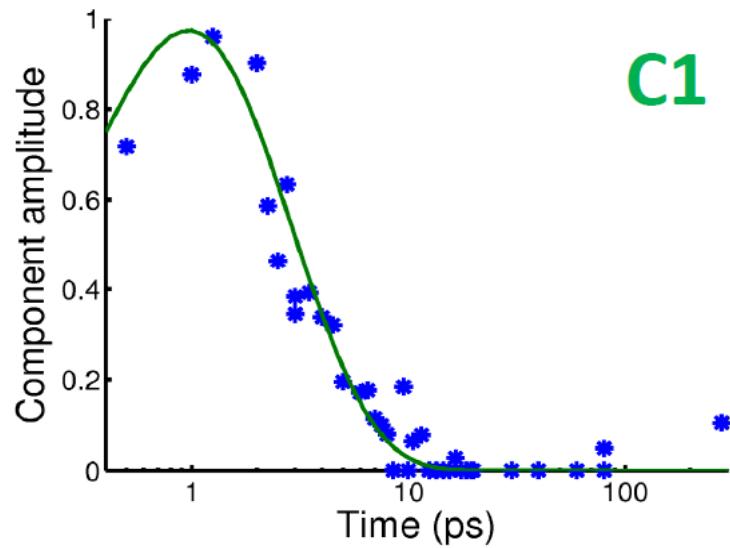
# TR-WAXS: Laser on minus laser off



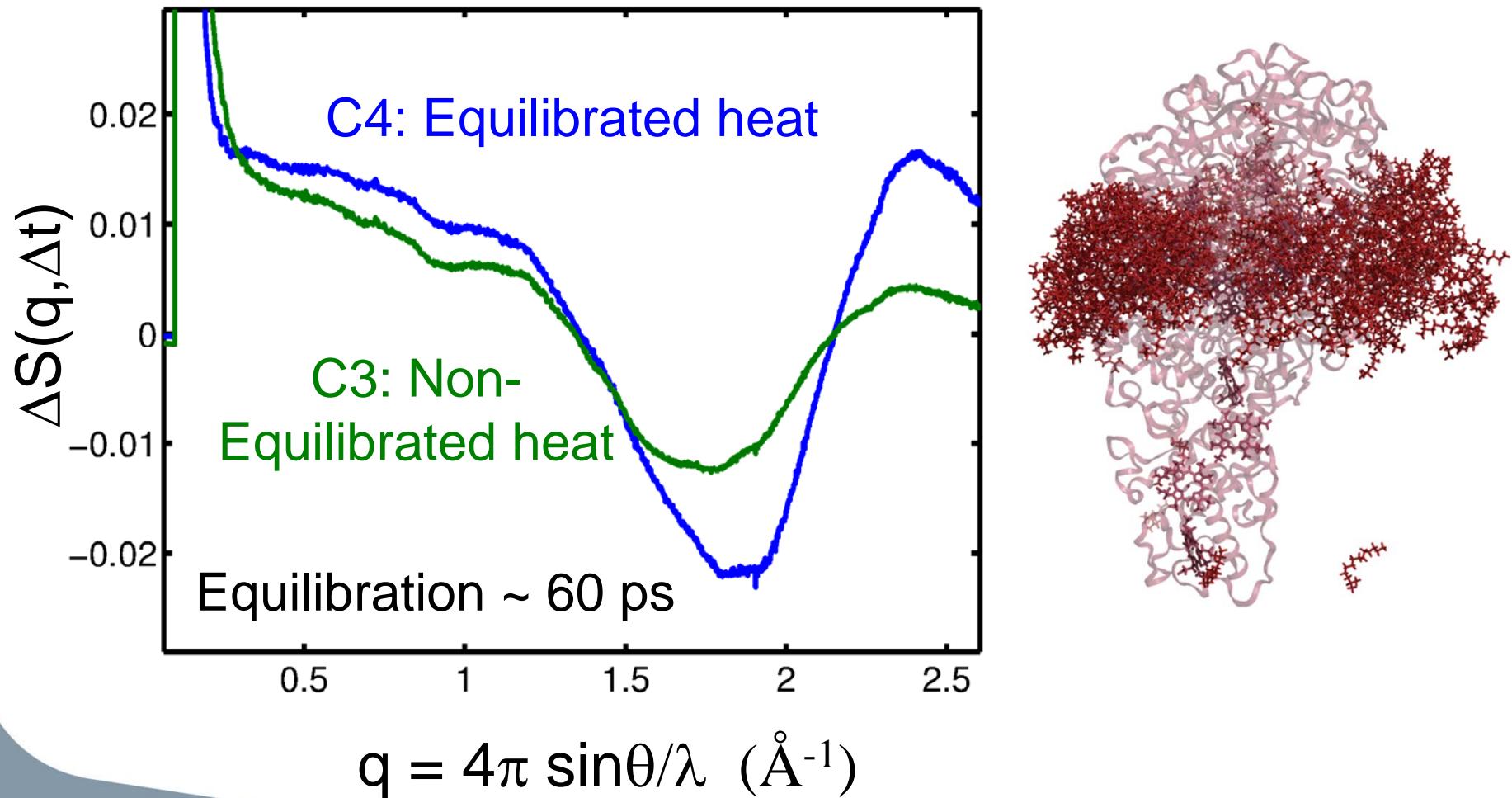
# Spectral decomposition



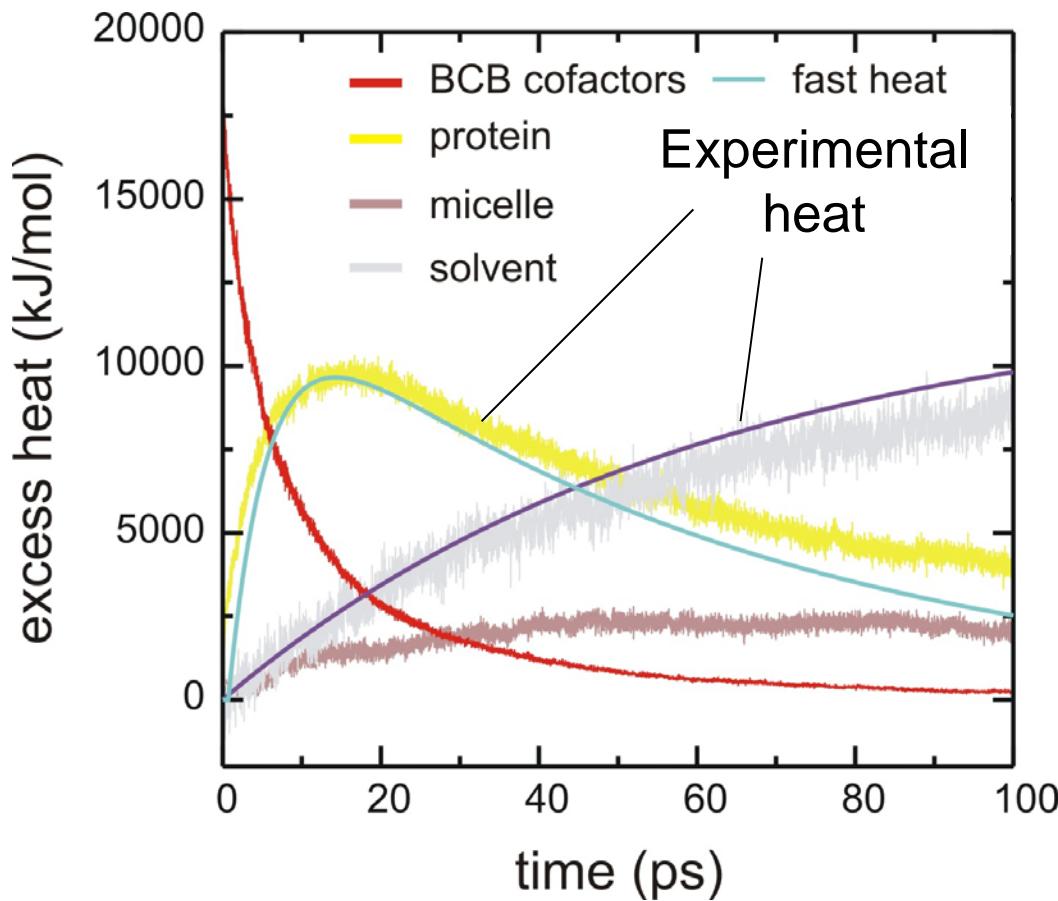
# Linear recombination of amplitudes



# Equilibrated & non-equilibrated heating

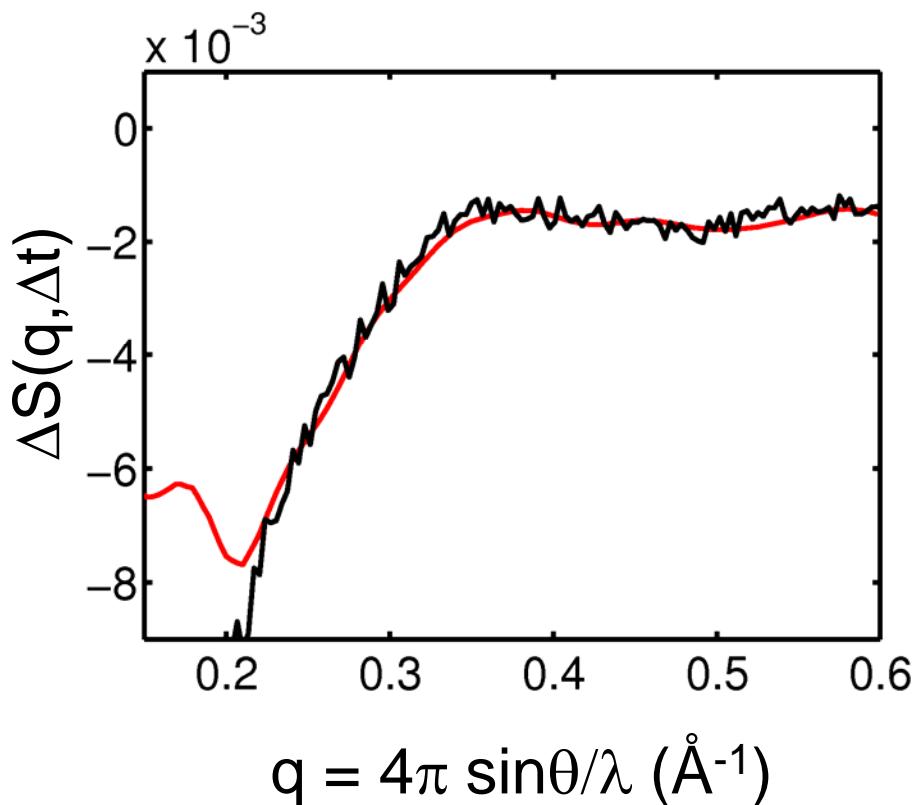


# MD simulations: flow of heat

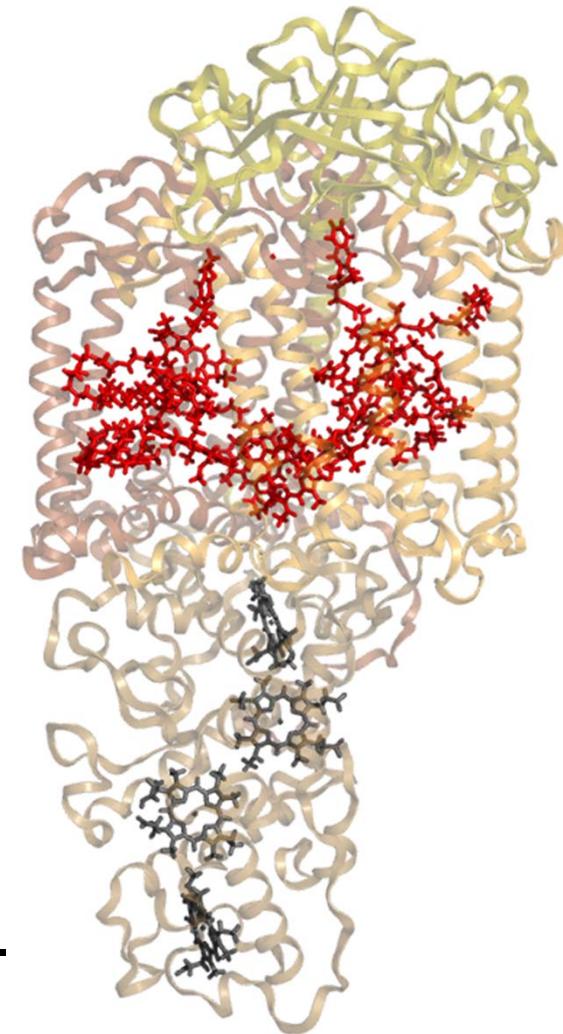


- MD simulations & TR-WAXS measurements both show heat equilibrates within ~ 100 ps.

# Ultrafast-component

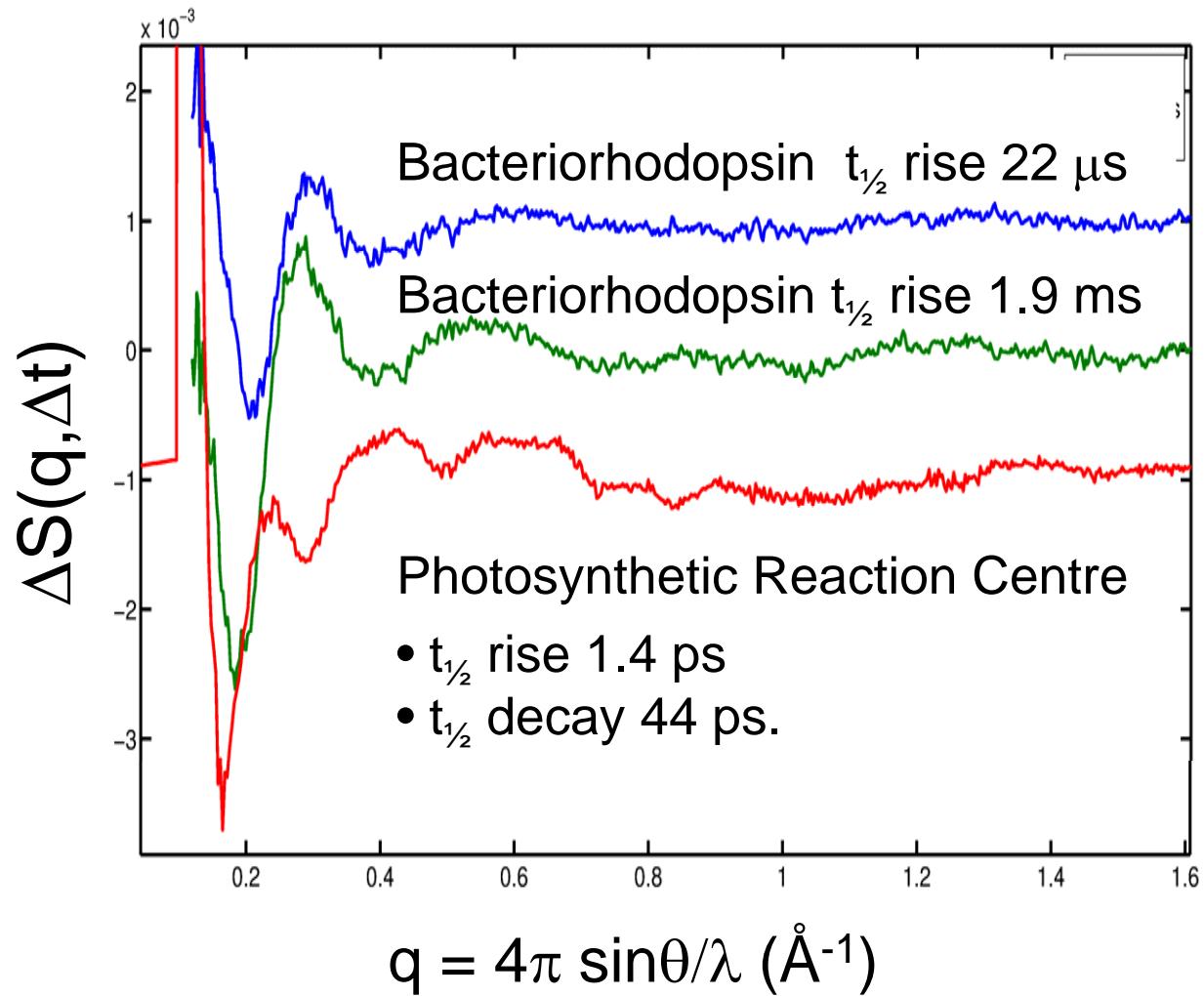


- Fitting with only co-factors moving.
- Rise: Sub-ps.
- $t_{1/2}$  decay  $\sim 3\text{-}2$  ps.

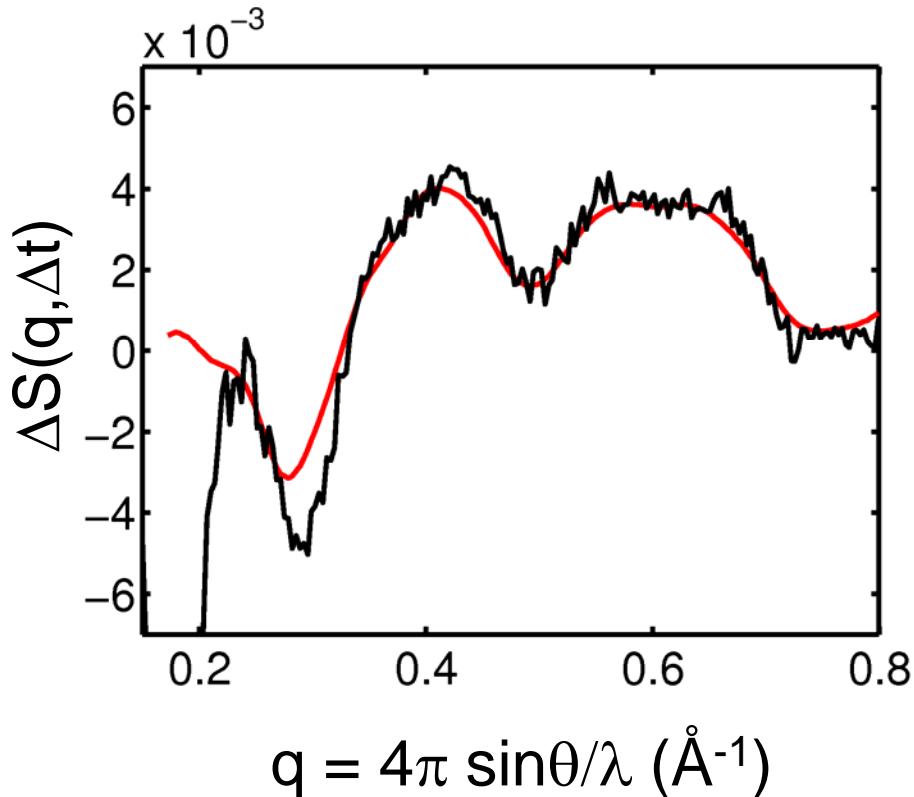


Arnlund *et al.*, *Nature Methods* (2014)

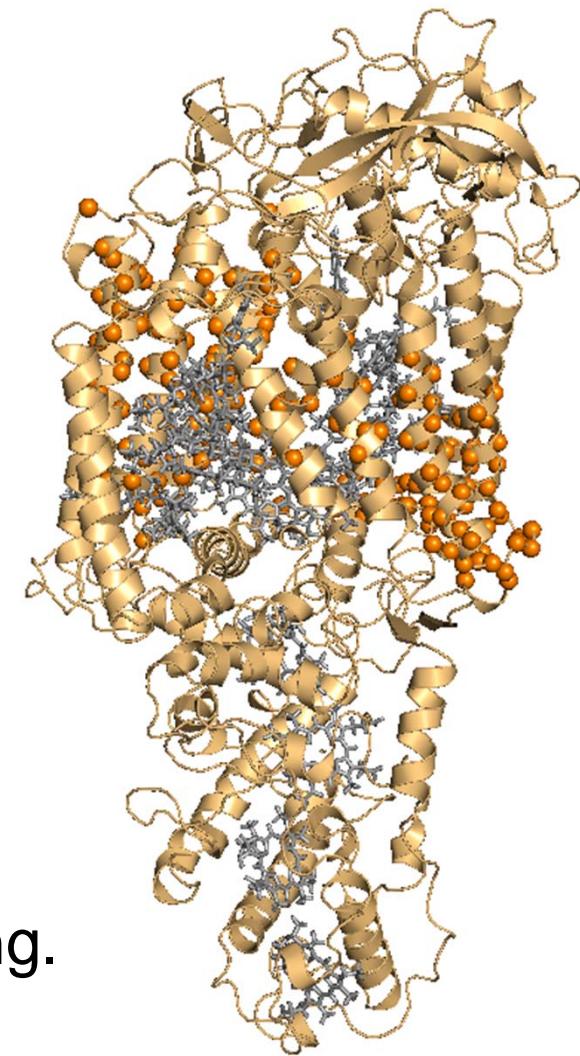
# c.w. TR-WAXS on bacteriorhodopsin



# Fitting protein component

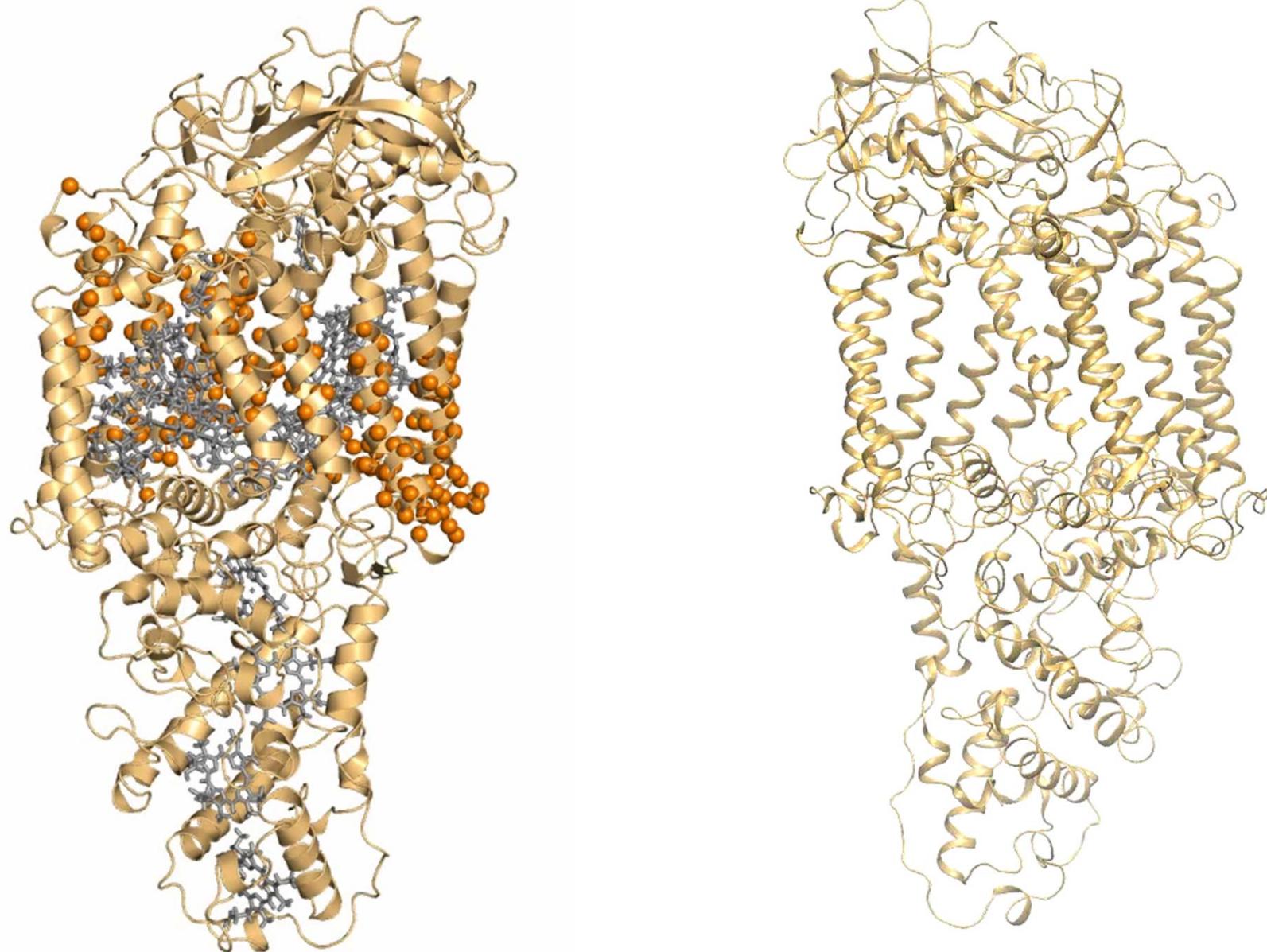


- Fitting with all protein atoms moving.
  - $t_{1/2}$  rise  $\sim 1.4$  ps
  - $t_{1/2}$  decay  $\sim 44$  ps.



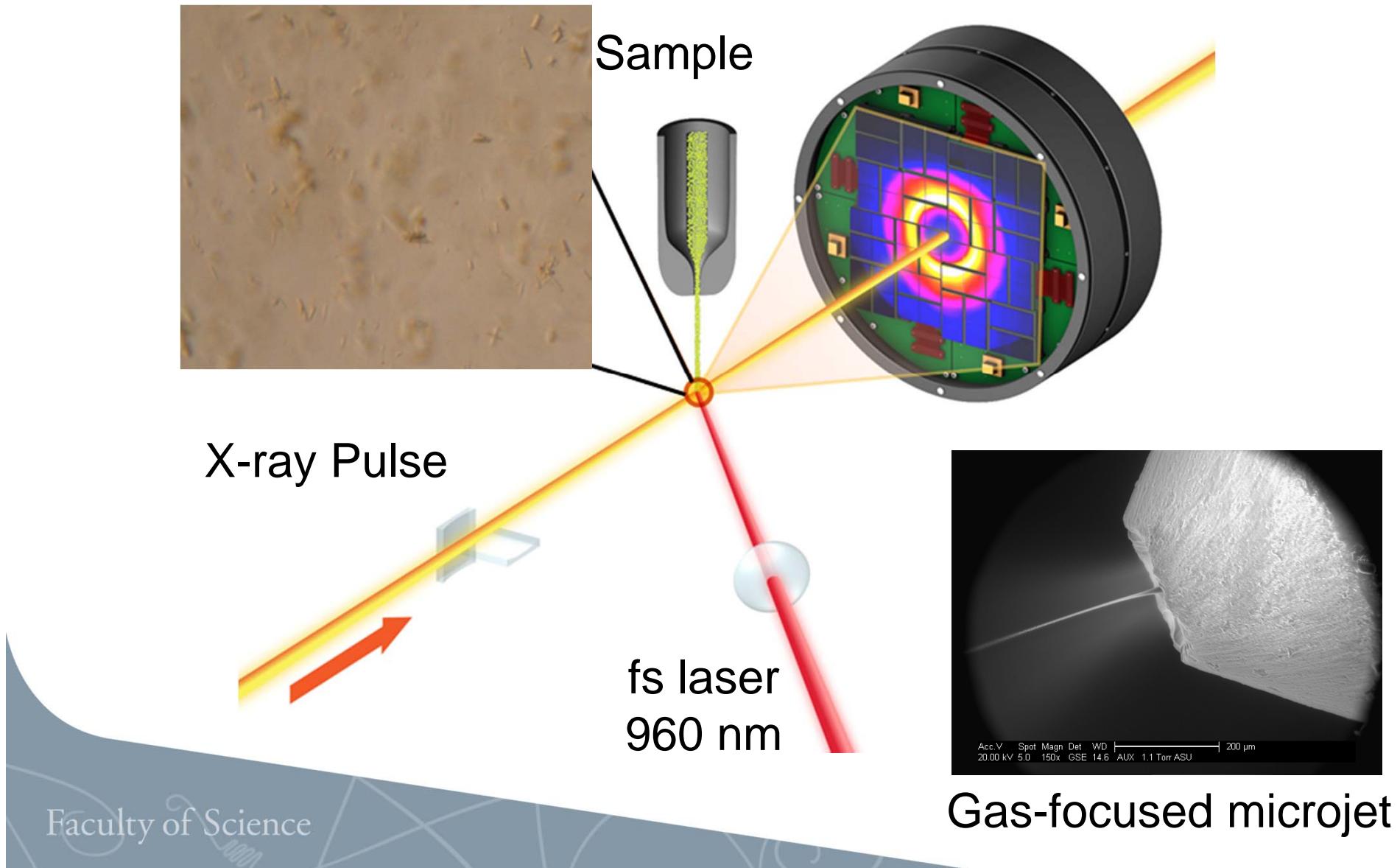
Arnlund *et al.*, *Nature Methods* (2014)

# Overall conformational change

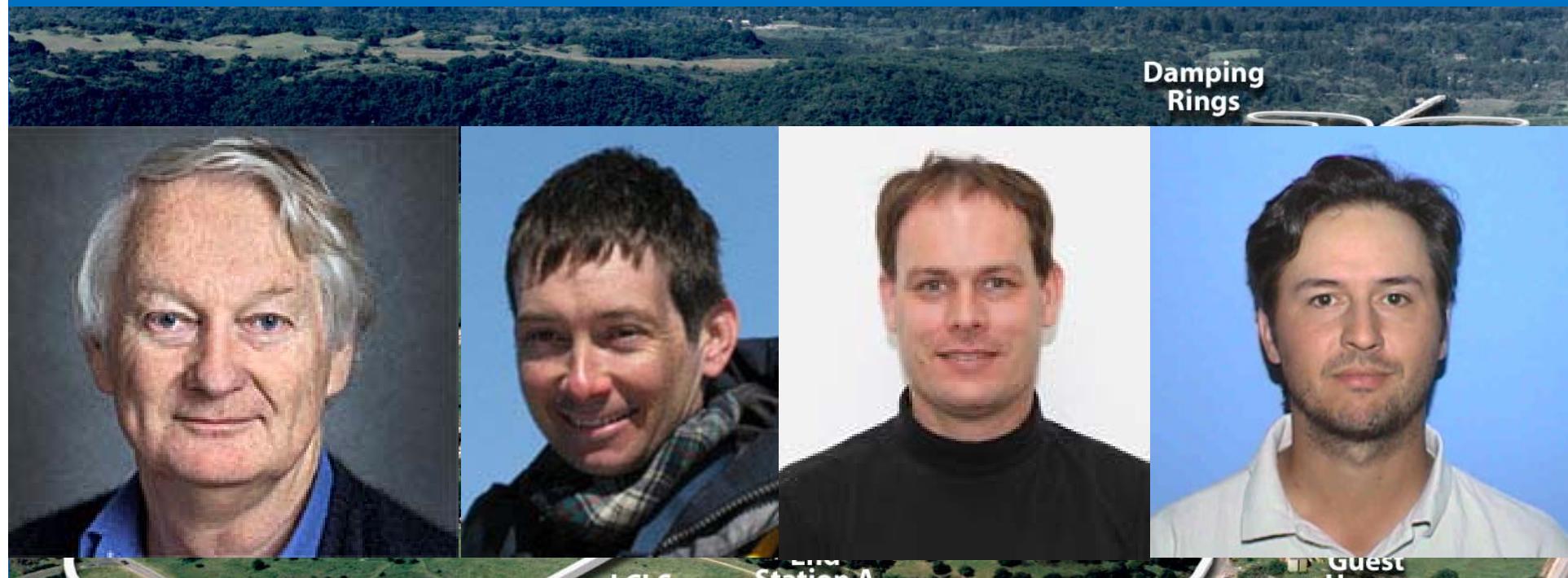




# Time Resolved Serial Crystallography



# Linac Coherent Light Source (LCLS) Stanford, USA



John Spence

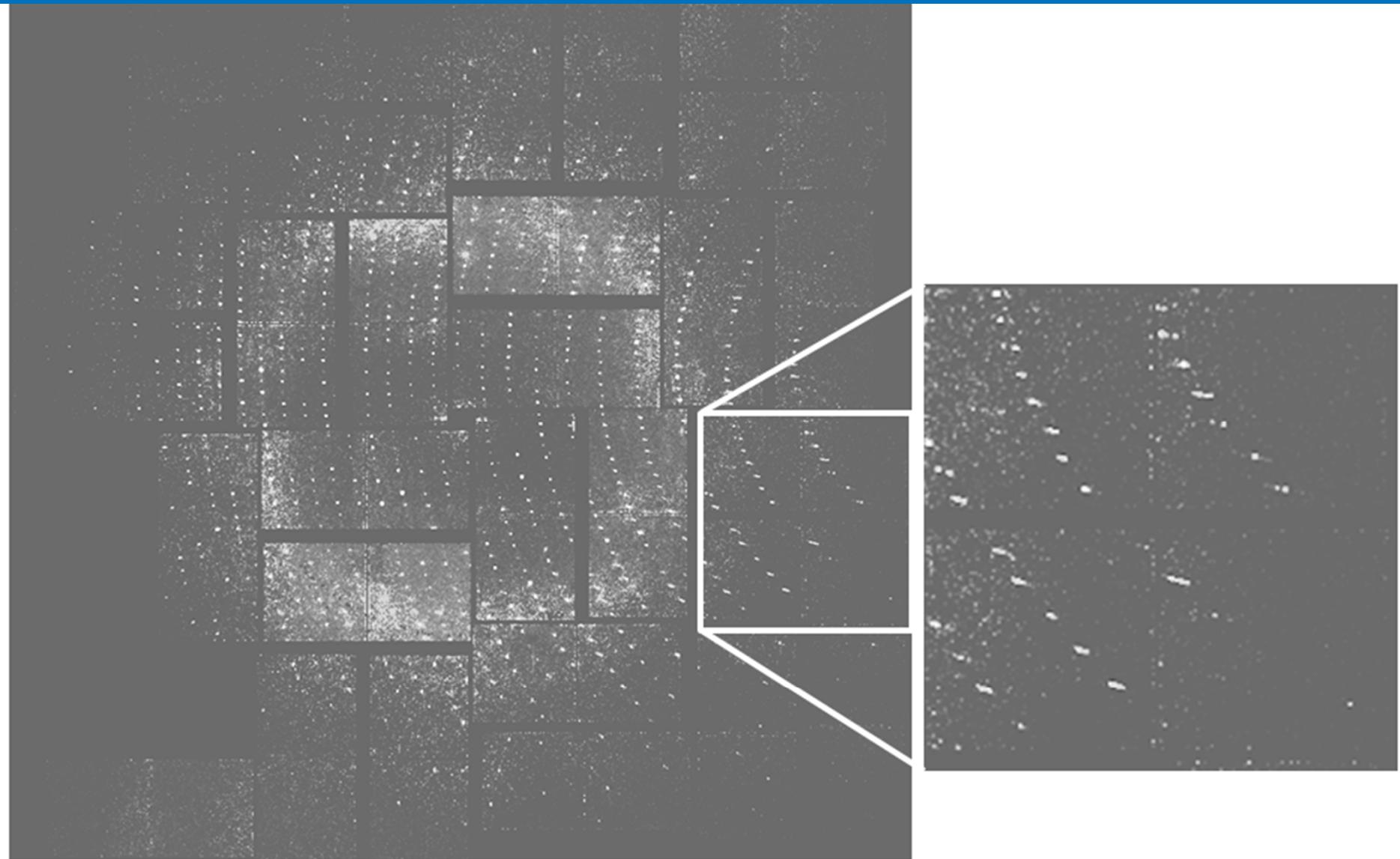
Henry Chapman

Anton Party

Sebastien Boutet

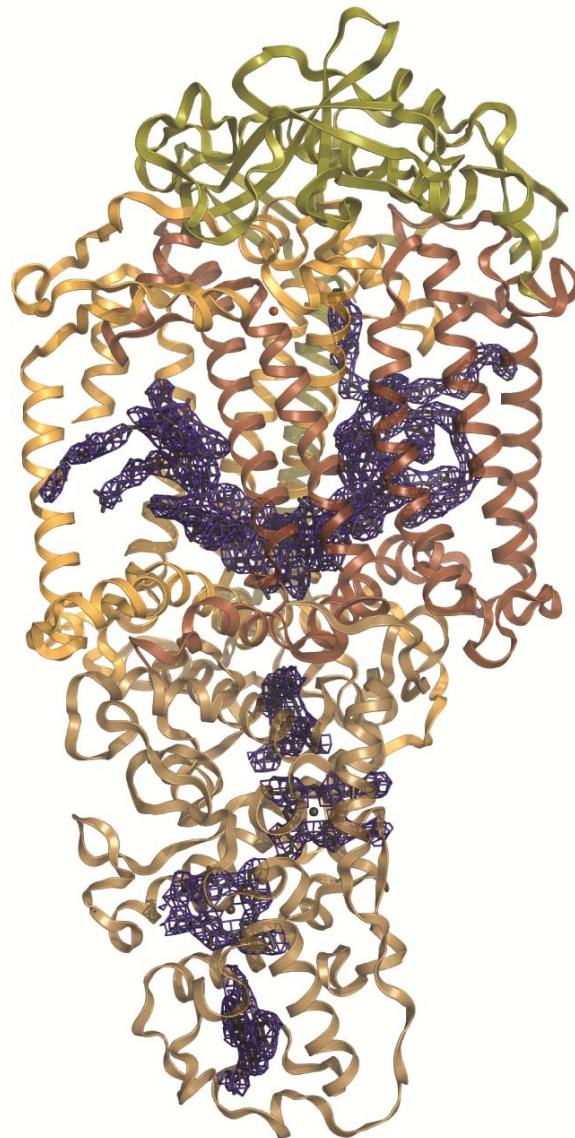


# Data extend to 2.8 Å resolution

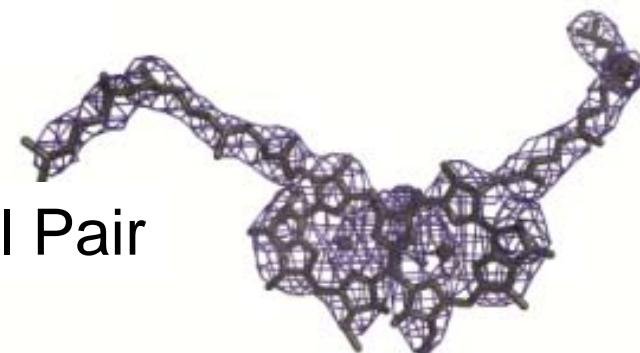


Johansson *et al.*, *Nature Communications* (2013).

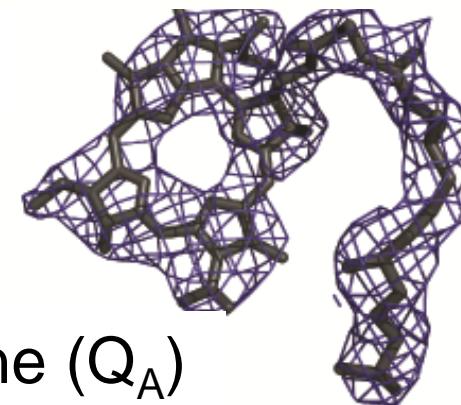
# 3.5 Å SFX structure



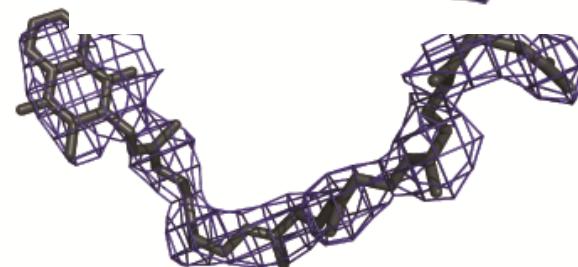
Special Pair



Pheophytin



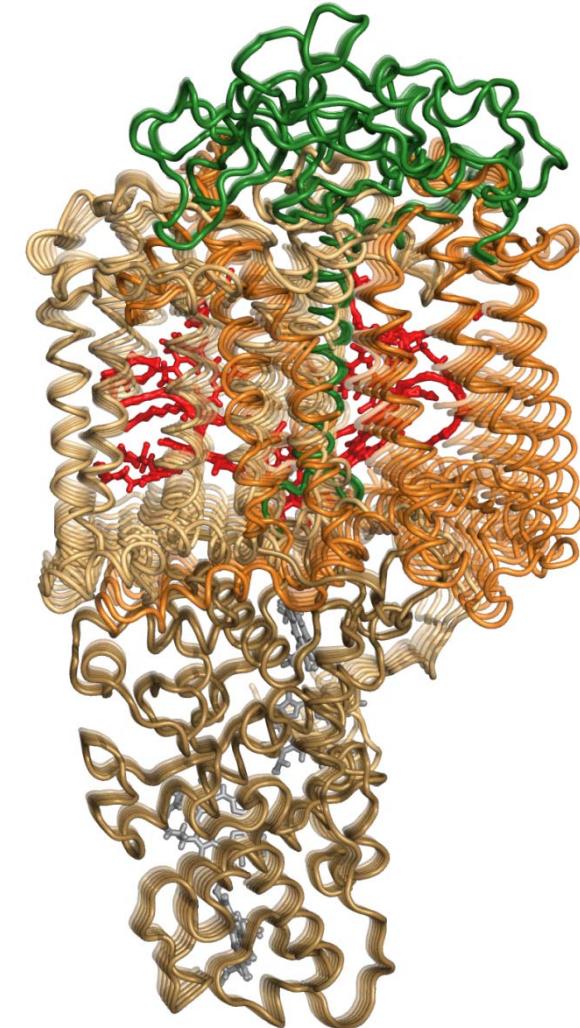
Menaquinone ( $Q_A$ )



Johansson *et al.*, *Nature Communications* (2013)

# Ultrafast conformational gating?

- Photosynthetic reaction centres remarkably efficient.
  - Multiple contributing factors.
- Structural changes allow the energy surface for an electron on *the way out* to differ from that *back home*.
- May help extend the lifetime of the charge separated state.



# Serial Femtosecond Crystallography @LCLS

<b>CFEL-DESY</b>	<b>H. Chapman</b> , J. Schulz, A. Barty, M. Liang, A. Aquila, T. White, D. Deponte, S. Stern, A. Martin, C. Caleman, K. Nass, F. Stellato, F. Wang, H. Fleckenstein, L. Gumprecht, L. Holmegaard, N. Coppola, S. Bajt, M. Barthelmess,
<b>ASU</b>	<b>J. Spence, P. Fromme</b> , U. Weierstall, B. Doak, M. Hunter, R. Kirian, X. Wang, K. Schmidt, I. Grotjohann, R. Fromme
<b>Gothenburg</b>	<b>L. Johansson, D. Arnlund</b> , G. Katona, E. Malmerberg
<b>SLAC-PULSE</b>	M. Bogan, D. Starodub, R. Sierra, C. Hampton, D. Loh
<b>SLAC-LCLS</b>	<b>S. Boutet, G. Williams</b> , M. Seibert, J. Kryzwinski, C. Bostedt, M. Messerschmidt, J. Bozek, W. White, R. Coffee
<b>Uppsala</b>	J. Hajdu, Nic Timneanu, J. Andreasson, M. Seibert, F. Maia, M. Svenda, <b>J. Davidsson</b>
<b>MPG CFEL ASG</b>	<b>I. Schlichting</b> , R. Shoeman, L. Lomb, S. Kessemeyer, T. Barends, J. Steinbrener, M. Bott, D. Rolles, S. Epp, A. Rudenko, L. Strüder, R. Hartmann, L. Foucar, N. Kimmel, P. Holl, T. Barends, J. Ullrich
<b>LLNL</b>	S. Hau-Riege, M. Frank
<b>LBNL</b>	S. Marchesini, J. Holton
<b>Cornell</b>	V. Elser, S. Gruner
<b>CAMP Team</b>	Led by <b>J. Ullrich</b> and <b>I. Schlichting</b>
<b>LCLS detector</b>	C. Kenney, R. Herbst, J. Pines, P. Hart, J. Morse
<b>Accelerator</b>	Led by <i>P. Emma</i>

# Time resolved WAXS @ LCLS

<b>Gothenburg</b>	<b>D. Arnlund, L. Johansson, C. Wikstrand, R. Dods, E. Malmerberg,</b> G. Katona, J. Sjöhamn, S. Westenhoff.
<b>CFEL-DESY</b>	<b>A. Barty, H. Chapman,</b> J. Schulz, M. Liang, A. Aquila, T. White, D. Deponte, S. Stern, A. Martin, K. Nass, F. Stellato.
<b>ASU</b>	<b>J. Spence</b> , P. Fromme, U. Weierstall, B. Doak, R. Kirian, D. Wang, K. Schmidt, I. Grotjohann, R. Fromme, D. James.
<b>SLAC-LCLS</b>	<b>S. Boutet, G. Williams</b> , M. Seibert, J. Kryzinski, C. Bostedt, M. Messerschmidt, J. Bozek, W. White, R. Coffee
<b>Uppsala</b>	<b>J. Davidsson</b>
<b>MPI Heidelberg</b>	<b>I. Schlichting</b> , R. Shoeman, T. Barends, S. Bari
<b>LCLS detectors</b>	C. Kenney, R. Herbst, J. Pines, P. Hart, J. Morse
<b>LCLS accelerator</b>	Led by P. Emma
<b>LCLS fs laser</b>	D. Milathianaki, A. Fry.
<b>LLNL</b>	M. Frank
<b>Göttingen</b>	<b>G. Groenhof</b>
<b>APS BioCARS</b>	<b>R. Henning</b> , I. Kosheleva.
<b>TDU</b>	K. Skov Kjær, T. Brandt van Driel, M. Meedom Nielsen.

## Time resolved SFX @ SACLA

**Riken**

**Eriko Nango, So Iwata, Rie Tanaka,** Toshiaki Hosaka,  
Tomoyuki Tanaka, Ayumi Yamashita, Jun Kobayashi, Toshi  
Arima, Minoru Kubo, Tetsunari Kimura, Song Changyong.

**CNRS**

**Antoine Royant.**

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**Gothenburg**

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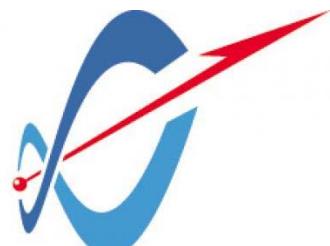
**Magnus Andersson, Erik Malmerberg,** Sebastian Westenhoff,  
Gergely Katona, Annemarie Wöhri, Linda Johansson

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