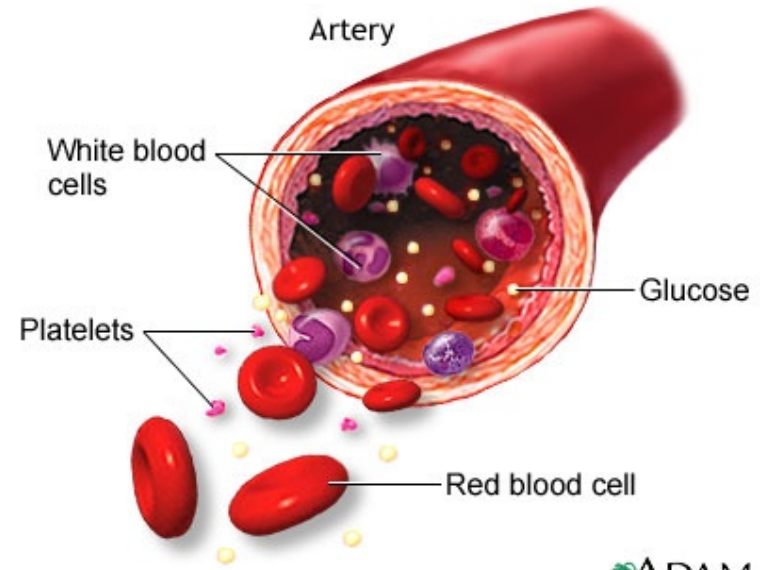
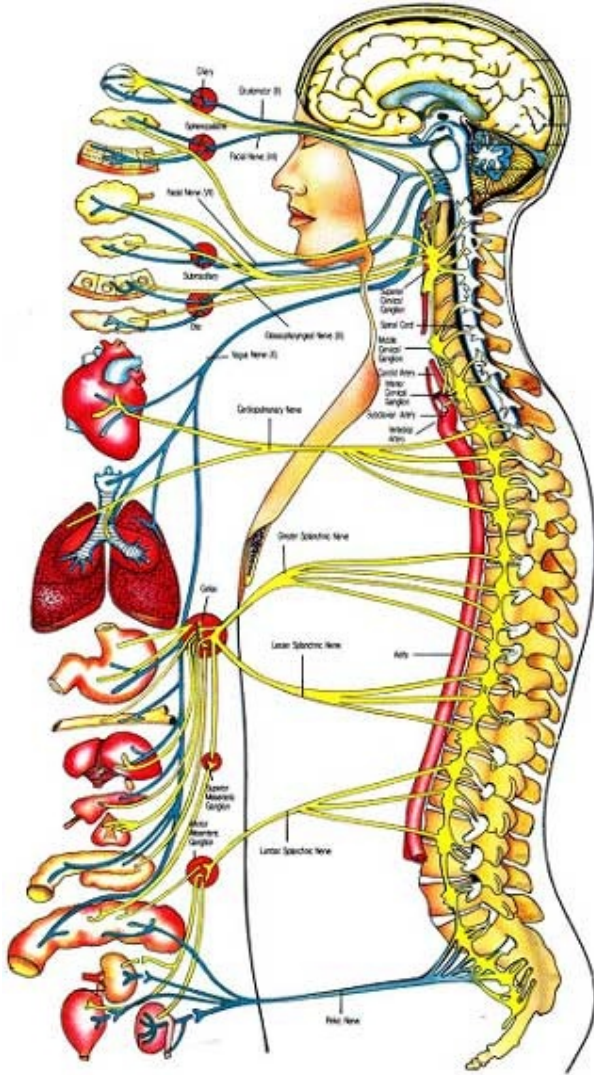
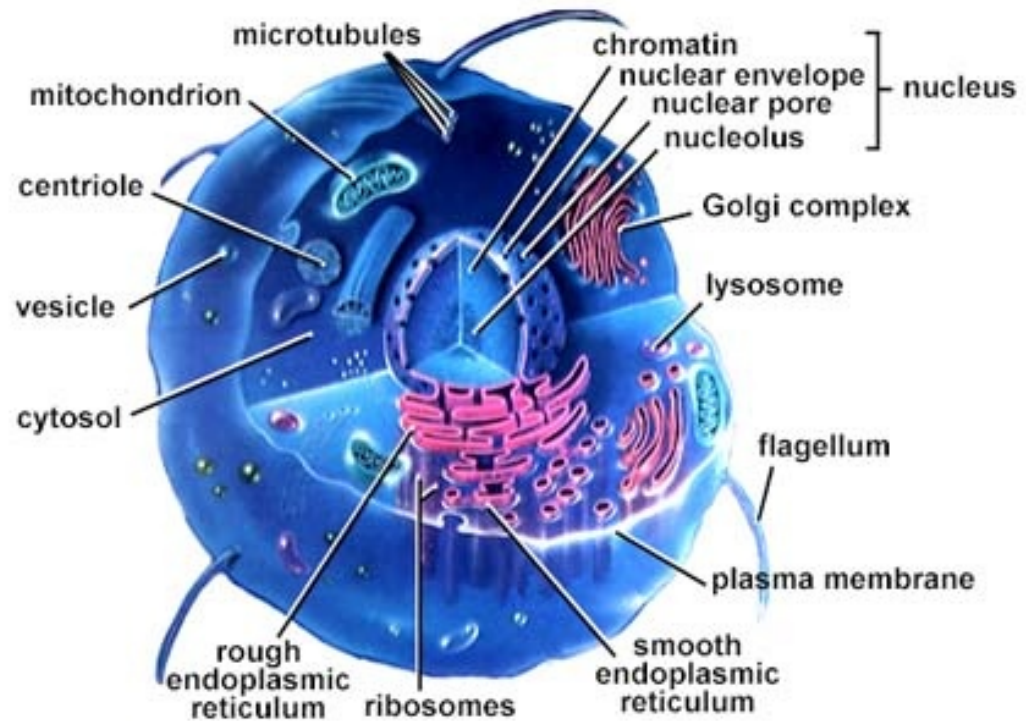
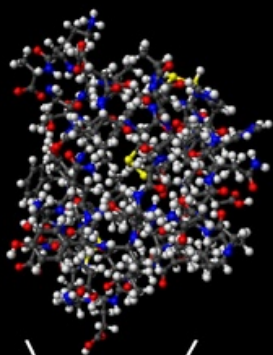


The Cell

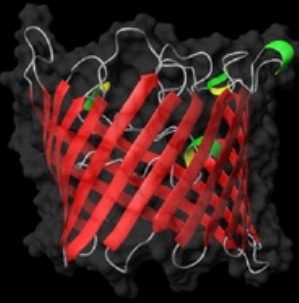
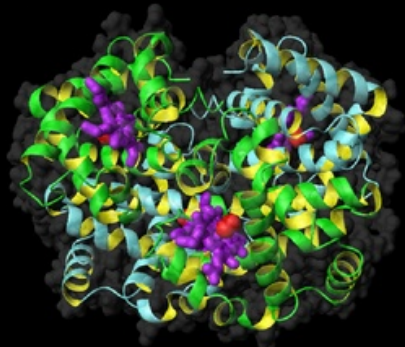
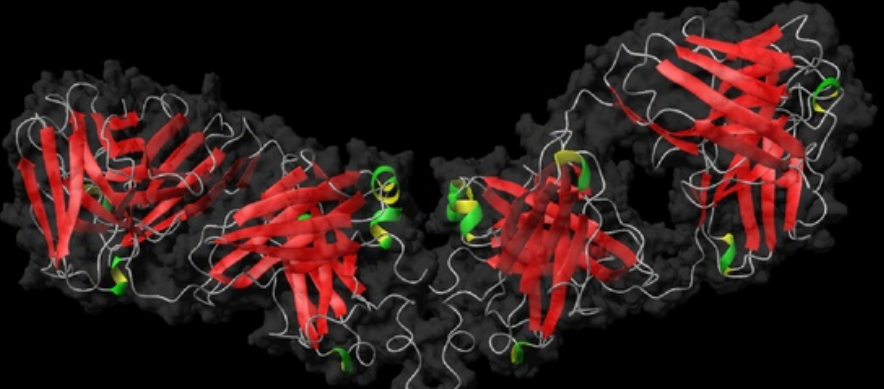
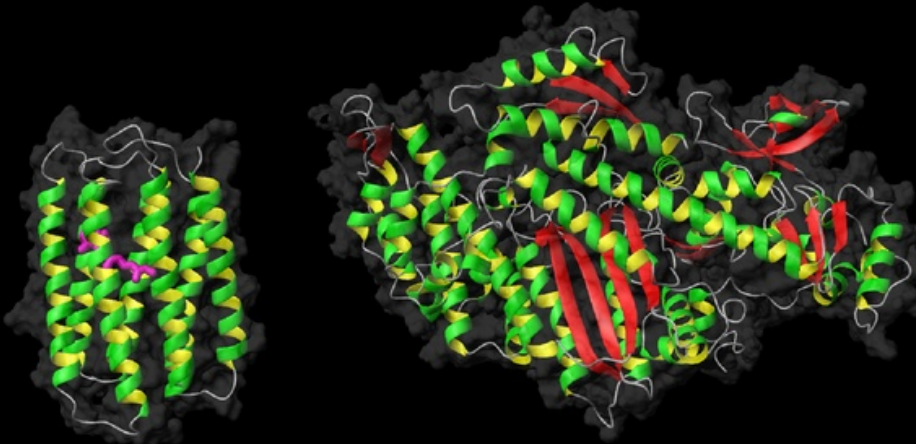


ADAM.



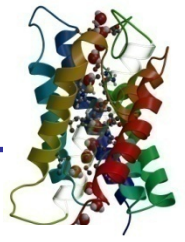


x 2



***Proteins are
nature's
nanomachines***

Aquaporin Water Channels



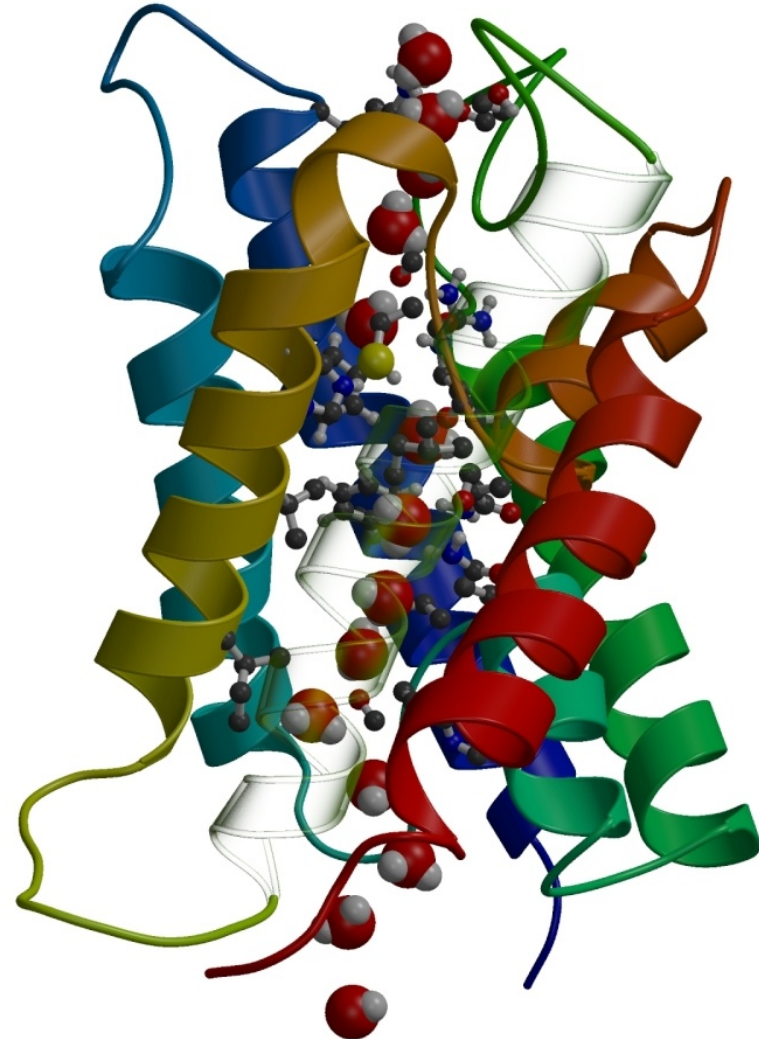
Aquaporins are highly selective, efficient water channels ($10^9/s$)

vital for water/solute balance in e.g. kidney, brain, eye

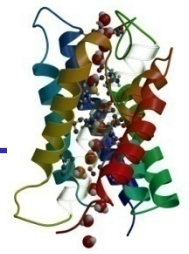
Malfunction associated with diabetes (AQP2), brain oedema (AQP4), glaucoma (AQP1), cell migration/cancer (AQP1/3).

Plasmodium falciparum expresses one AQP

- ***water permeation***
- ***permeation of other solutes?***
- ***inhibition***

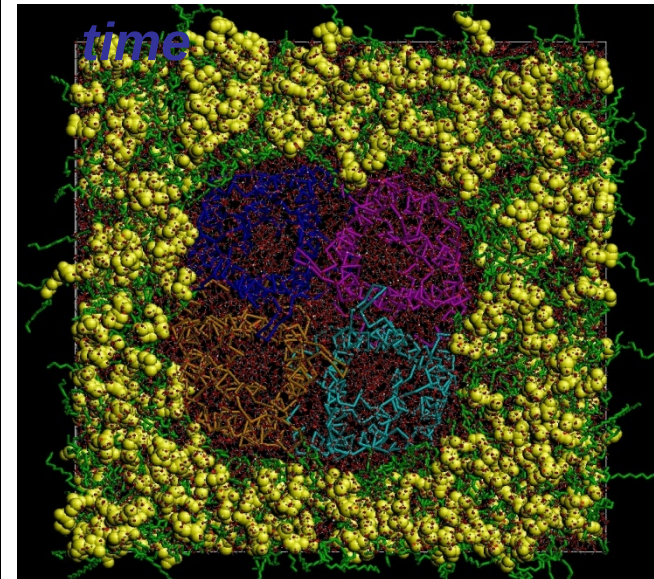
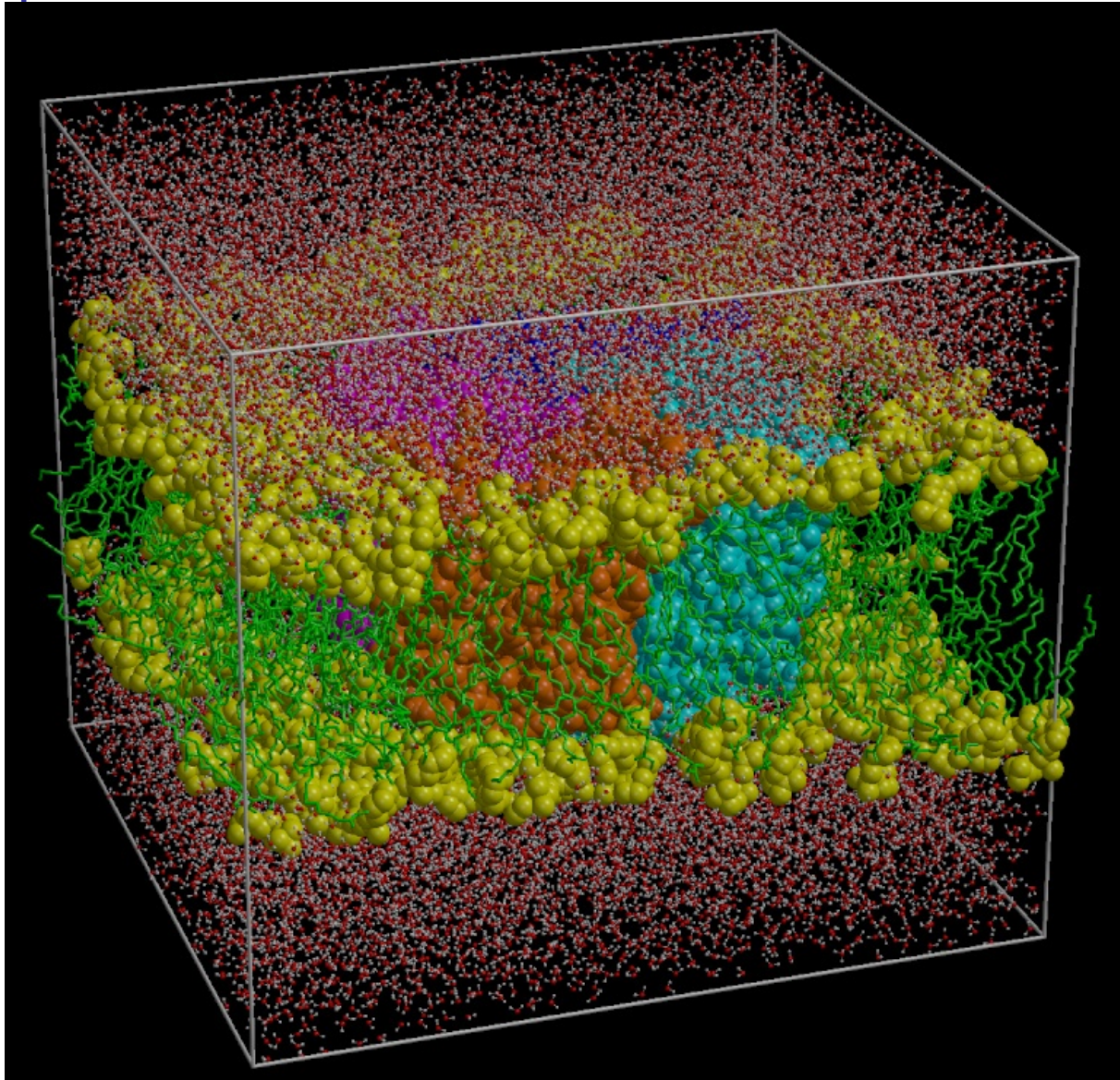


MD simulations of water permeation

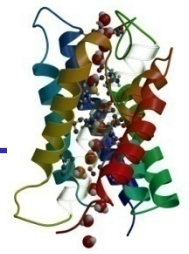


Gromacs:

- ca. 100 000 atoms
- full electrostatics, periodic boundary
- 10-100 ns simulation

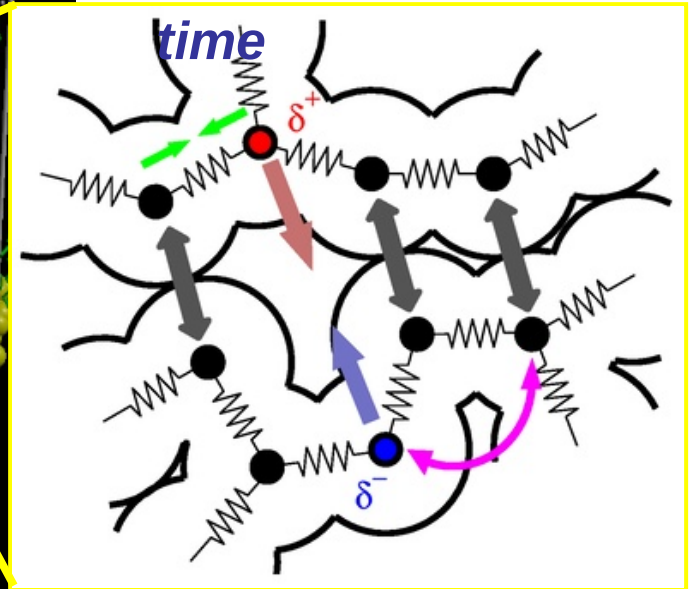
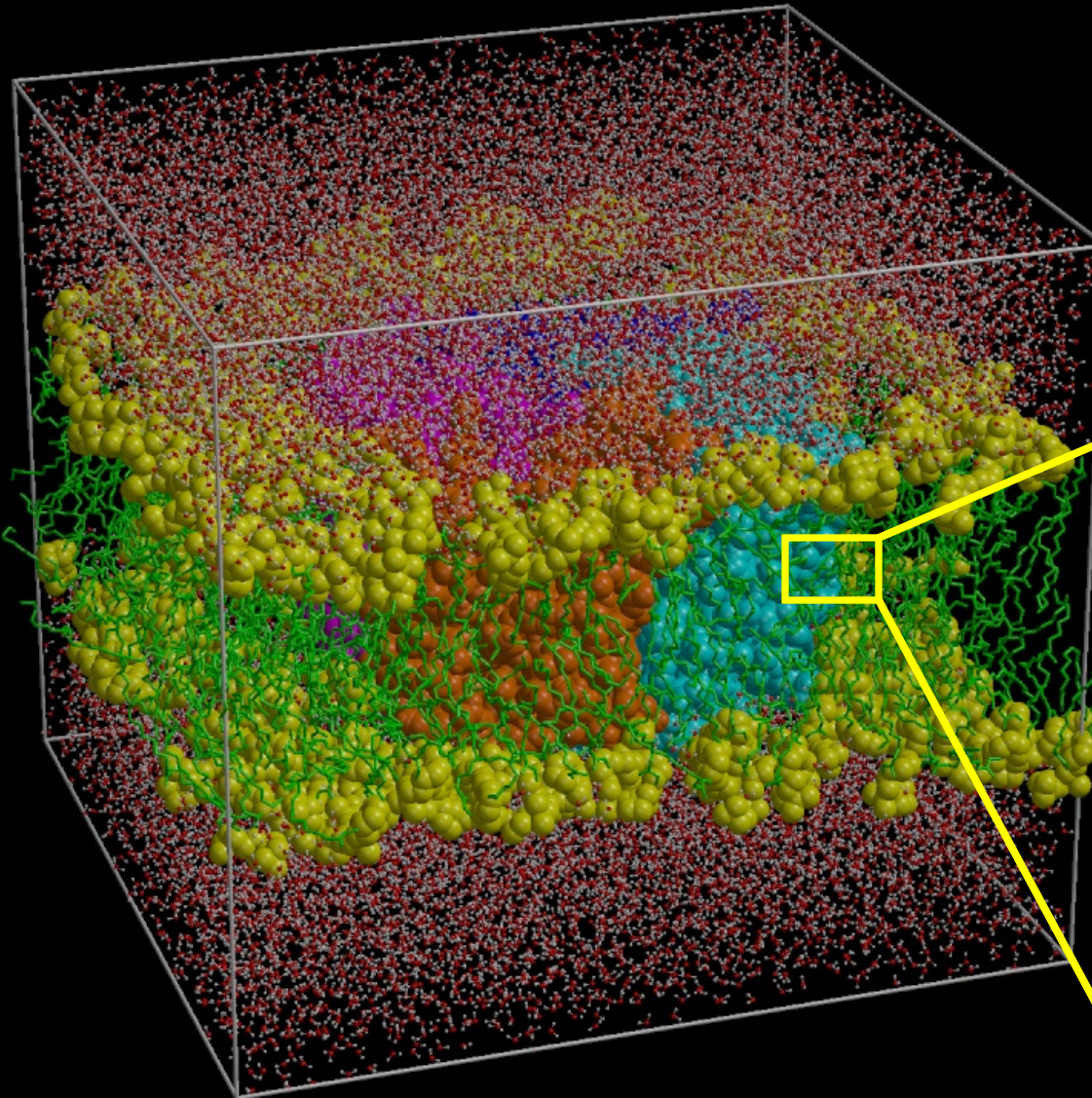


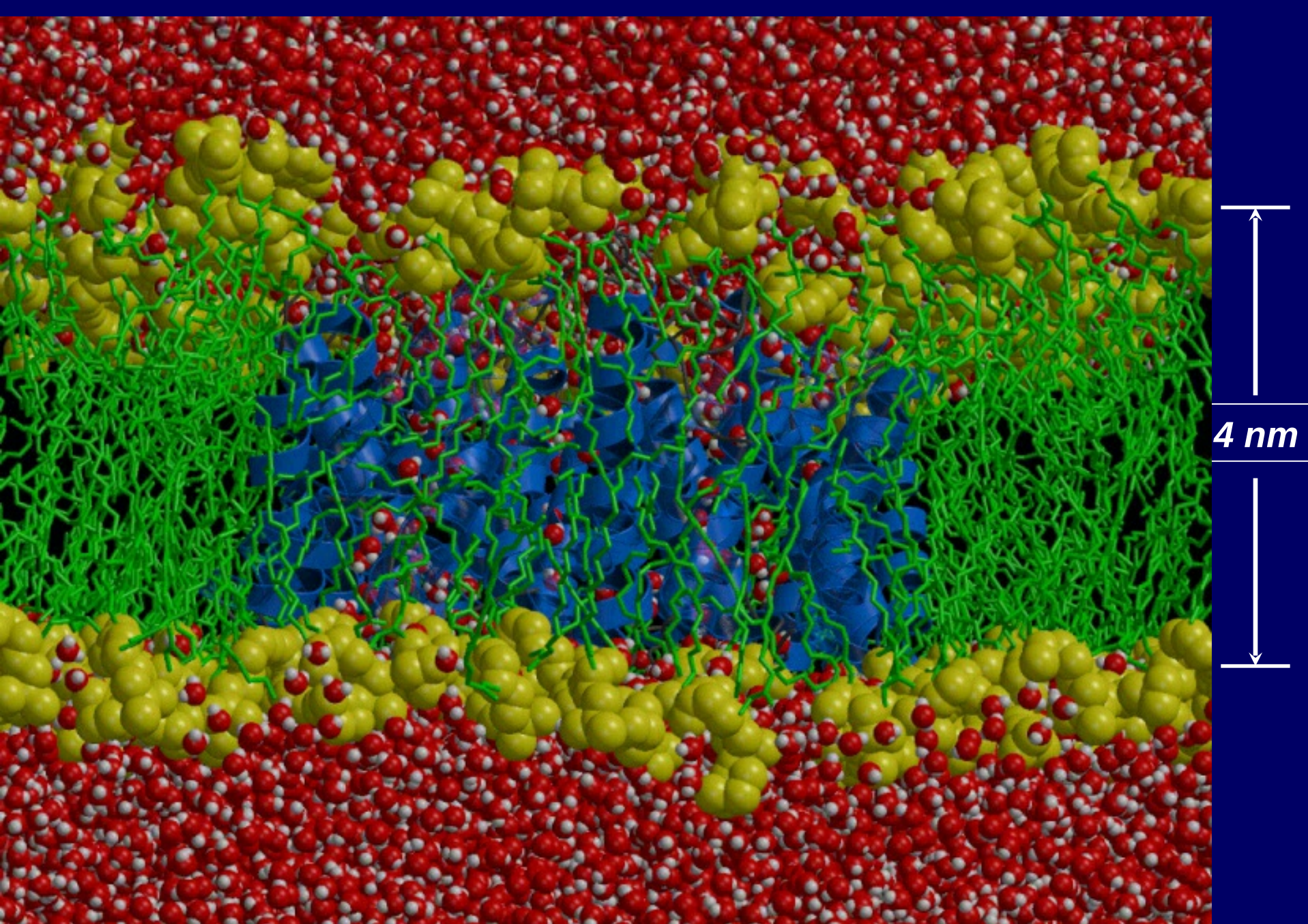
MD simulations of water permeation



Gromacs:

- ca. 100 000 atoms
- full electrostatics, periodic boundary
- 10-100 ns simulation





4 nm

Molecular dynamics simulation, 1s $\hat{=}$ $2 \cdot 10^{-11}$ s

,Real time' water permeation

***one out of several
spontaneous
permeation events***

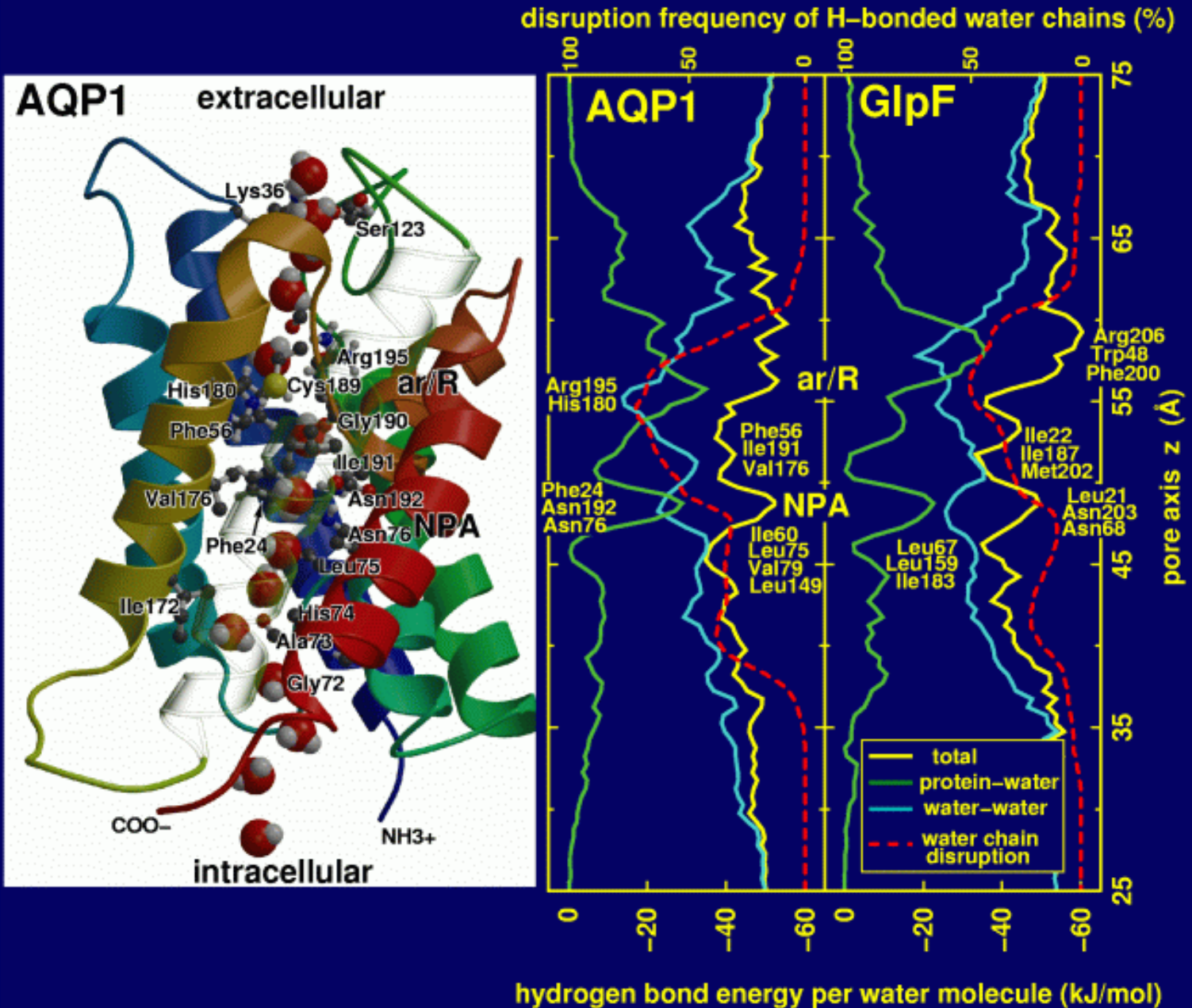
$$pf = 7.5 \cdot 10^{-14} \text{ cm}^3/\text{s}$$

$$(\text{exp: } 3.2 - 11.7 \cdot 10^{-14} \text{ cm}^3/\text{s})$$

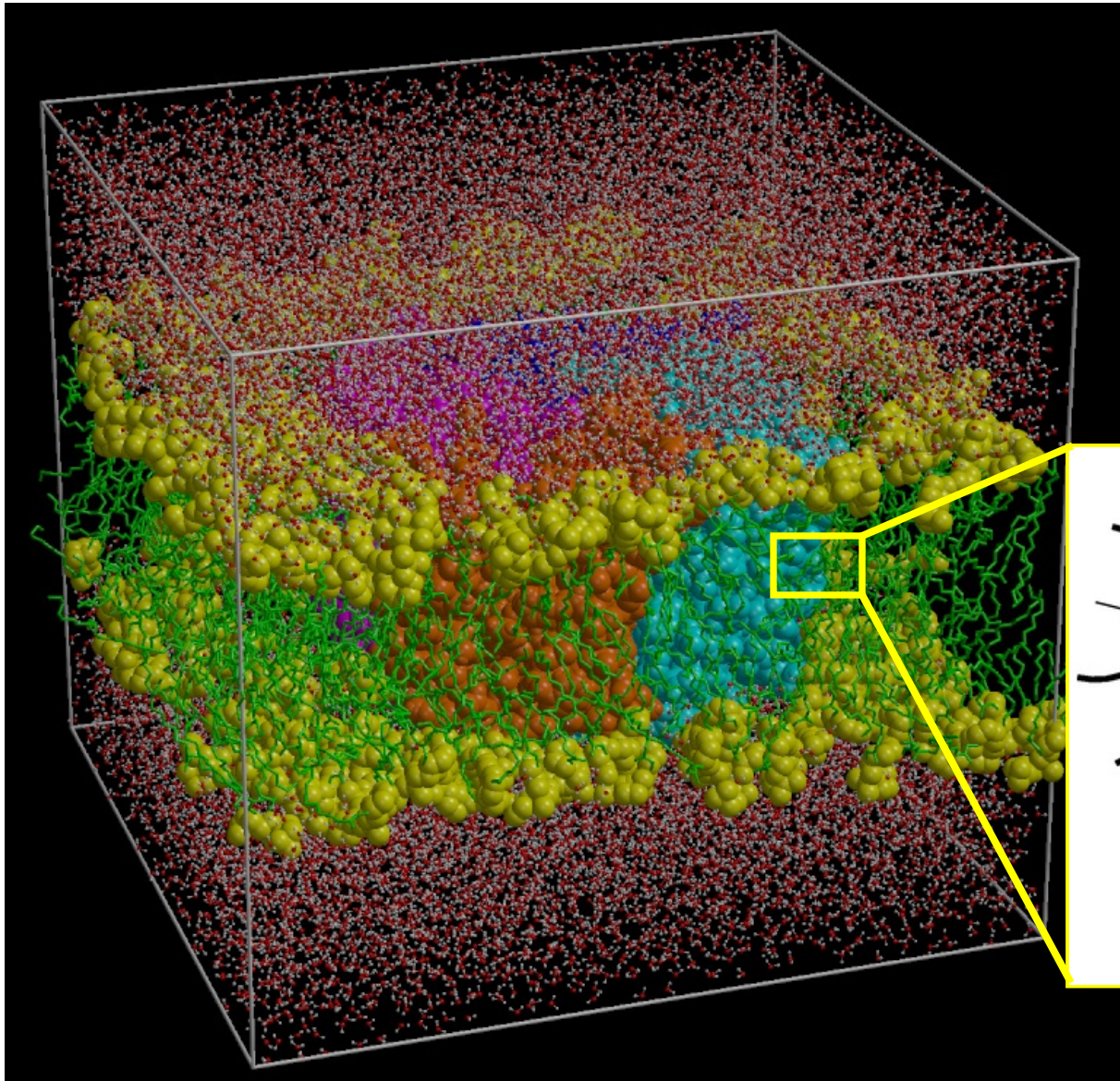
***(outside the channel,
only few water
molecules are shown)***



Water pathway and hydrogen bonding



Why are these calculations so expensive?

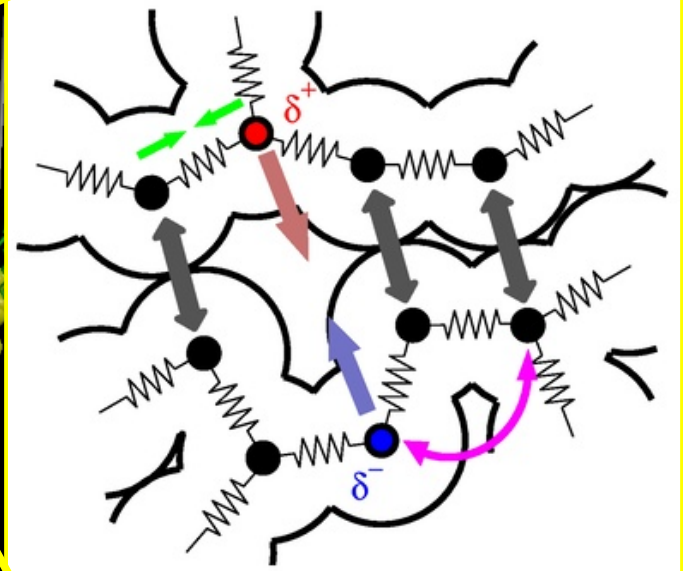


ca. 100 000 atoms

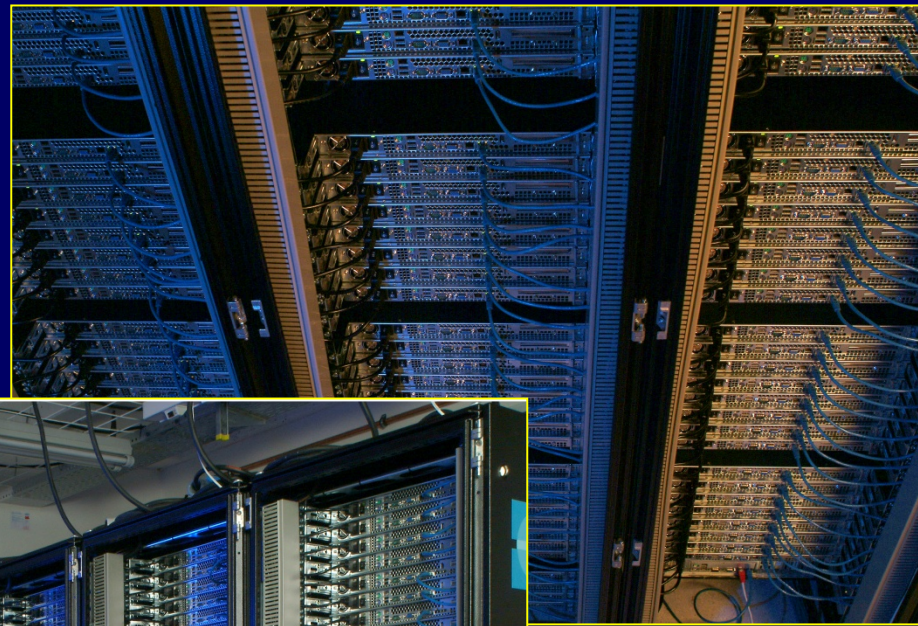
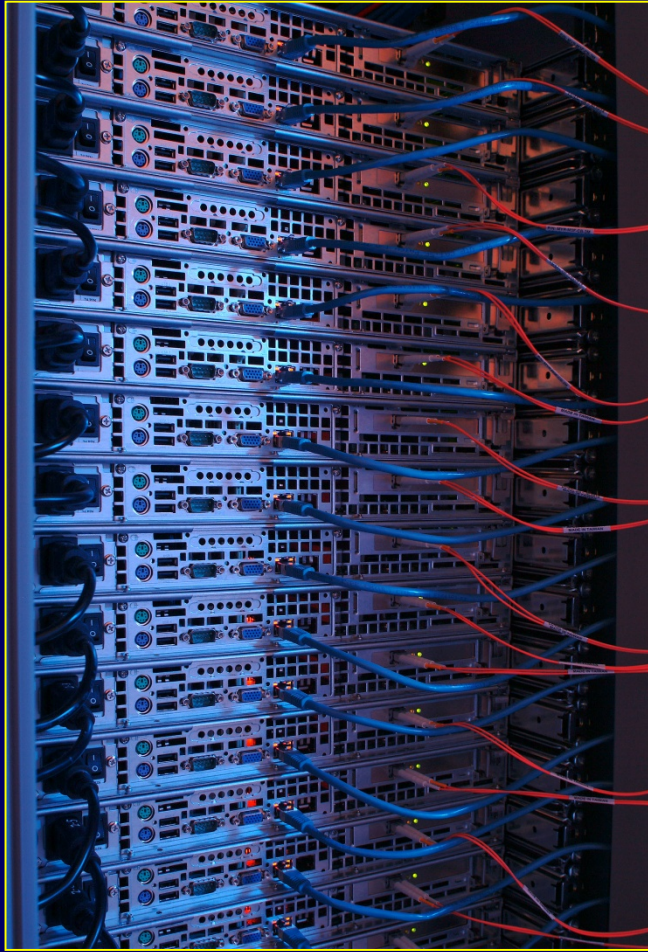
→ in principle 10^{10}
interactions

time step $\sim 10^{-15}$ s

→ 10^9 steps for $1\mu\text{s}$



High performance parallel computing



*Molecular dynamics simulations of biological processes requires huge computational resources.
Typical simulation: 1-10 exaflop*

„High performance“ parallel computing



*In-house linux cluster (~ 40 users):
~ 16000 CPU cores, 450 GTX GPUs
+ external sources, e.g. GWDG, Garching*

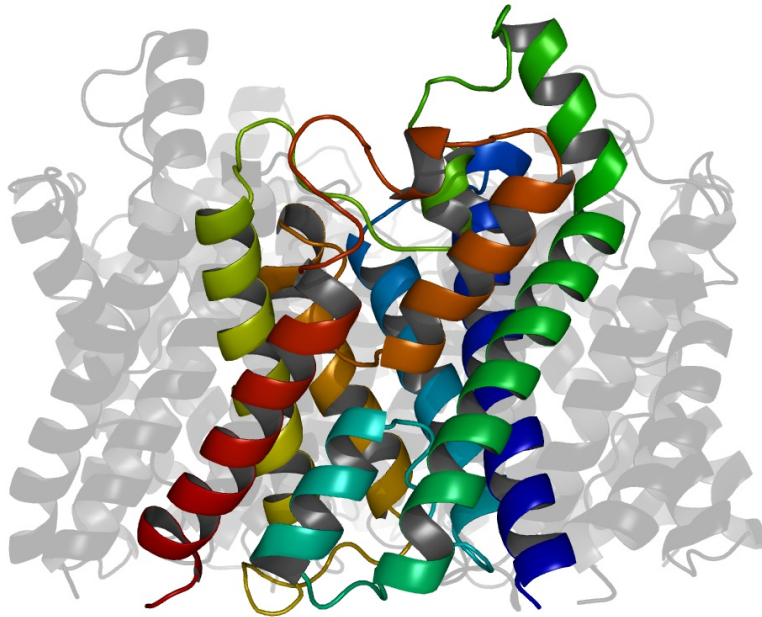


*Backup traffic:
> 10 TB / week*

*Long term storage:
> 200 TB / year*

Molecular dynamics in drug design

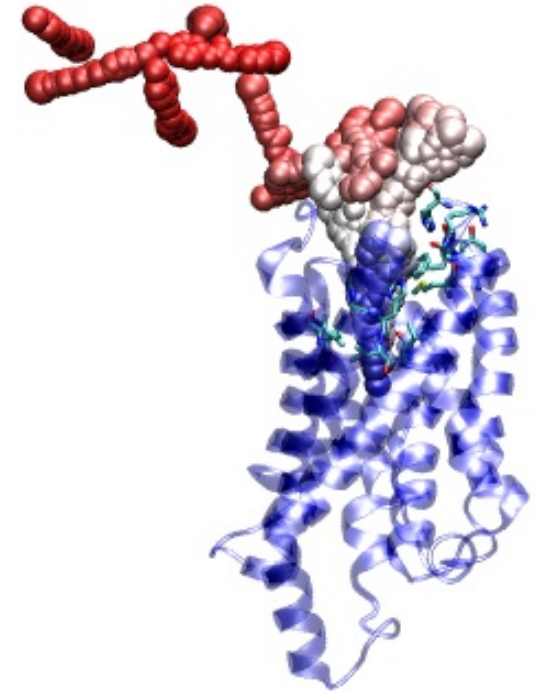
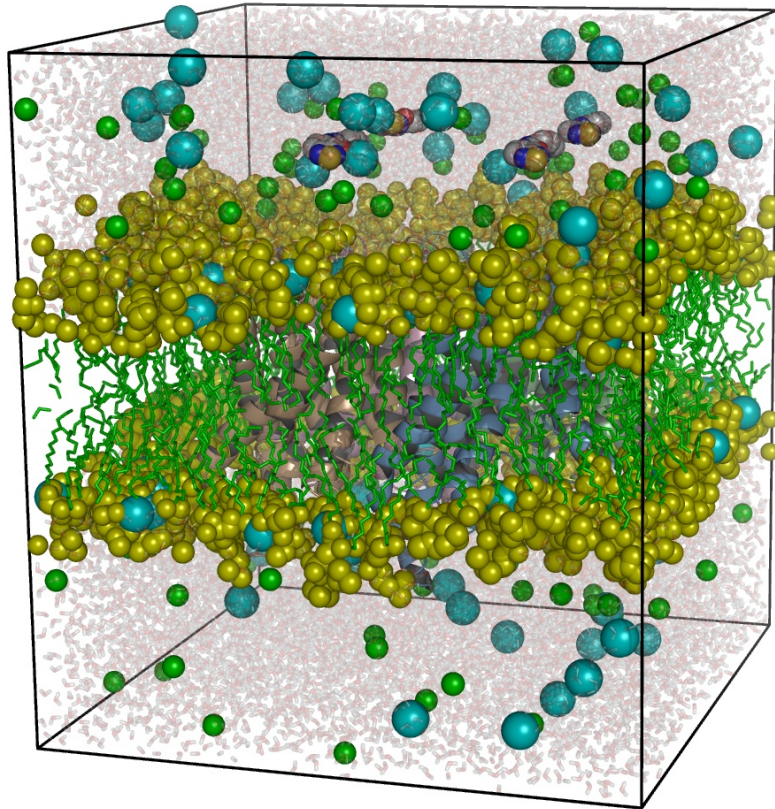
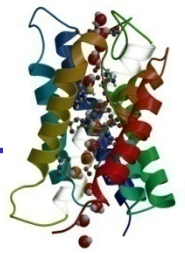
AQP9 inhibitor design



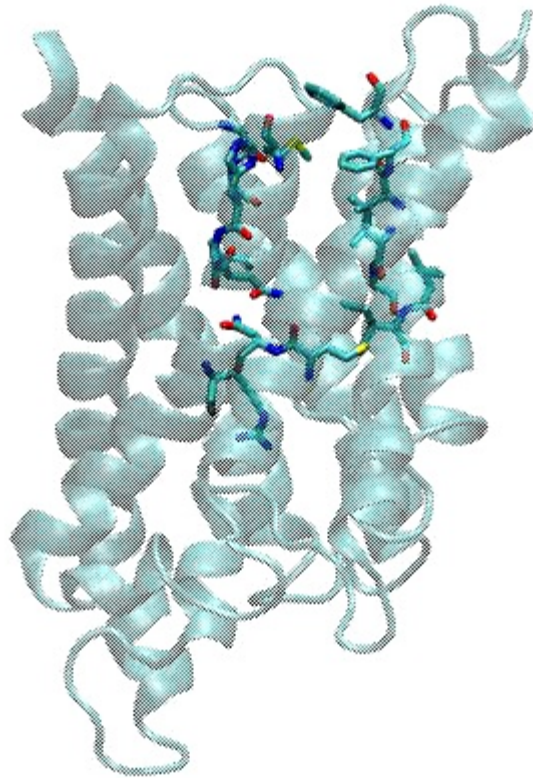
- aquaglyceroporin expressed mainly in the liver
- plays a major role in glycerol metabolism
- possible diabetes target
- structure unknown, homology model based on bacterial GlpF

Collaborations:
Michael Rützler
(Aarhus)

Simulation of ligand binding to AQP9

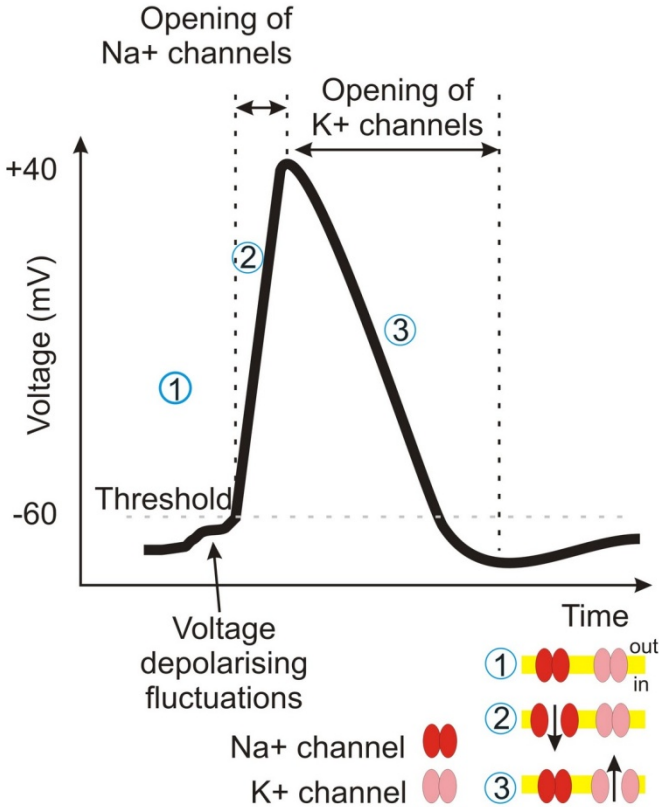
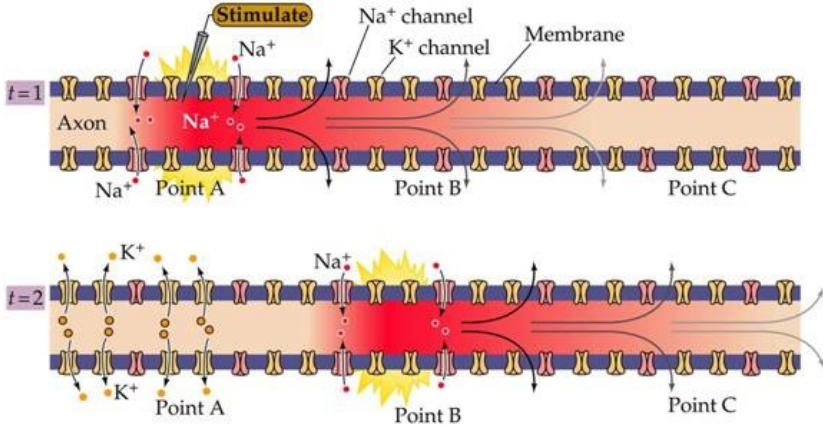
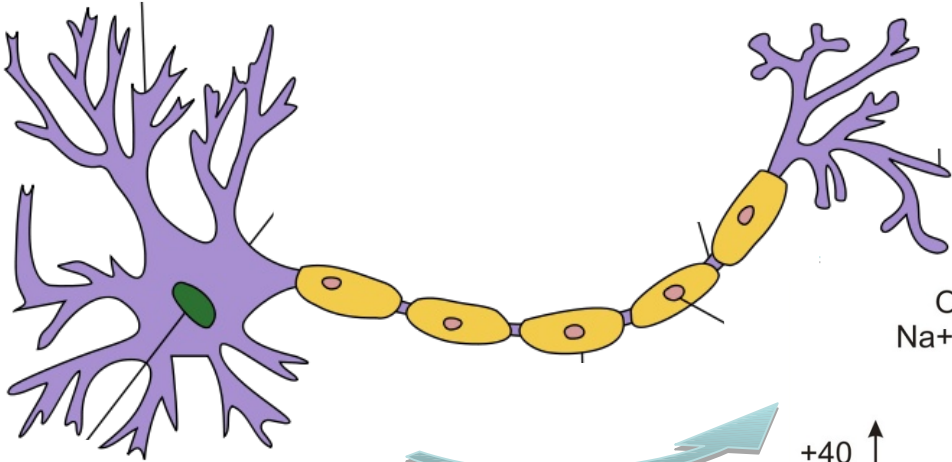
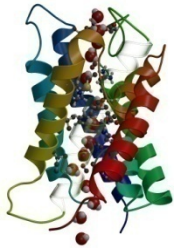


ID	EC50 [μM]	Max. inhibition [%]	
HTS1	2.7	100	4x HTS2 20 simulations à 100ns 4 intracellular associations
<u>HTS2</u>	0.4	90	1 extracellular association

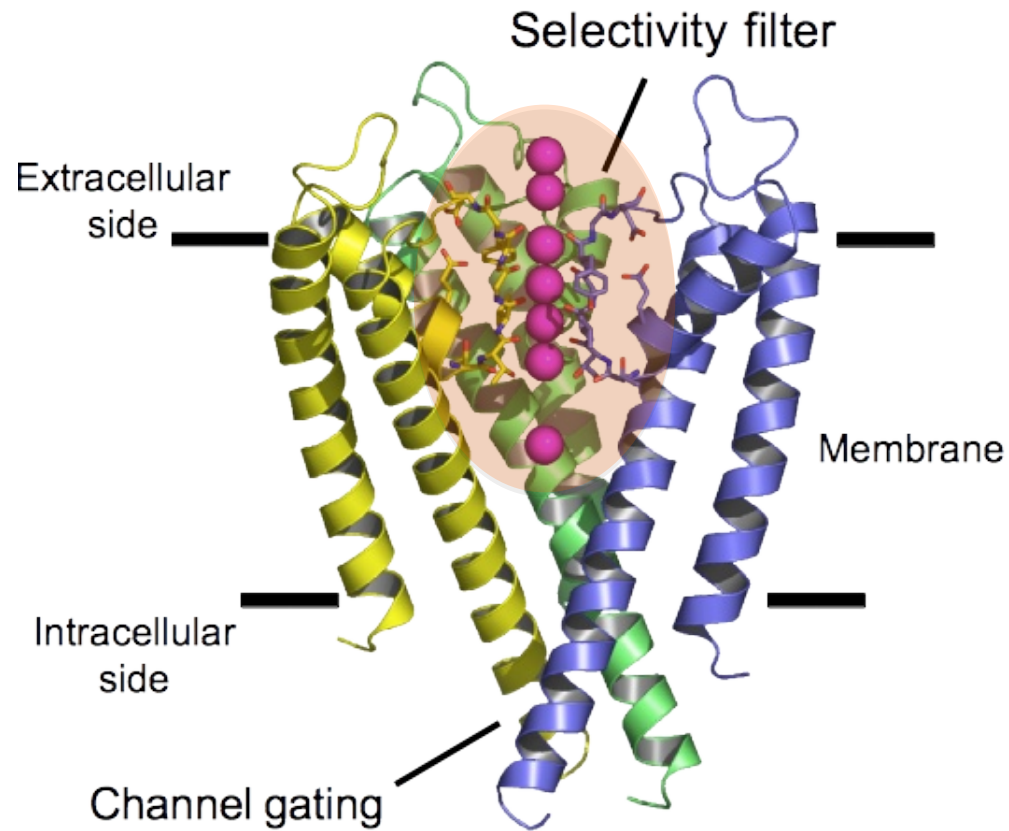


K⁺ permeation through potassium channels

Ion Channels in Excitable Cells

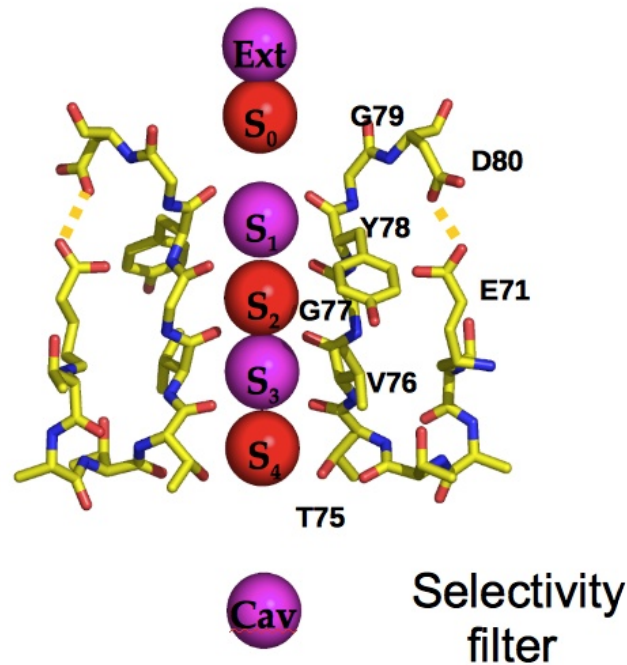


Mechanism of Selective Potassium Translocation across Membranes



Potassium channels

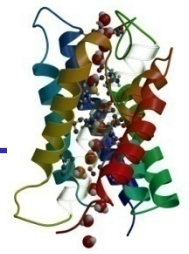
Mechanism of Selective Potassium Translocation across Membranes



Potassium channel SF:

- combines strong selectivity for K^+ with impressive efficiency
- catalyses transmembrane ion transfer to $\sim 10^8$ ions per second
- \sim diffusion limited ion conduction

Mechanism of Selective Potassium Translocation across Membranes



Chemistry of ion coordination and hydration revealed by a K^+ channel–Fab complex at 2.0 Å resolution

Yufeng Zhou, João H. Morais-Cabral*, Amelia Kaufman & Roderick MacKinnon

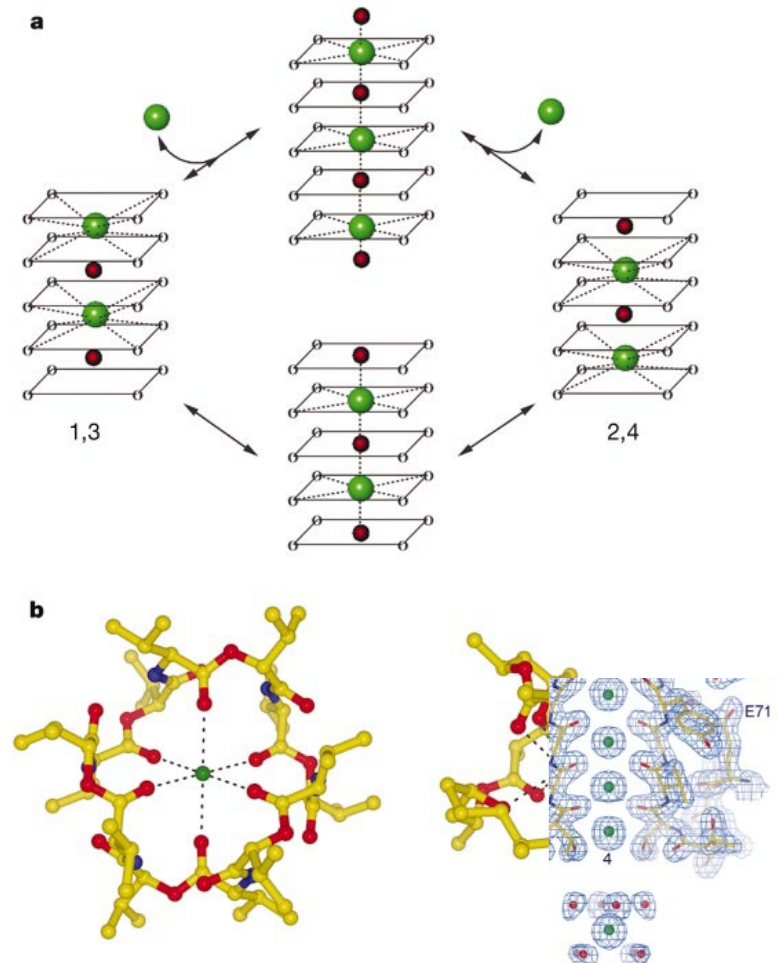
Howard Hughes Medical Institute, Laboratory of Molecular Neurobiology and Biophysics, Rockefeller University, 1230 York Avenue, New York, New York 10021, USA

Energetic optimization of ion conduction rate by the K^+ selectivity filter

João H. Morais-Cabral*, Yufeng Zhou & Roderick MacKinnon

Howard Hughes Medical Institute, Laboratory of Molecular Neurobiology and Biophysics, Rockefeller University, 1230 York Avenue, New York, New York 10021, USA

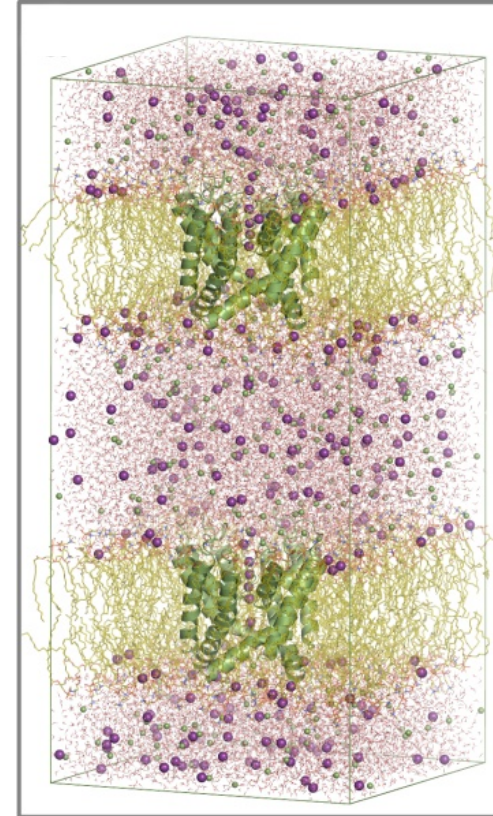
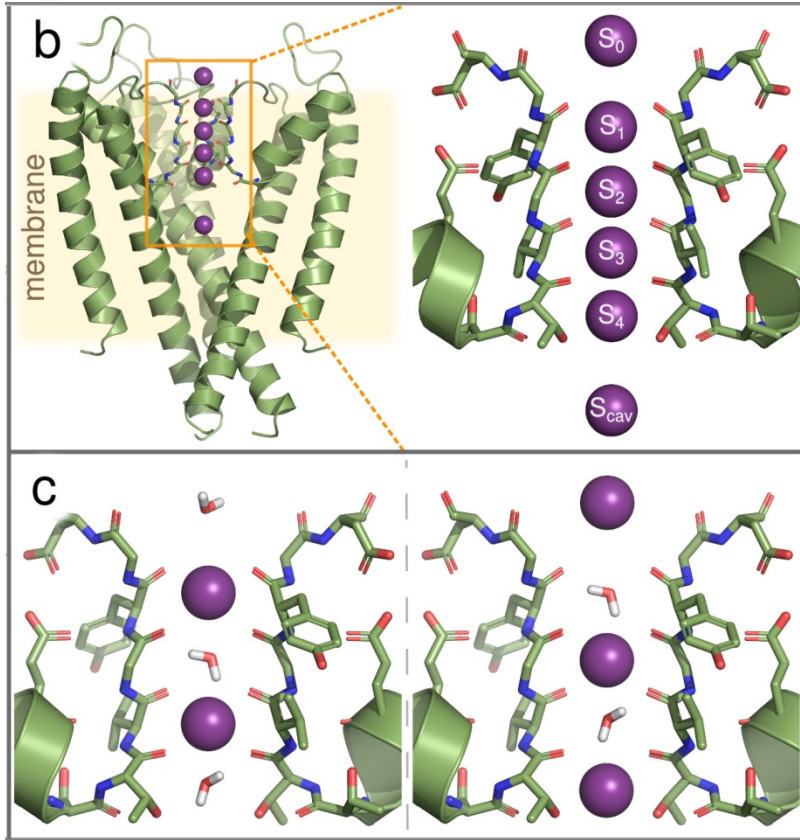
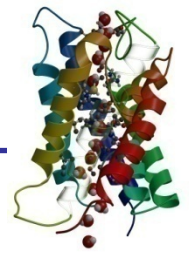
Adjacent peaks in the K^+ electron density profile are separated by about 3.2 Å (Fig. 3b). Potassium ions have a diameter of 2.7 Å, so they could, in principle, fit in the filter side by side, but this would seem to be an unstable binding configuration for electrostatic reasons. A survey of a database of small-molecule structure (Cambridge Crystallographic Data Centre, <http://www.ccdc.cam.ac.uk>) showed us that two K^+ ions only very rarely occur with a separation distance of less than 3.5 Å. Therefore, although K^+ ions



Nature, 2001; R. MacKinnon Nobel lecture, 2003

prokaryotic K^+ channel KcsA

Molecular mechanism of Potassium Permeation

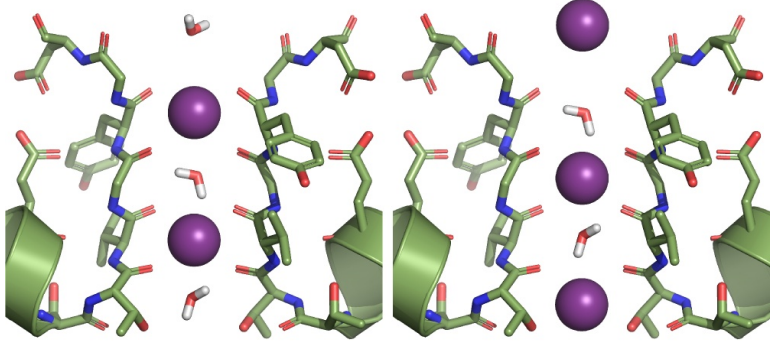


Open state structures of KcsA have recently become available

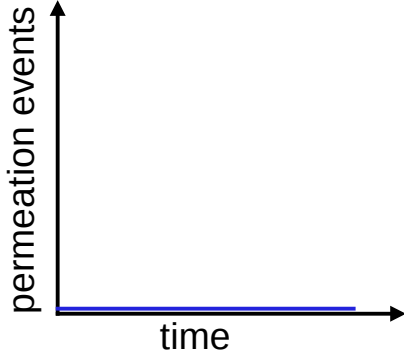
Computational electrophysiology
Molecular Dynamics



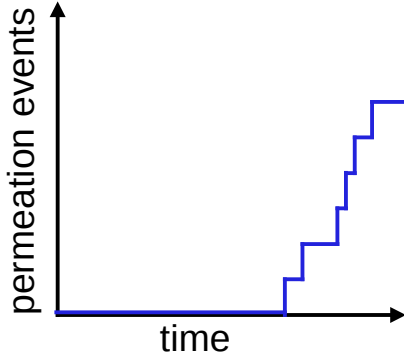
How does the Selectivity Filter Conduct Ions?



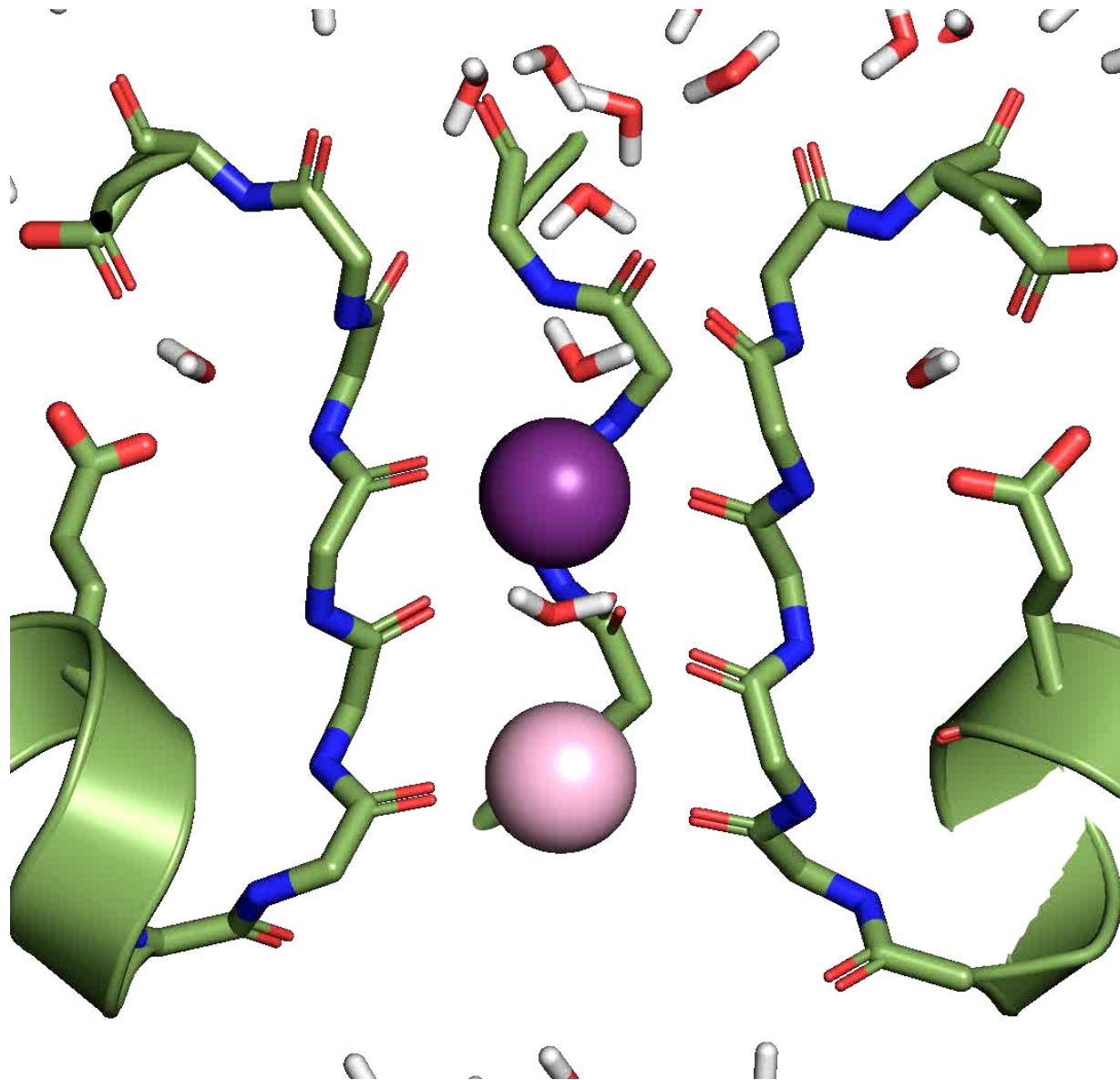
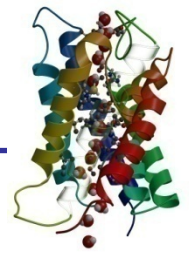
Simulation
most of the time



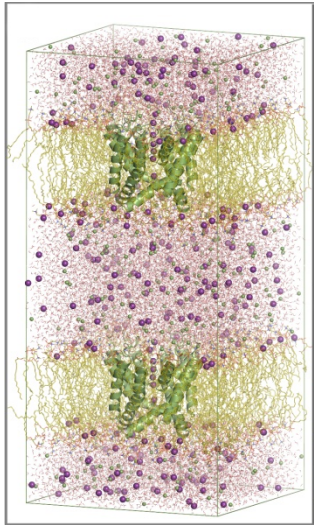
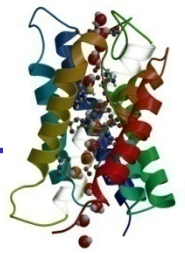
rarely



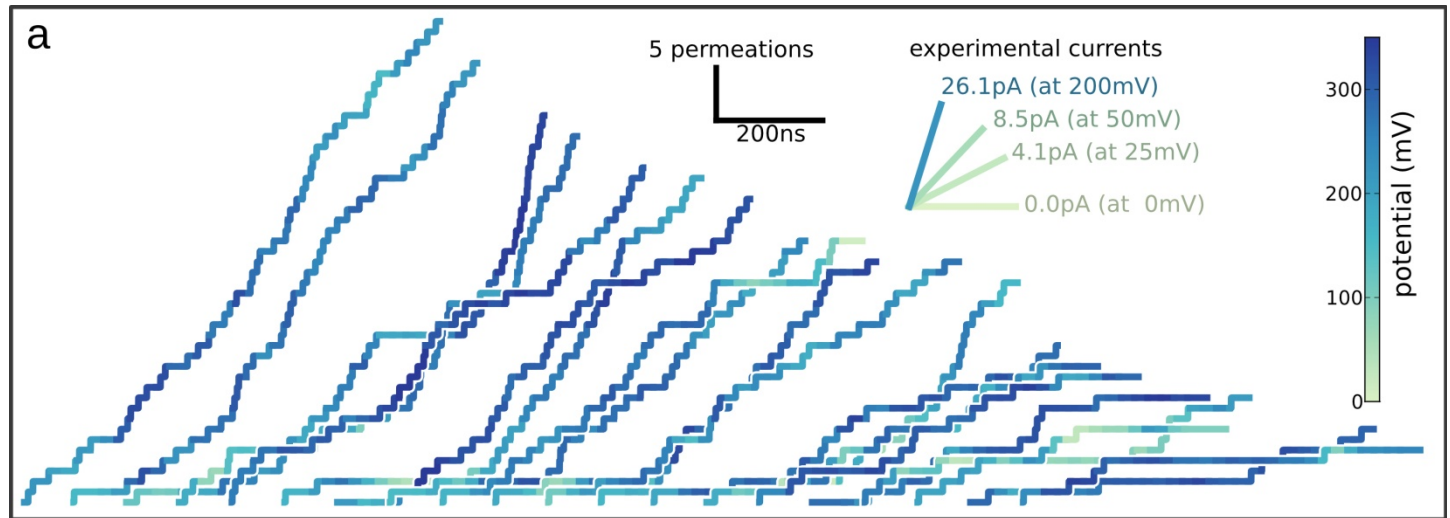
Potassium Permeation Through Open State KcsA



Potassium Permeation Through Open State KcsA



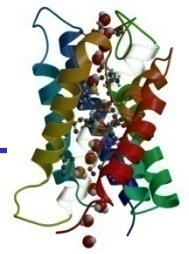
Ion current elicited by K^+ gradient



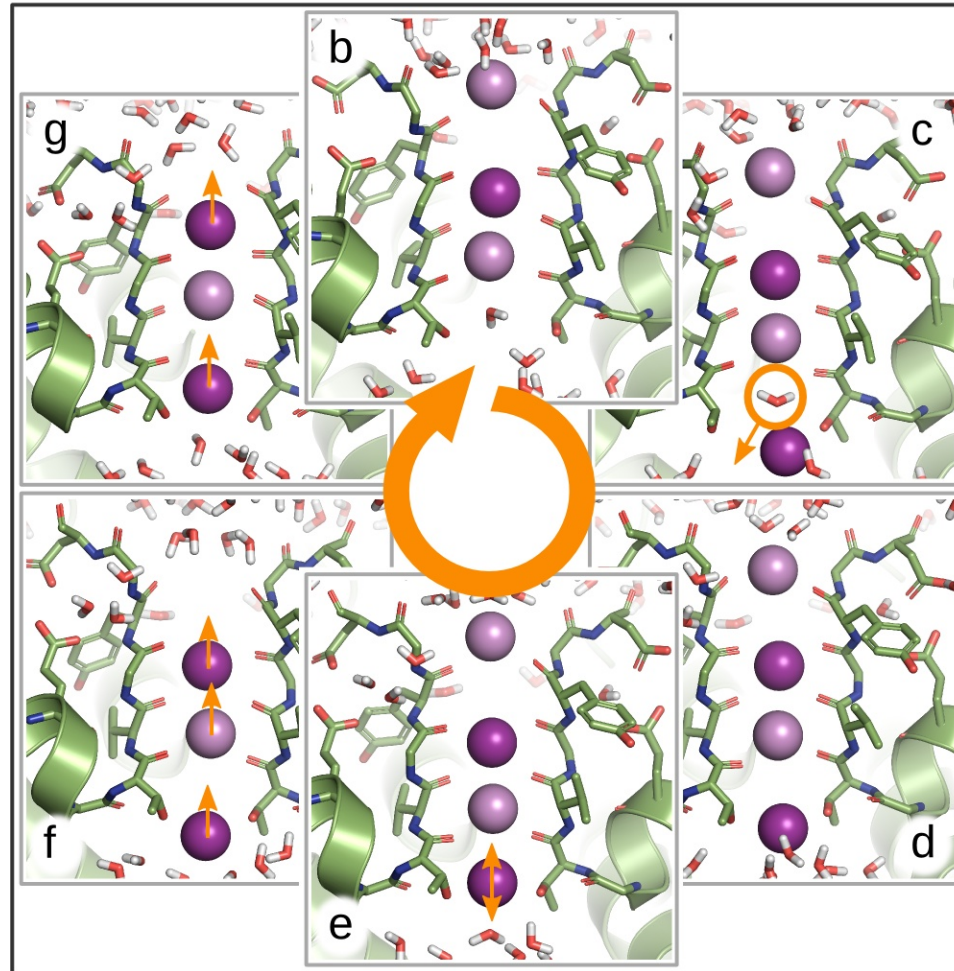
Accumulated ion permeation events on a microsecond timescale (>1500 individual permeation events)

Conductance agrees with experiment within experimental error

Mechanism of Diffusion-controlled Potassium Permeation

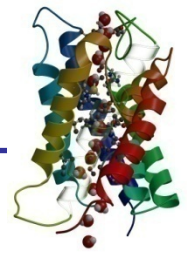


Detailed analysis of ion conduction mechanism



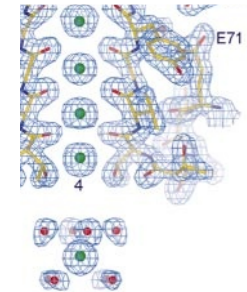
Same mechanism is seen in MthK and Kv1.2 channels

Experimental Insights



Crystallography: Tim Gruene/George Sheldrick, Göttingen

		KcsA, PDB ID: 1r3j refinement of Tl ⁺			MthK, PDB ID: 3ldc refinement of K ⁺	
		res. id	abs. occ.	rel. occ.	res. id	abs. occ.
Bind ing site	S ₁	C401	1.02 ± 0.04	1.0	A1	0.92 ± 0.07
	S ₂	C402	0.93 ± 0.03	0.9	A2	0.80 ± 0.07
	S ₃	C403	0.92 ± 0.04	0.9	A3	1.00 ± 0.09
	S ₄	C404	0.99 ± 0.04	1.0	A4	1.00 ± 0.09

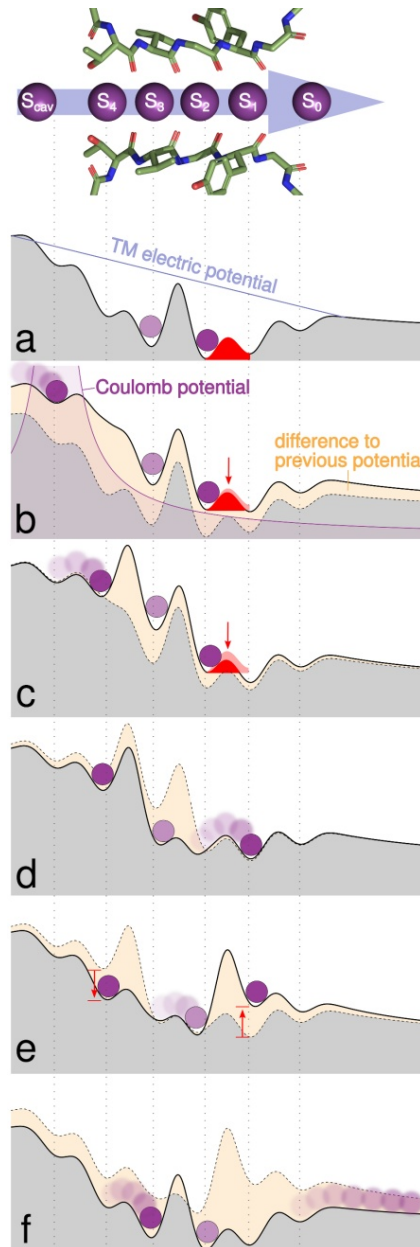
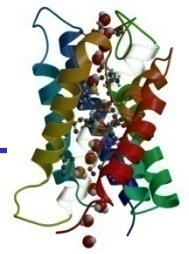


Occupancies refined against anomalous data

PDB IDs 1R3J (KcsA, Tl⁺), 3LDC (MthK, K⁺), 2QKS (Kir3.1, K⁺) :

Tl⁺, K⁺: 4x ~full occupancy

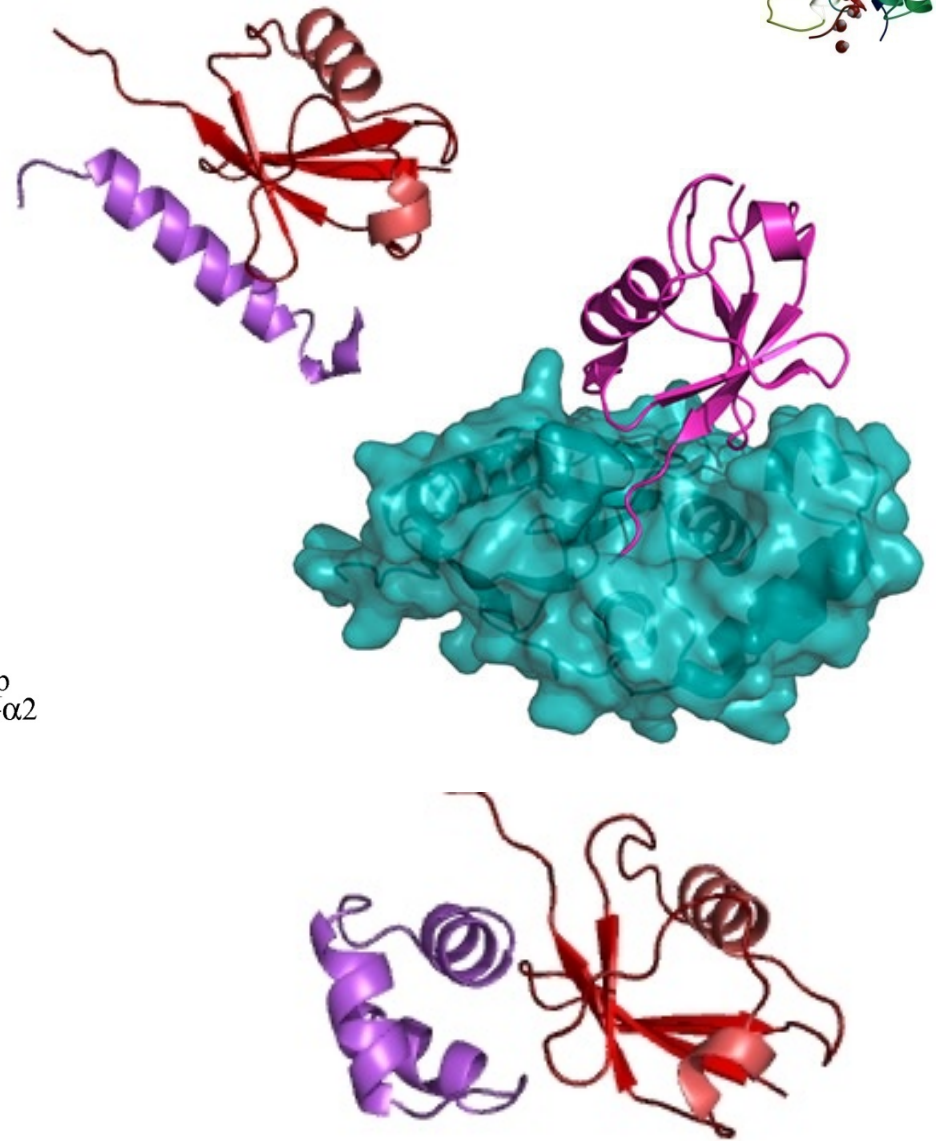
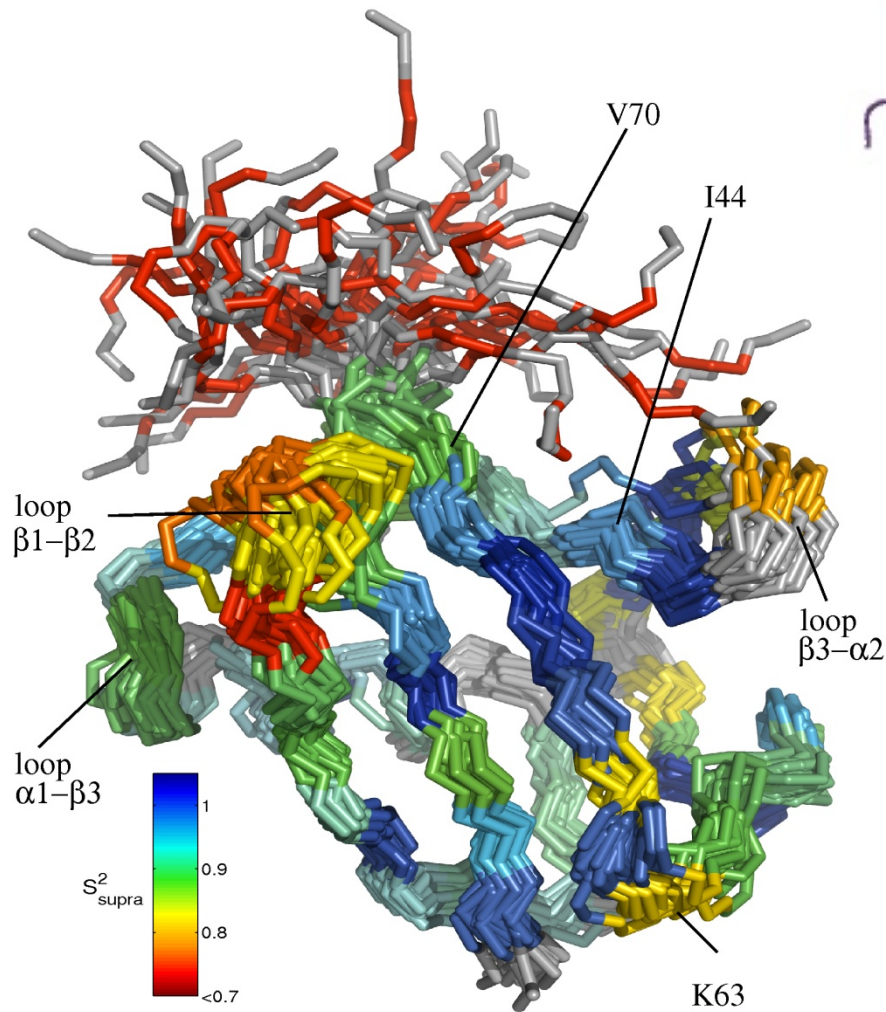
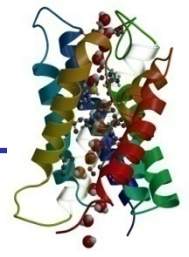
Mechanism of Selective Potassium Translocation across Membranes



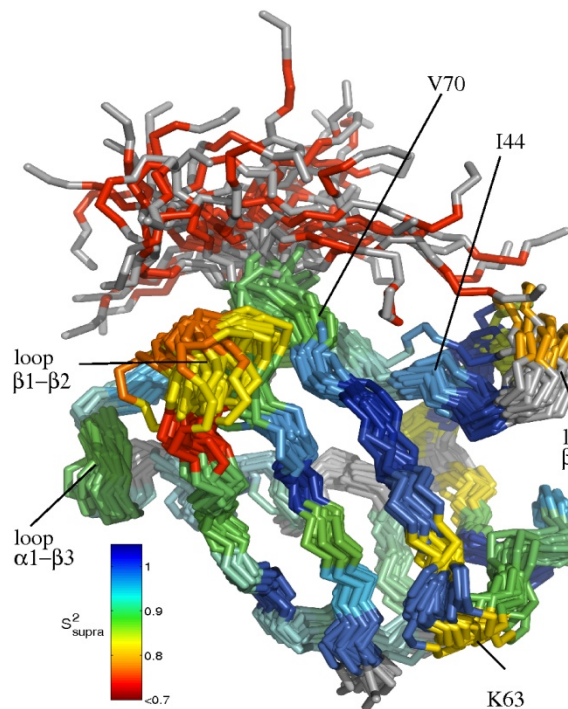
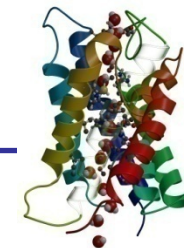
Molecular dynamics in protein recognition and design

Molecular dynamics in protein recognition and design: The case of ubiquitin

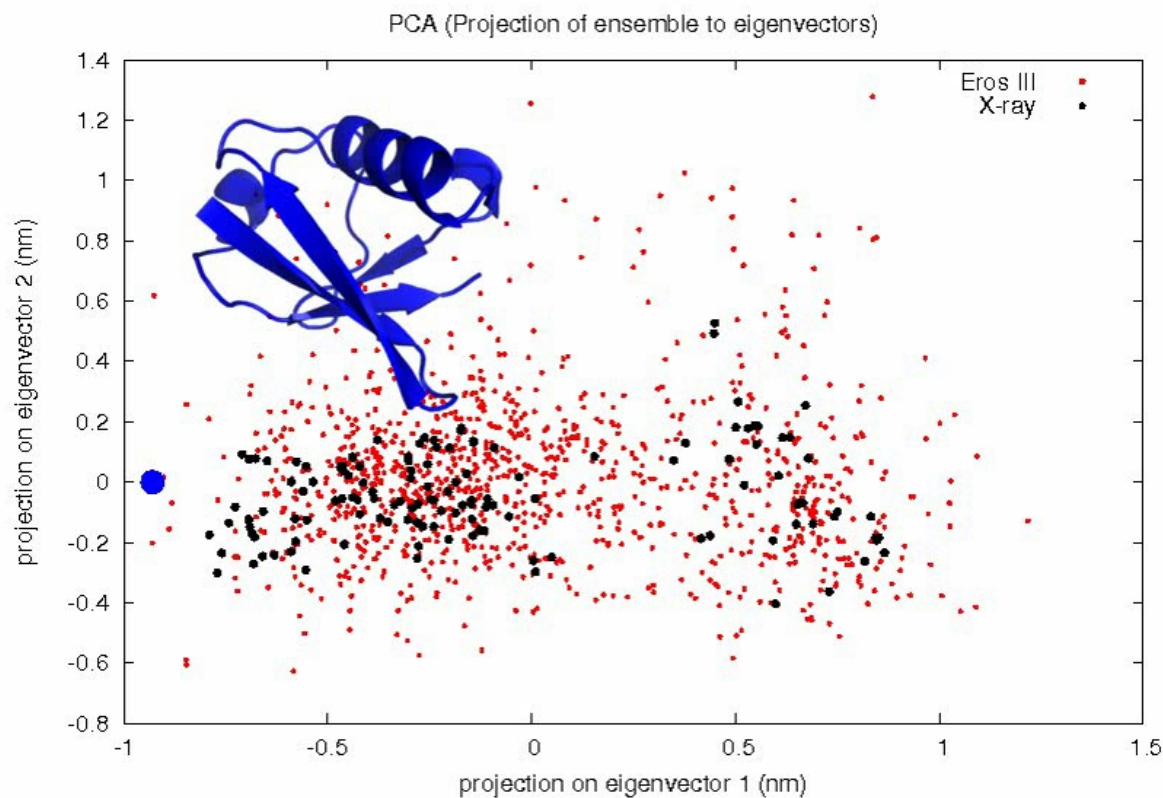
Ubiquitin binds to many partners in different conformations

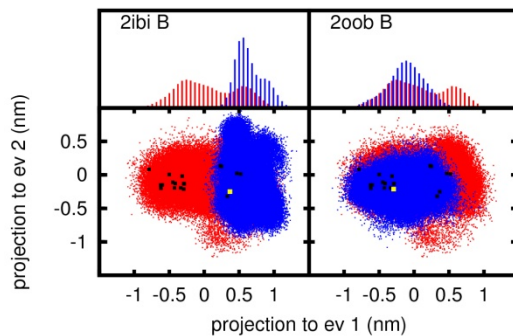
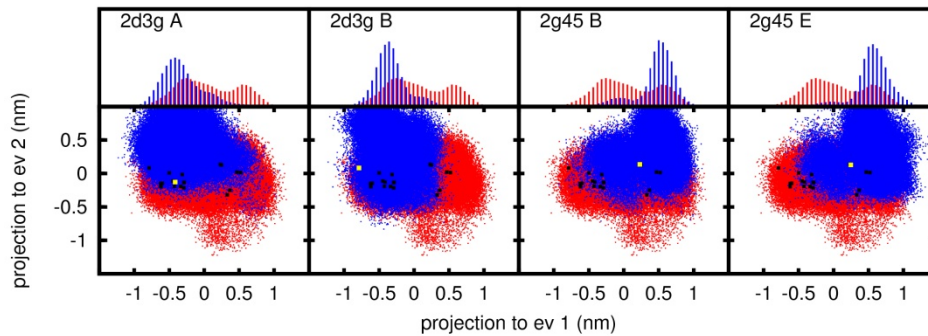
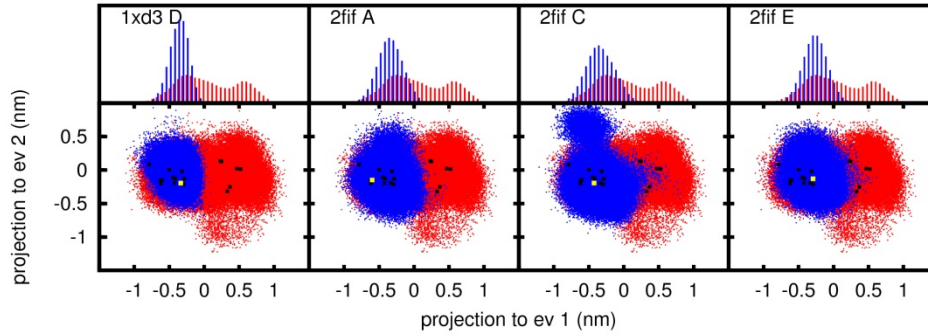
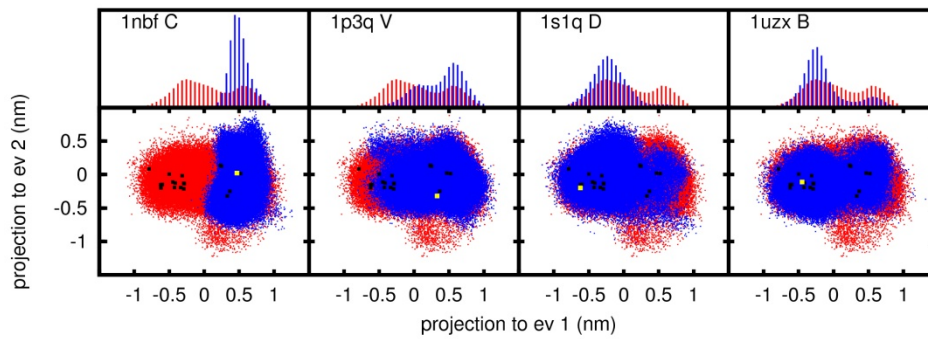


Ubiquitin complexed and free in solution



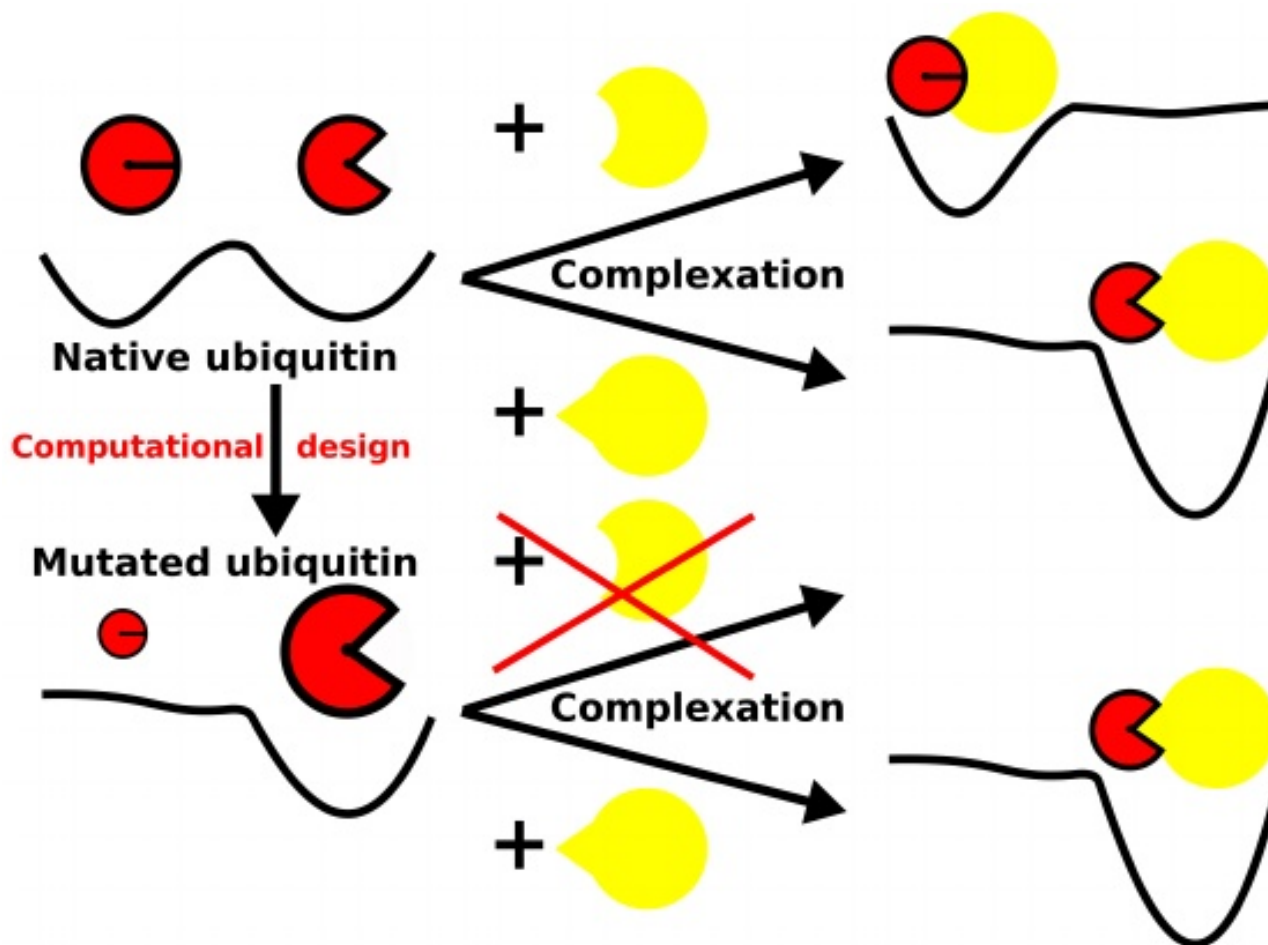
Overlap of bound and unbound ubiquitin ensembles suggests conformational selection



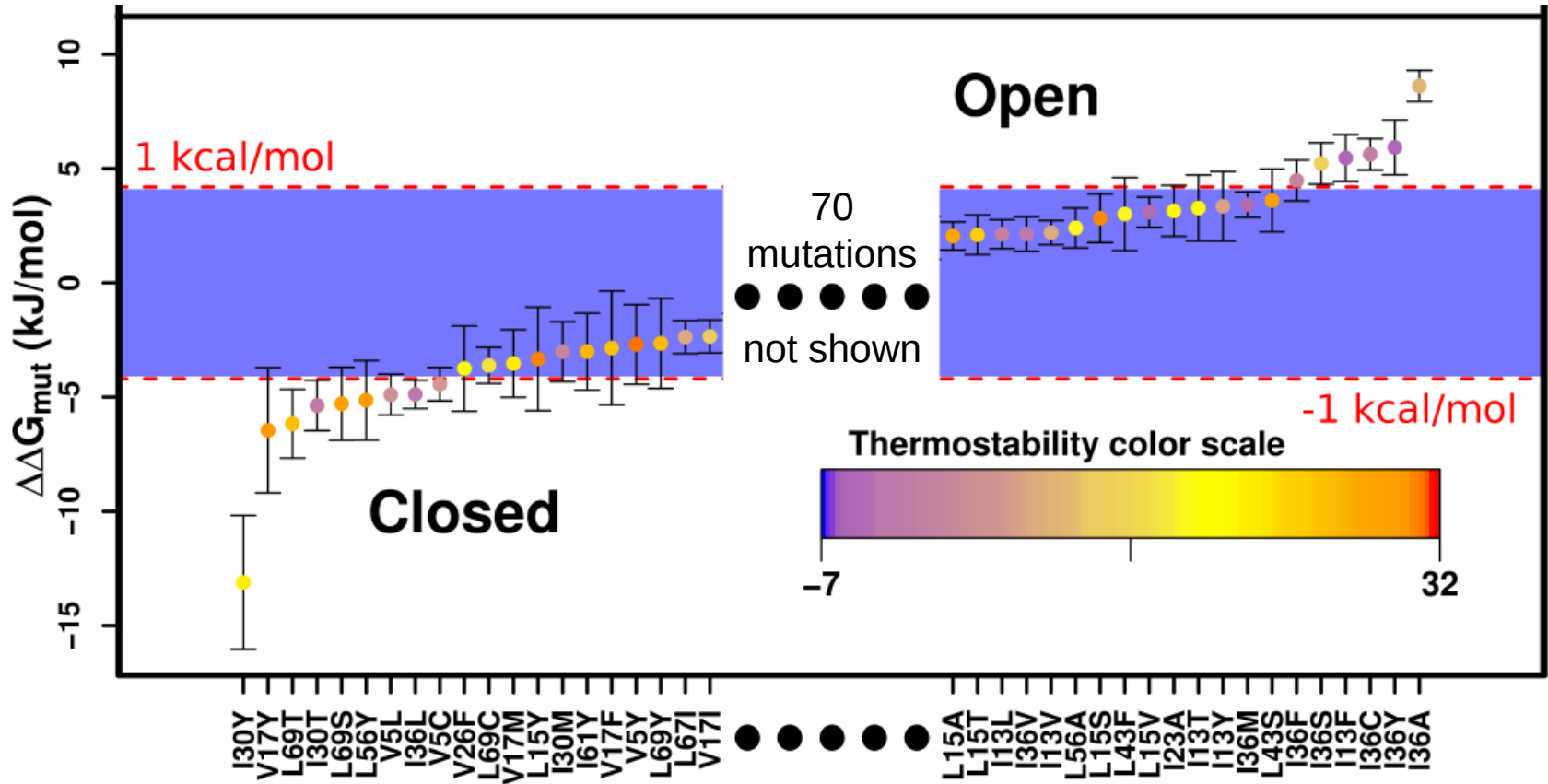
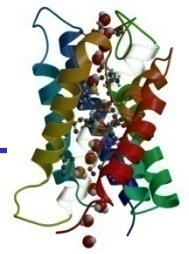


PCA: ubiquitin **free** vs **complexed**

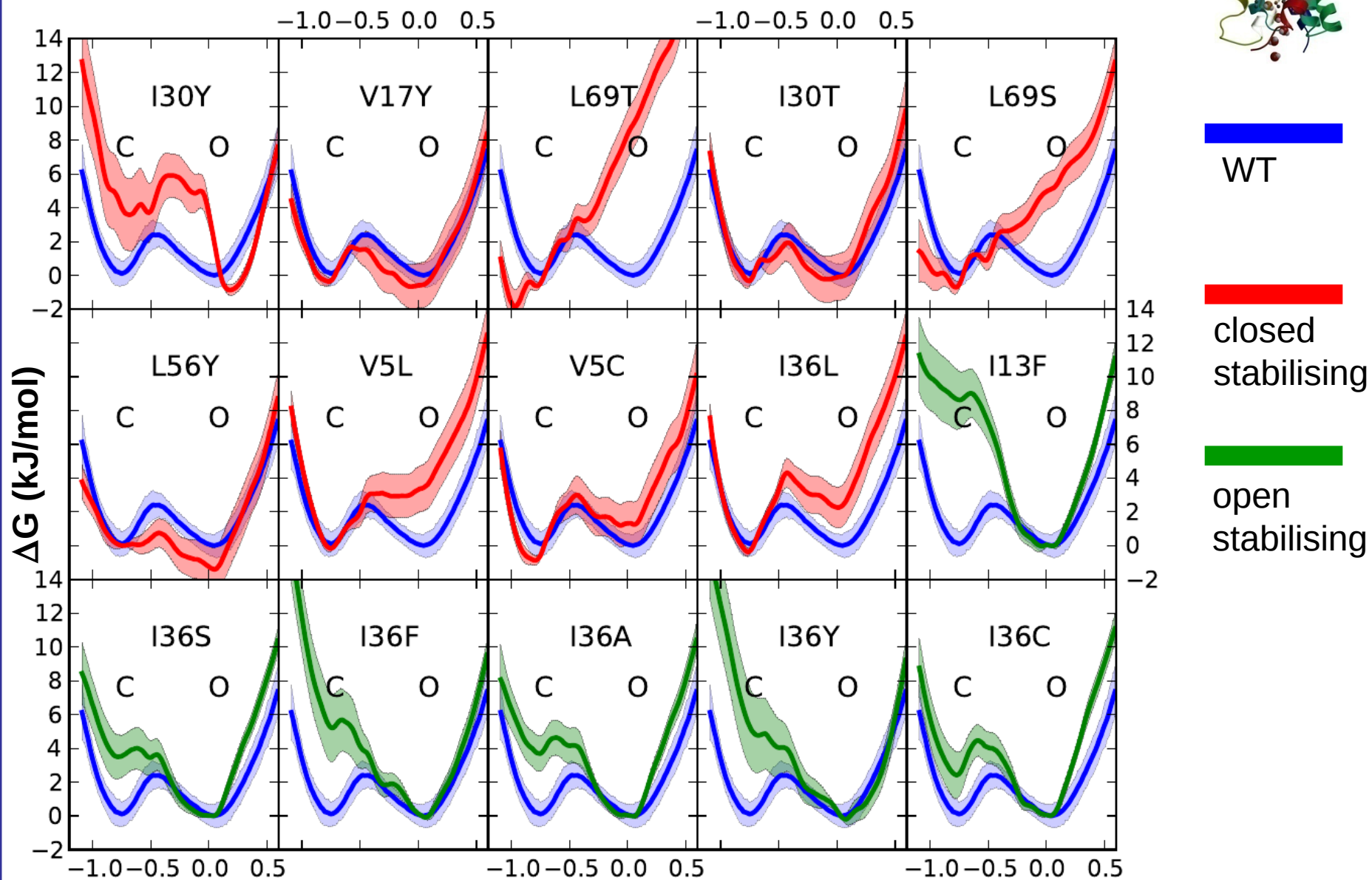
Ubiquitin mutants with altered affinity



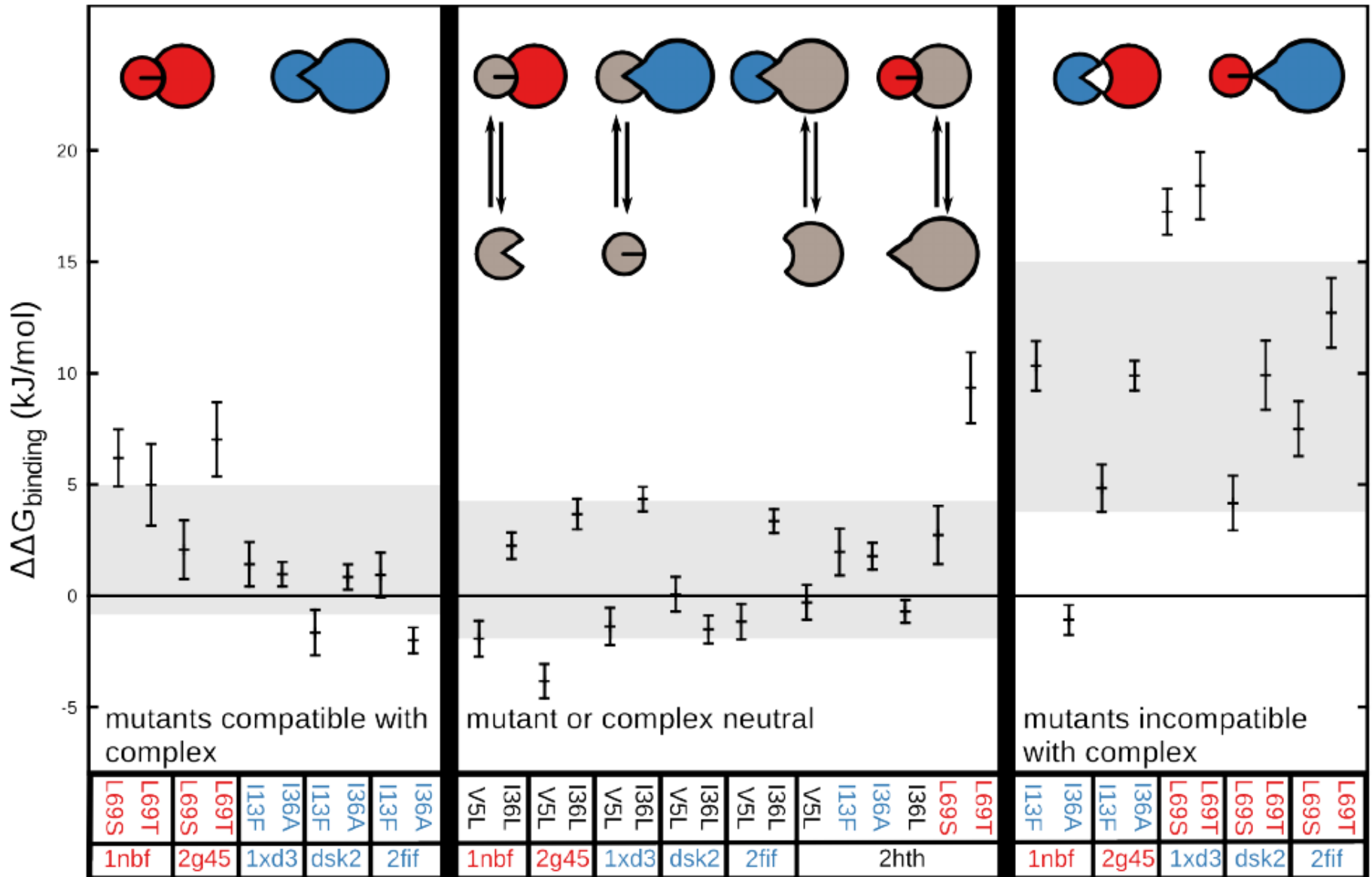
Alchemical screening of 112 mutants



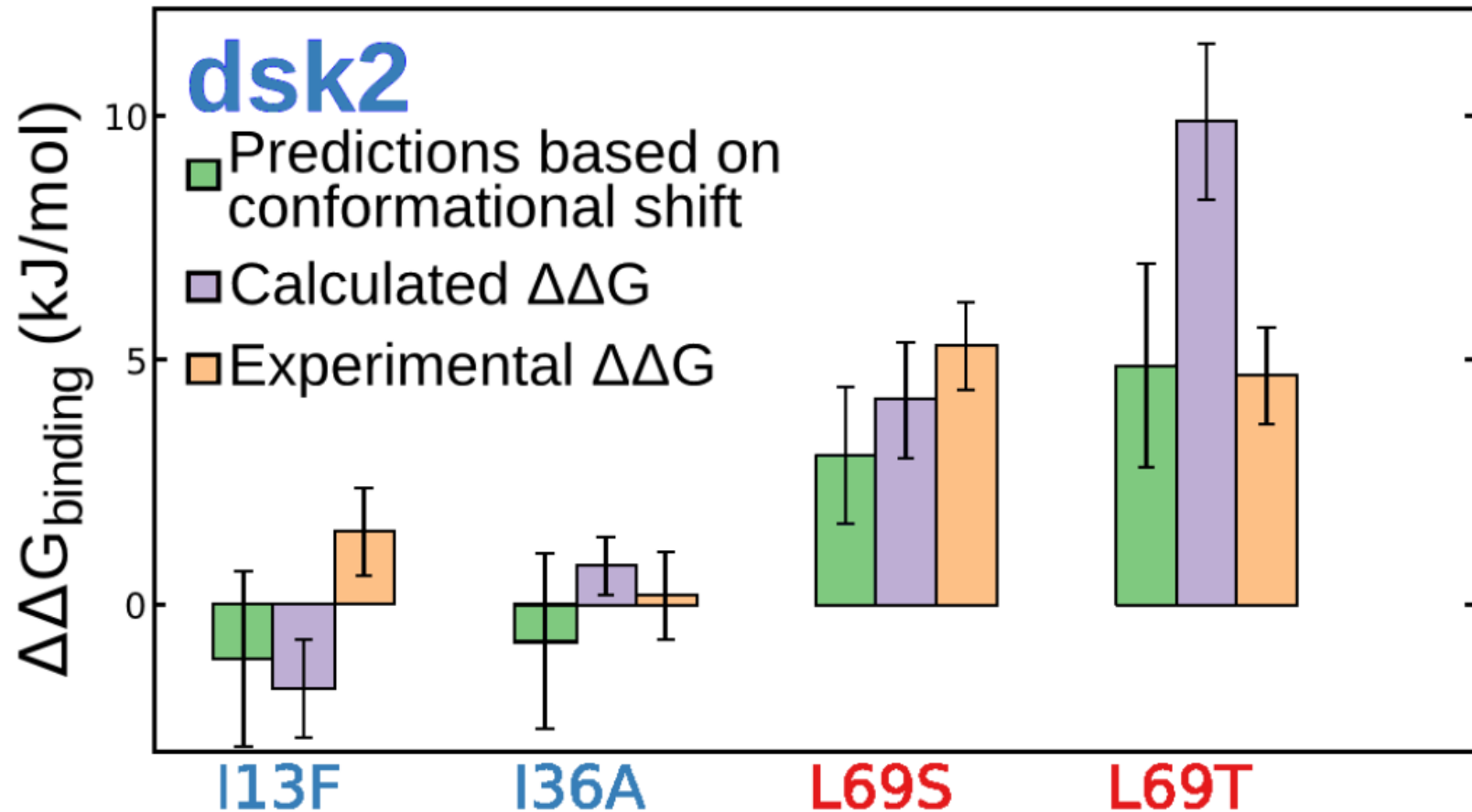
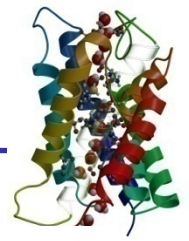
Validation of ubiquitin mutants by umbrella sampling



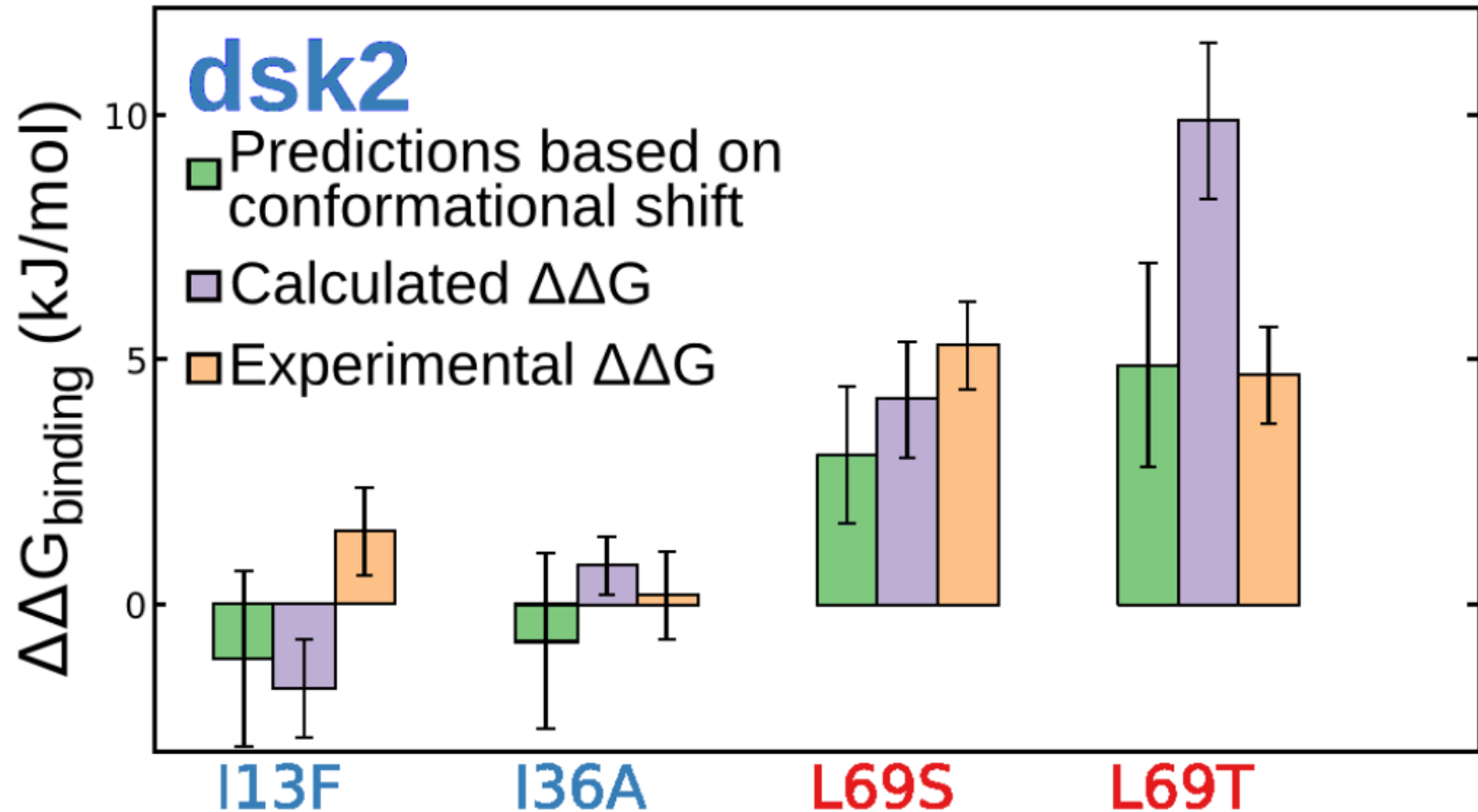
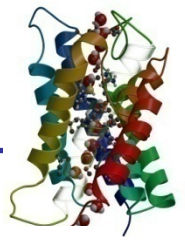
Complex formation of ubiquitin mutants



Complex formation of ubiquitin mutants



Complex formation of ubiquitin mutants



→ Conformational shift induces change in function

Team:

Water channels:

Guillem Portella
(Barcelona)

Shreyas Kaptan



Rodolfo Briones



Camilo Aponte
(Heidelberg)



Jochen Hub
(uni Göttingen)

Peter Pohl
Claudia Steinem
Ulf Diederichsen
Marina Benatti
Tim Salditt
Tom Walz



Collective dynamics:

Hadas Leonov Colin Smith Martin Vesper



Axel Munk
Robert Tampe
Christian Griesinger



Affinity prediction:

Daniel Seeliger
(Boehringer)

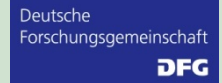


Sören Wacker

Joan Cerda
Sabine Flitsch
Eric Beitz
Peter Johnson
Michael Rützler

Vytautas Gapsys

Han Sun



Molecular recognition:



Jan Peters Dirk Matthes Servaas Michielssens

Oliver Lange
Christian Griesinger
Adam Lange



Physical & Chemical Graduate School Göttingen

Ion channels:

Anna Stary
(Vienna)



Ulrich Zachariae
(Dundee)



Björn Forsberg

David Köpfer



Chen Song
(Oxford)

Gert Vriend
Kornelius Zeth
Xention
Claudia Steinem
Markus Zweckstetter



Positions available!